

Original Research Article

<https://doi.org/10.20546/ijcmas.2020.908.239>

## *In vitro* Evaluation and Sensitivity of Antibiotics and Fungicides against *Xanthomonas axonopodis* pv. *punicae*

Sumant H. Kabade\*, R. W. Ingle, Punam N. Usendi and Rahul S. Shete

Department of Plant Pathology, Dr. Panjabrao Deshmukh Krishi Vidyapeeth,  
Akola- 444104 (M.S.), India

\*Corresponding author

### ABSTRACT

#### Keywords

*Xanthomonas axonopodis* pv.,  
*Punica granatum*,  
Bacterium,  
Streptomycine sulphate,  
Copper oxychlorite,  
Inhibition zone,  
Streptocycline

#### Article Info

Accepted:  
20 July 2020  
Available Online:  
10 August 2020

Cultivation of high yielding varieties of pomegranate with intensive care and management in the recent past under irrigated condition with early stage exploitation of plants has lead to certain severe pest and disease problems. Bacterial blight disease of pomegranate caused by *Xanthomonas axonopodis* pv. *punicae* is one of the most destructive disease of pomegranate (*Punica granatum*) inflicting considerable quantitative and qualitative losses. Mostly the disease occurred on leaves, stems and fruits. Extensive agricultural practices along with large amount of pesticide as well as antibiotic are required to control this disease. The antibiotic sensitivity against eight isolates was studied by paper disk inhibition method. Streptomycine sulphate (250ppm) + COC (2500ppm) was significantly superior than all the treatments showing maximum inhibition zone in all eight isolates of *Xanthomonas axonopodis* pv. *punicae*.

### Introduction

Pomegranate (*Punica granatum* L.) is a favourite table fruit in tropical and sub-tropical regions of the world which belongs to family *Puniceae*. Among the major diseases leaf spot, fruit spot and will results in reduction of pomegranate fruit yield and put the growers in to hardship. Pomegranate grows very well on the moderately alkaline

soils as well as slightly acidic soils. Bacterial blight infection results in appearance of water soaked oily spot symptoms on leaves, stems and fruits which consequently decreases fruit production and market value. The continuous presence of the pathogen in the orchard throughout the season has led to many fold speculations on possible survival of the pathogen on some alternate hosts grown in and around the garden. Severity of incidence

and losses varies among different isolates and influenced by existing climatic conditions and geographical distribution (Mondal and Sharma, 2009; Mondal and Singh, 2009; Petersen *et al.*, 2007; Mondal *et al.*, 2012).

## **Materials and Methods**

### **Collection of diseased sample and isolation of pathogen**

The bacterial blight diseased samples were collected from different districts of Maharashtra state. The isolate of pathogen was obtained from infected leaves of Pomegranate showing typical symptoms of bacterial blight by tissue isolation method. A bacterial suspension of each specimen was then cultured on Nutrient Sucrose Agar (NSA) medium. Following incubation, colonies similar to *Xanthomonas* were maintained on NSA medium at room temperature by adopting subsequent subculturing at periodical and regular intervals. Three days old cultures were used for further studies.

### **Pathogenicity test of the isolate**

For this study the healthy seedling of Pomegranate cultivar 'Bhagwa' was obtained from the central nursery of state agriculture university P.D.K.V. Akola, Dist. - Akola, Maharashtra state. The seedling was grown under aseptic in vitro condition and veinlets of some leaves were injected with 48 hold bacterial suspension with the help of sterile syringe. Some plant leaves were injured by sharp needle and inoculated by spraying bacterial suspension. The inoculated plants were covered with polythene sheets and incubated for 10-12 days at 25 to 28°C temperature. After 10– 12 days observations made for typical symptoms of bacterial blight on leaves, the organism was reisolated from artificially inoculated leaves and used for further antibacterial studies.

