



## Original Research Article

# Hydrogen Cyanide Production Ability by bacterial antagonist and their Antibiotics Inhibition Potential on *Macrophomina phaseolina* (Tassi.) Goid

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## ABSTRACT

This research was undertaken for the purpose of isolation and purification of indigenous *Pseudomonas* spp and *Bacillus* spp. and evaluating its ability in hydrogen cyanide synthesis and also evaluating the potential of super-strains on reduction in the fungal growth of *Macrophomina phaseolina*. According to this, the research was carried out in laboratory tests. 2 strains (obtained from rhizosphere soil from 2 isolates of sunflower plants) *B. subtilis* and 2 strains of *P. fluorescence* were sub-cultured, purified and refreshed. Then these strains were evaluated for the capability in cyanide synthesis by a quantitative and qualitative method. Among these isolates on qualitative analysis of HCN indicated a strong production of HCN in *Pf*<sub>1</sub> and *CPf*<sub>5</sub> was estimated. Isolates of *B. subtilis* did not produce HCN. In quantitative estimation, *Pf*<sub>1</sub> and *CPf*<sub>5</sub> recorded the maximum OD value of 0.094 and 0.085 respectively but *Bs*<sub>10</sub> recorded least OD value (0.015). The antibiotics produced by all the four effective bacterial isolates were effective against *M. phaseolina* and recorded reduction in growth of the pathogen ranged between 61.13 to 69.62 per cent reduction over control. Antibiotics produced by *Pf*<sub>1</sub> were maximum per cent reduction in the fungal growth (69.92%) followed by *CPf*<sub>5</sub> (65.59%). The *Bacillus* isolates *Bs*<sub>10</sub> and *CBS*<sub>4</sub> were efficacy against the pathogen with 63.36 per cent and 61.13 per cent reduction over control. All the isolates were on par in inhibition of mycelial growth.

## Keywords

Bacterial antagonist, HCN, Antibiotic, *M. phaseolina*

## Introduction

Sunflower (*Helianthus annus*, L.) is an important oil seed crop in India. It is one of the fastest growing plants which belong to family Asteraceae (Compositae) (Rodriguez *et al.*, 2002). *M. phaseolina* is a soil-borne fungus that causes charcoal rot disease of many crops in arid and

semiarid areas of the world (Dhingra and Sinclair, 1978). On sunflower is subjected to be attacked with a variety of fungal pathogens, which affect its yield and its oil quality (Sangawan *et al.*, 2005). Very high incidence and spreading of charcoal rot on sunflower was recorded in Slovakia

(Bokor, 2007) and in the Czech Republic in 2007 (Veverka *et al.*, 2008). *M. phaseolina* is an anamorphic and soil borne fungus with a broad host range that includes 75 plant families and more than 500 species worldwide (Salik, 2007; Khan, 2007). Charcoal rot is of great economic importance in arid areas of the world. It causes decrease in stem height, girth, root region and head weight (Raut, 1983; Kolte, 1985). Production of HCN by certain strains of *Pseudomonas fluorescens* has been involved in suppression of soil borne pathogens (Voisard *et al.*, 1989). HCN is produced by many rhizobacteria and is postulated to play a role in biological control of pathogens (Defago *et al.*, 1990). Hajji *et al.* (1989) reported that broad-spectrum of antibiotics such as gliotoxin known to have antimicrobial activity (Howell *et al.*, 1993; Sivasithamparam and Ghisalberti, 1998; Pozo *et al.*, 2004). Antibiotics are low molecular weight secondary metabolites produced by antagonists and directly inhibited the pathogen growth (Sabitha *et al.*, 2001). This objective aimed to investigate the qualitative and quantitative capability of rhizobacteria isolates in HCN production and on reduction in the fungal growth of *M. phaseolina*.

## Materials and Methods

### Isolation of antagonists from the rhizosphere region of sunflower plants

Antagonistic of bacteria were isolated from the rhizosphere soil collected from different sunflower growing areas of Tamil Nadu. The plants were pulled out gently with intact roots and the excess soil adhering on roots was removed gently. Ten gram of rhizosphere soil was

transferred to 250 ml Erlenmeyer flask containing 100 ml of sterile distilled water. After thorough shaking, the antagonist in the suspension was isolated by serial dilution plate method (Pramer and Schmidt, 1956).