### **In vitro evaluation and sensitivity test of antibiotics & fungicides against *Xanthomonas axonopodi* spv. punice by Paper disc method**

Different chemicals at concentration 100, 300, 500 ppm were evaluated in vitro applying inhibition zone technique (paper disc method) and using Nutrient Agar (N.A.) as basal culture medium. Fresh Nutrient Agar medium was prepared and dispersed in 100 ml quantities in conical flask (200 ml. Cap.), plugged and autoclaved at 15 lbs/cm<sup>2</sup> pressure for 15-20 minute. The desired concentration of chemicals i.e. 100, 300, 500 ppm was prepared by using appropriate quantities of chemical required for 100, 300, 500 ppm concentration. In this desire concentration of chemicals 5 mm discs of Whatman No. 1 filter paper was dipped for few minutes. After sterilization of media, it was allowed to cool down to 35°C before pouring. Approximately 20 ml liquid media was poured in previously sterilized Petri plates and allowed them to solidify. Pouring of plates were always be done by using Laminar Air Flow cabinet under aseptic condition. After solidification of media of Petri plates, the bacterial suspension was spread on Nutrient Agar with glass spreader. After uniform spreading of bacterial suspension 5 mm disc of Whatman No.1 filter paper previously dipped in desired concentration of chemicals was placed in centre of medium. Three replication for each concentration of chemical was maintained. The paper disc soaked in sterile distilled water served as control Three replication for each concentration of chemical was maintained. The paper disc soaked in sterile distilled water served as control.

All these Petri plates after treatment were incubated at 28 ± 2°C for 48 hours. Observation on radial growth of test pathogen.

## Result and Discussion

### Isolation and pathogenicity test

Plates showing well separated, typical, yellow, mucoid, colonies of *Xanthomonas* bacterium were used to check pathogenicity on leaves of Pomegranate to confirm the isolate. After inoculation of pathogen in seedlings, seedlings were covered with plastic bags for maintain humidity for 3-4 days. The symptoms of the disease were developed within 10 to 18 days after inoculation on leaves small, water soaked, brown to black coloured lesions, which later on developed into angular to irregular shaped spots along the veins and veinlets of the leaf lamina. The re-isolation attempted from artificially infected / diseased plant tissues on Nutrient Agar consistently yielded *X. axonopodis* pv. *punicae*, thus fulfilling Koch's postulates and association of *X. axonopodis* pv. *punicae* with pomegranate was confirmed.

### Pathogenic variability among different isolates of *Xanthomonas axonopodi* sp. *punicae*

Amongst eight isolates Xap-3 isolate had more lesion size i.e. 3.5 mm after 18 days of inoculation which showed water soaked circular to irregular dark brown spots with yellow halo symptoms followed by Xap-7 which showed 1.5 mm lesion size after 18 days after inoculation showing water soaked circular to irregular, light brown spots. The data is Presented in Table 1.

### *In vitro* evaluation and sensitivity test of antibiotics & fungicides against *Xanthomonas axonopodis* sp. *punicae* isolates

The sensitivity of antibiotics and fungicides was tested against eight isolates of *Xanthomonas axonopodis* pv. *punicae* (Xap-1 to Xap-8) by Paper disc inhibition method and

data is Presented in Table 2 and Fig. 2. The streptomycin used in different concentration viz; 100, 250, 500 ppm. The results (Table 2) revealed that lower concentration of streptomycin i.e. 100 ppm showed maximum growth inhibition in Xap-2 i.e. 15.67 mm whereas the minimum growth inhibition was observed in Xap-1 i.e. 6.00 mm. For the concentration 250 ppm, maximum inhibition was seen in Xap-8 (20.67mm) and minimum inhibition in Xap-7 (11.67mm). The higher concentration of streptomycin i.e. 500 ppm reported maximum inhibition in Xap-2 (23.67mm) and minimum inhibition in Xap-1 (18.67mm). The results showed that COC and Salicylic acid did not inhibit the growth which shows that isolates are not sensitive to COC and Salicylic acid. The results (Table 2, Fig 2) revealed that among the isolates Xap-6 (39mm) showed maximum growth inhibition whereas minimum growth inhibition was shown by xap-8 (28.67mm). Streptomycin 250ppm + COC 2500 ppm showed maximum growth inhibition in Xap-4 (24.67) and minimum inhibition in Xap-2 (21.67mm). The findings (Table 2, Fig. 2) revealed that Streptomycin sulphate 250ppm + COC 2500ppm (T8) was significantly superior over rest of the treatments showing maximum growth inhibition zone (39mm) in Xap-6, followed by Xap-2 (34.67mm). Whereas growth inhibition zone was minimum in Xap-8 (28.67mm).