From the final dilutions of  $10^{-5}$  and  $10^{-6}$ , one ml of each aliquot was pipetted out, poured in sterilized Petri dish containing King's B medium and nutrient agar medium separately and they were gently rotated clockwise and anti clockwise for uniform distribution and incubated at room temperature ( $28 \pm 2^\circ \text{C}$ ) for 24 hours. The colonies were viewed under UV light at 366 nm. Colonies with characteristics of *Bacillus* spp., *Pseudomonas* spp. were isolated individually and purified by streak plate method (Rangaswami, 1993) on Nutrient agar medium and King's B medium respectively. The pure cultures were maintained on respective agar slants at  $4^\circ \text{C}$ .

### Hydrogen cyanide (HCN) production Qualitative assay

HCN production of fungal and bacterial biocontrol agents was tested qualitatively following the method of Bakker and Schipper (1987). The antagonistic bacteria were streaked on King's B medium amended with glycine at 4.4g/ l. sterile filter paper saturated with picric acid solution (2.5 g of picric acid; 12.5 g of  $\text{Na}_2\text{CO}_3$ , 1000 ml of distilled water) was placed in the upper lid of the Petri plate. The dishes were sealed with Parafilm and incubated at  $28^\circ \text{C}$  for 48 h. A change of colour of the filter paper from yellow to light brown, brown or reddish-brown was recorded as weak (+), moderate (++) or strong (+++) reaction respectively.

### **Quantitative assay**

Antagonistic bacteria were grown in King's B broth amended with glycine (4.4g/ l) and Uniform strips of filter paper (10 x 0.5 cm<sup>2</sup>) were soaked in alkaline picrate solution and kept hanging inside the conical flask. After incubation at 28 ± 2°C for 48 h the sodium picrate in the filter paper was reduced to a reddish compound in proportion to the amount of HCN evolved. The colour was eluted by placing the filter paper in a test tube containing 10 ml of distilled water and its absorbance was read at 625 nm (Sadasivam and Manickam, 1992).

### **Antibiotic production - bacterial antagonists**

#### **Extraction of crude antibiotic metabolites**

The bacterial biocontrol agents *viz.*, Bs<sub>1</sub>, Bs<sub>10</sub>, Pf<sub>1</sub> and Pf<sub>2</sub> grown for five days in pigment production broth and were centrifuged at 5000 rpm for 30 min. The supernatant was adjusted to pH 2.0 with concentrated HCl and extracted with equal volume of benzene. The benzene layer was evaporated in a water bath and the residue was resuspended in 0.1 N NaOH (Rosales *et al.*, 1995).

#### **Effect of bacterial antibiotics on the growth of *M. phaseolina***

The effect of antibiotics extracted from bacterial antagonists was tested against the growth of *M. phaseolina* by filter paper disc assay (Lam and Ng, 2001). Three sterile filter paper discs were placed on solidified PDA in Petri dishes. The crude antibiotic extracted was pipetted on to the filter paper @150 µl/disc. A five-mm-

mycelial disc of the fungus was placed at the centre of the plate and incubated at 28 ± 2°C. Filter paper without antibiotic served as control. Surface area of inhibition was measured by tracing the area of inhibition in a trace paper, plotting it on a graph sheet and comparing with control.

### **Statistical analysis**

The pot culture and laboratory experiments were conducted by following Completely Randomized Design (CRD). The field experiment was laid out in Randomized Block Design (RBD). The percentage values were transformed into "Arcsine" and "Square-root". The statistical analysis of the experiment was done by following the methods suggested by Gomez and Gomez (1984). Per cent values were transformed by arcsine or square root transformation.

## **Results and Discussion**

### **Biochemical characterization of *P. fluorescens* isolates**

The two effective isolates of *P. fluorescens* gave positive result to following test *viz.*, KOH test, producing fluorescent pigment, growth at 4°C, arginine dihydrolase and gelatin liquefaction. These isolates gave negative reaction to Gram's reaction, growth at 41°C and levan formation. (Table 1).

### **Biochemical characteristics of *B. subtilis* isolates**

All the two effective isolates of *B. subtilis* showed positive reaction to gram reaction,

**Table.1** Characterization of *P. fluorescens*

S. No.	Diagnostic tests	Pf <sub>1</sub>	CPf <sub>5</sub>
1.	Gram reaction	-	-
2.	KOH test	+	+
3.	Pigment production in King's B medium	+	+
4.	Growth at 4°C	+	+
5.	Growth at 45°C	-	-
6.	Arginine dihydrolase	+	+
7.	Gelatin liquefaction	+	+
8.	Levan formation	-	-

**Table.2** Characterization of *B. subtilis*

S. No.	Diagnostic tests	Bs10	CBs <sub>4</sub>
1.	Gram reaction	+	+
2.	KOH test	-	-
3.	Growth at 45°C	+	+
4.	Growth in 7% NaCl	+	+
5.	Citrate utilization	+	+
6.	Anaerobic growth	-	-
7.	Starch hydrolysis	+	+
8.	Catalase test	+	+

**Table.3** Production of HCN from bacterial antagonists

S. No.	Isolates	HCN	
		Qualitative	Quantitative (O.D. Value)
1.	Bs10	-	0.015
2.	CBs5	-	0.004
3.	Pf1	+++	0.094
4.	CPf5	+++	0.085

HCN production - negative, + weak, ++ moderate, +++ strong

growth in 45°C, growth in NaCl, citrate utilization, starch hydrolysis and catalase test these isolates gave negative reaction in KOH test and anaerobic growth (Table 2).

### Production of HCN

Study on qualitative analysis of HCN indicated a strong production of HCN in *Pf*<sub>1</sub> and *CPf*<sub>5</sub> was estimated. Isolates of *B. subtilis* did not produce HCN (Table 3). In quantitative estimation, *Pf*<sub>1</sub> and *CPf*<sub>5</sub> recorded the maximum OD value of 0.094 and 0.085 respectively but *Bs*<sub>10</sub> recorded least OD value (0.015). Role of HCN in disease suppression has been demonstrated by several scientists in various crops (Stutz *et al.*, 1986; Voisard *et al.*, 1989; efago *et al.*, 1990). HCN is the common secondary metabolite produced by rhizosphere *Pseudomonas* (Schippers, 1988). Meena *et al.* (2001) compared the HCN production of several strains of *P. fluorescens* and their efficacy in controlling root rot of groundnut caused by *M. phaseolina*. *Pseudomonas* releasing HCN were reported in the rhizosphere of tobacco in soils suppressive to *T. basicola*, causal agent of black root rot of tobacco (Ramette *et al.*, 2006).

### Effect of bacterial antibiotics on the growth of *M. phaseolina*

The antibiotics produced by all the four effective bacterial isolates were effective against *M. phaseolina* and recorded reduction in growth of the pathogen ranged between 61.13 to 69.62 per cent reduction over control (Fig.1). Antibiotics produced by *Pf*<sub>1</sub> were maximum percent reduction in the fungal growth (69.92%) followed by *CPf*<sub>5</sub> (65.59%). The *Bacillus* isolates *Bs*<sub>10</sub> and *CBS*<sub>4</sub> were next only to pseudomonads in their efficacy against the pathogen with 63.36 per cent and 61.13

per cent reduction over control. All the isolates were on par in inhibition of mycelial growth. Bainton *et al.* (2002) reported that the naturally occurring fluorescent *Pseudomonads* produced the antibiotic, 2-4 DAPG. *Bacillus* spp. produced different inhibitory agents which have been categorized in peptide derivative family (Stein, 2005; *et al.*, 2002). Bacilysoicin, a novel and broad spectrum phospholipid antibiotic was purified from *B. subtilis* strain 168 (Tamehiro *et al.*, 2002).

In conclusion, Cyanogenic rhizobacteria might have the potential of biological control of *M. phaseolina*. The effect of each strain was different due to species and method. Growth inhibition was the most in *Pf*<sub>1</sub>.

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### References

- Bainton, N.J., Michael Lynch, J., Naseby, D. and Alexander Way, J. 2002. Survival and ecological fitness of *Pseudomonas fluorescens* genetically engineered with dual biocontrol mechanisms. Can. J. Microbiol., 56(2):706-709.