Similar results observed by Abhang *et al.*, (2015). The efficacy of bioagents, botanicals and chemicals was studied by paper disc method. The Copper oxychloride (0.2%) + streptomycin sulphate (200 ppm) was found significantly effective in inhibiting growth of *Xanthomonas axonopodis* pv. *citri*. Raju *et al.*, (2012) carried out investigation to screen the different bactericides against *Xanthomonas axonopodis* pv. *punicae*. The inhibition zone of 3.3 cm was observed in the treatment of Streptomycin + Copper oxychloride.

Ambadkar *et al.*, (2015) studied in vitro efficacy of different antibiotics for management of bacterial blight and disease of pomegranate caused by *Xanthomonas axonopodis* pv. *punicae*. They found that antibiotic streptomycin showed maximum inhibition zone of 22.21 and 31.00 per cent at 250 and 500 ppm against *Xanthomonas axonopodis* pv. *punicae* followed by

tetracycline (18.26 and 27.53 %) and bacterinol (17.40 and 27.15 %). Abhang *et al.*, (2015), the efficacy of bioagents, botanicals and chemicals was studied by paper disc method. The Copper oxychloride (0.2%) + streptomycin sulphate (200 ppm) was found significantly effective in inhibiting growth of bacteria.

**Table.1** List of different isolates collected from various villages of Maharashtra state

Sr. no	Name of isolate	Name of Village	Taluka	District	Pathogen	Affected plant part
1	Xap-1	Mardi	Man	Satara	<i>Xanthomonas axonopodis</i> pv. <i>punicae</i>	Leaf
2	Xap-2	Anjangaon	Mhada	Solapur	<i>Xanthomonas axonopodis</i> pv. <i>punicae</i>	Leaf
3	Xap-3	Agoti -1	Indapur	Pune	<i>Xanthomonas axonopodis</i> pv. <i>punicae</i>	Leaf
4	Xap-4	Bherdapur	Newasa	Ahmednagar	<i>Xanthomonas axonopodis</i> pv. <i>punicae</i>	Fruit
5	Xap-5	Satephal	Jafrabad	Jalna	<i>Xanthomonas axonopodis</i> pv. <i>punicae</i>	Leaf
6	Xap-6	Kolara	Chikhali	Buldhana	<i>Xanthomonas axonopodis</i> pv. <i>punicae</i>	Leaf
7	Xap-7	Ekamba	Malegaon	Washim	<i>Xanthomonas axonopodis</i> pv. <i>punicae</i>	Leaf
8	Xap-8	Akola	Akola	Akola	<i>Xanthomonas axonopodis</i> pv. <i>punicae</i>	Leaf

Sr. No	Treatment No	Name of chemicals	Concentration of chemicals (ppm)
1	T1	Streptomycin sulphate	100
2	T2	Streptomycin sulphate	250
3	T3	Streptomycin sulphate	500
4	T4	Streptomycin	100
5	T5	Streptomycin	250
6	T6	Streptomycin	500
7	T7	Copper Oxychlorite	2500
8	T8	Streptomycin sulphate + Copper oxychlorite	250 + 2500
9	T9	Streptomycin + Copper oxychlorite	250 + 2500
10	T10	Salicylic acid	200
11	Control		

**Table.2** Pathogenic variability among different isolates of *Xanthomonas axonopodis* pv. *punicae* on Pomegranate 45-60 day old seedlings

Sr. No	Isolates	Lesion size (mm)
		After 18 days
1	Xap1	3
2	Xap2	3
3	Xap3	3.5
4	Xap4	3
5	Xap5	2
6	Xap6	2
7	Xap7	1.5
8	Xap8	2

**Table.3** *In vitro* evaluation and sensitivity test of antibiotics & fungicides against *Xanthomonas axonopodis* pv. *punice* isolates

Sr No	Name of treatment	Concentration (ppm)	Zone of inhibition (mm) * average of three replication							
			Xap1 (Satara)	Xap2 (Solapur)	Xap3 (Pune)	Xap4 (Ahmednagar)	Xap5 (Jalna)	Xap6 (Buldhana)	Xap7 (Washim)	Xap8 (Akola)
T1	Streptomycin sulphate	100	0.00	0.00	0.00	0.00	0.00	15.50	0.00	0.00
T2	Streptomycin sulphate	250	0.00	0.00	0.00	0.00	0.00	16.67	0.00	0.00
T3	Streptomycin sulphate	500	0.00	0.00	0.00	0.00	0.00	19.17	0.00	0.00
T4	Streptocycline	100	6.00	15.67	0.00	0.00	0.00	12.67	0.00	15.00
T5	Streptocycline	250	15.67	20.33	15.33	18.00	16.33	18.33	11.67	20.67
T6	Streptocycline	500	18.67	23.67	23.00	20.67	22.00	23.67	21.67	22.67
T7	Copper Oxchlorite	2500	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
T8	Streptomycin sulphate + COC	250+2500	31.33	34.67	31.00	34.00	33.00	39.00	30.67	28.67
T9	Streptocycline + COC	250+2500	24.00	22.33	21.67	24.67	24.00	24.00	23.00	23.33
T10	Salicyclic acid	200	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
T11	Control		0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
F test			<b>Sig.</b>	<b>Sig.</b>	<b>Sig.</b>	<b>Sig.</b>	<b>Sig.</b>	<b>Sig.</b>	<b>Sig.</b>	<b>Sig.</b>
SE(M) ±			<b>0.95</b>	<b>0.97</b>	<b>0.57</b>	<b>0.68</b>	<b>0.37</b>	<b>0.66</b>	<b>0.64</b>	<b>0.31</b>
CD (P=0.1)			<b>3.80</b>	<b>3.88</b>	<b>2.29</b>	<b>2.74</b>	<b>1.48</b>	<b>2.63</b>	<b>2.57</b>	<b>1.26</b>

Fig.1 Pathogenicity test



Plate 8 Pathogenicity test of *Xanthomonas axonopodis* pv. *punicae* on Pomegranate seedlings