- Bakker, A.W. and Schipper, B. 1987. Microbial cyanide production in the rhizosphere in relation to potato yield reduction and *Pseudomonas* spp. mediated plant growth stimulation. *Soil Biol. Biochem.*, 19: 451-457.
- Bokor, P. 2007. *Macrophomina phaseolina* causing a charcoal rot of sunflower through Slovakia. *Biologia. Bratislava*, 62: 136-138.
- Defago, G., Berling, C.H., Burger, U., Haas, D., Kahr, G., Keel, C., Voisard, C., Wirthner, P. and Wuthrich, B. 1990. Suppression of black root rot of tobacco and other root diseases by strains of *Pseudomonas fluorescens* potential applications and mechanisms. *In: Biological control of soil bore plant pathogens*, D. Hornby (Ed.). CAB International, Wallingford, Oxon, U.K. pp. 93-98.
- Defago, G., Berling, C.H., Burger, U., Haas, D., Kahr, G., Keel, C., Voisard, C., Wirthner, P. and Wuthrich, B. 1990. Suppression of black root rot of tobacco and other root diseases by strains of *Pseudomonas fluorescens* potential applications and mechanisms. *In: Biological control of soil bore plant pathogens*, D. Hornby (Ed.). CAB International, Wallingford, Oxon, U.K. pp.93-98.
- Dhingra, O.D. and Sinclair, J.B. 1978. Biology and pathology of *Macrophomina phaseolina*. *Universidade Federal de Vicosa, Brasil*, p.166.
- Gomez, K.A. and Gomez, A.A. 1984. *Statistical procedures for Agricultural Research*. John Wiley and Sons, New York, p.680.
- Hajji, E.L., Rebuffat, M.S., Le Doan, T., Klein, G., Satre, M. and Bodo, B. 1989. Interaction of trichorzianines A and B with model membranes and with *Amoeba dictyostelium*. *Biochem. Biophys. Acta.*, 57: 97- 104.
- Howell, C.R., Stipanovic, R.D. and Lumsden, R.D. 1993. Antibiotic production by Strains of *Gliocladium virens* and its relation to the biocontrol of cotton seedling disease. *Biocont. Sci. Tech.*, 3: 435-441.
- Kolte, S.J. 1985. Sunflower diseases of annual oilseed crops, Vol. III, CRC Press, Inc. Boca Raton, Florida, 33-44.
- Lam, S.K. and Ng, T.B. 2001. First simultaneous isolation of a ribosome inactivating protein and an antifungal protein from a mushroom (*Lyophyllum shimeji*) together with evidence for synergism of their antifungal effects. *Archi. Biochem. Biophysics.*, 393(2): 271-280.
- Meena, B., Marimuthu, T., Vidhyasekaran, P. and Velazhahan, R. 2001. Biological control of root rot of groundnut with antagonistic *Pseudomonas fluorescens* strains. *J. Pl. Dis. Protect.*, 108: 369-381.
- Pozo, M.J., Baek, J.M., Garcia, J.M. and Kenerley, C.M. 2004. Functional analysis of a serine protease-encoding gene in the biocontrol agent *Trichoderma virens*. *Fungal Genet. Biol.*, 41: 336-348.
- Pramer, D. and Schmid, E.L. 1956. *Experimental soil Microbiology*, Buffer Publ. Co., Minneapolis, USA. p.107.
- Ramette, A., Loy, M. and Defago, G. 2006. Genetic diversity and biocontrol protection of *fluorescens pseudomonas* producing phloroglucinols and hydrogen cyanide from swiss soils naturally suppressive or conducive to *Thieviopsis basicola* mediated black rot of tobacco. *FEMS Microbial Ecol.*, 55(3): 369-381.
- Rangaswami, G. 1993. *Diseases of crop plants in India*. Prentice Hall of India (Pvt). Ltd., New Delhi. p.498.

- Raut, J.G. 1983. Transmission of seed borne *Macrophomina phaseolina* in seed. *Sci. Technol.*, 11: 807-817.
- Rodriguez, J.D., de, J., Romero-Garcia, R., Rodriguez-Garcia J.L.A. and Sanchez. 2002. Characterization of proteins from sunflower leaves and seeds. *Relationship of biomass and seed yield. In: Janich J. and A. Whikey (Eds). Trends in New Crops and New Uses. ASHS press, Alexandria. pp: 143-149.*
- Rosales, A.M., Thomashow, L., Cook, R.J. and Mew, T.W. 1995. Isolation and identification of antifungal metabolites produced by rice associated antagonistic *Pseudomonas* sp. *Phytopathology*, 85: 1028-1032.
- Sabitha, D., Nakkeeran, S. and Chandrasekhar, G. 2001. *Trichoderma bioaresenel* in plant disease management and its scope for commercialization. IPS southern zone meeting, Calicut. pp. 43- 55.
- Sadasivam, S. and Manickam, A. 1992. *Biochemical Methods for Agricultural Sciences*. Wiley Eastern Ltd, New Delhi, p 246.
- Salik, N.K. 2007. *Macrophomina phaseolina* as causal agent for charcoal rot of sunflower. *Mycopath.*, 5(2): 111-118.
- Sangawan, M., Metha, N. and Saharan, G. 2005. *Diseases of Oil Seed Crops*. Indus Publication Co. India. P.11-15.
- Schippers, B. 1988. Biological control of pathogens with rhizobacteria. *Trans British Soc. Land. B. Biol. Sci.*, 318: 283-293.
- Sivasithamparam, K. and Ghisalberti, E.L. 1998. *Secondary metabolism in Trichoderma and Gliocladium. In: Kubicek, C.P. and G.E. Harman (Eds.) .Trichoderma and Gliocladium. Basic biology, Taxonomy & Genetics, 139-191.*