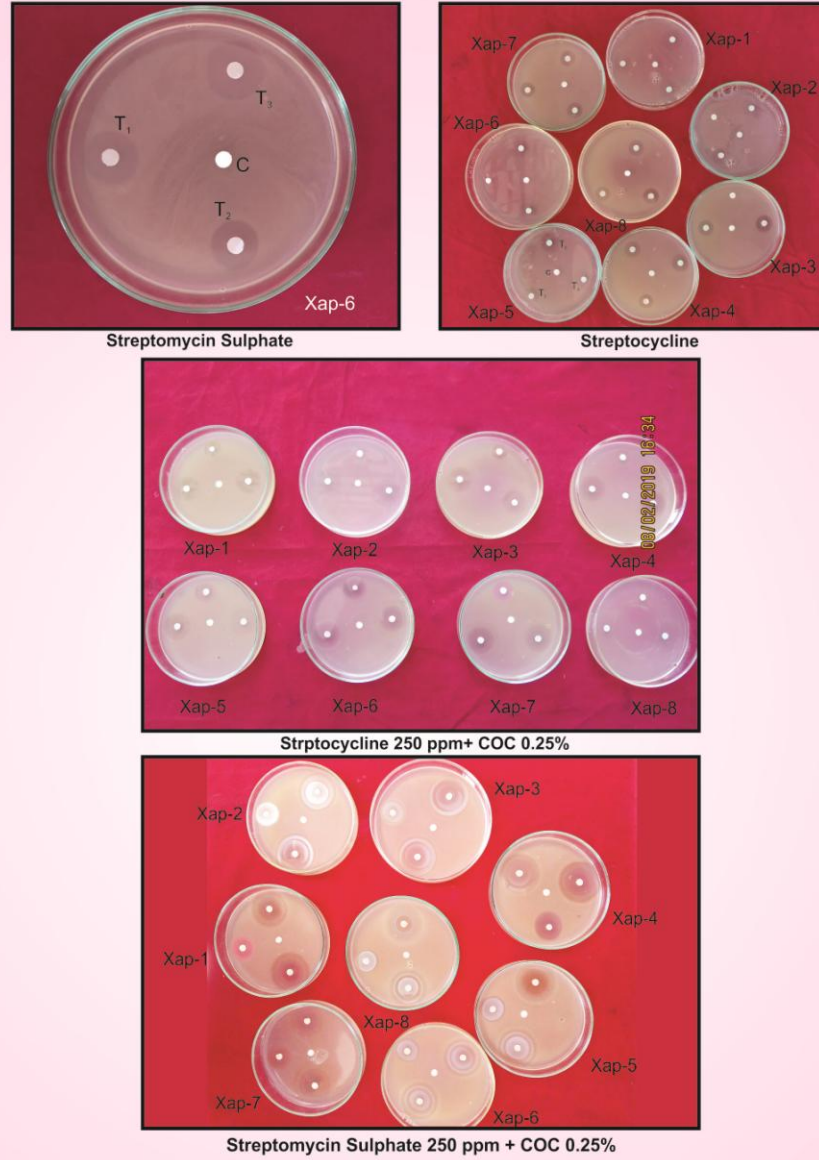


Plate 9 Pathogenicity test of *Xanthomonas axonopodis* pv. *punicae* on Fruit of pomegranate



Plate 10 Re-isolation of *Xanthomonas axonopodis* pv. *punicae*

**Fig.2** *In vitro* evaluation and sensitivity test of antibiotics & fungicides against *Xanthomonas axonopodis* pv. *punice* isolates



- |  |   |
|--|---|
| T <sub>1</sub> - Streptomycin Sulphate 100 ppm   | T <sub>2</sub> -Streptomycin Sulphate 250 ppm             |
| T <sub>3</sub> -Streptomycin Sulphate 500 ppm    | T <sub>4</sub> - Strptocycline100 ppm                     |
| T <sub>5</sub> -Strptocycline 250 ppm            | T <sub>6</sub> -Strptocycline500 ppm                      |
| T <sub>7</sub> - COC 0.25%                       | T <sub>8</sub> -Streptomycin Sulphate 250 ppm + COC 0.25% |
| T <sub>9</sub> -Strptocycline 250 ppm+ COC 0.25% | T <sub>10</sub> - Salicyclic acid 200 ppm                 |
| T <sub>11</sub> -Control                         |   |

**Plate 16** *In vitro* Evaluation and sensitivity test of antibiotics and fungicides against *Xanthomonas axonopodis* pv. *puniceae*



## References

- Petersen, Y., E. L. Mansvelt, E. Venter, and W. E. Langenhoven 2007. Detection of *Xanthomonas axonopodispv. Punicae* causing bacterial blight on pomegranate in South Africa. *J. of Austrian Pl. Path.*, 39: 544-546.
- Mondal, K. K. and D. Singh 2009. Bacterial blight of pomegranate-A technical bulletin of Division of Plant Pathology, Indian Agricultural Research Institute, New Delhi, pp: 8.
- Mondal, K. K. and J. Sharma 2009. Bacterial blight an emerging threat to pomegranate export of Indian Farming, 59: 22-23.
- Mondal, K. and C. Mani, 2012. Investigation of the antibacterial properties of nanocopper against *Xanthomonas axonopodispv. punicae*, the incitant of pomegranate bacterial blight. *Annals of Microbiology*, 62: 889-893.
- Abhang, P. B., M. V. Totawar, and R. Kadam, 2015. Biochemical characterization of *Xanthomonas axonopodispv. citri* for identification of Citrus canker disease. *J. Basic Sci.*, 30-33.
- Raju, J., B. Enagi, V. I. Jaylakshmi, , K. Angadi, S. G. Basha and P.S. Sonavane, 2012. Survey, surveillance and *in vitro* evaluation of chemicals against *Xanthomonas axonopodispv. punicae* causing bacterial blight of pomegranate. *J. Pl. Dis. Sci.* 7(2): 225-230
- Ambadkar C.V., A. S. Dhawan and V.N. Shinde 2015. Integrated management of bacterial blight disease (oily spot) of pomegranate caused by *Xanthomonas axonopodispv. punicae*. *Int. J. Pl. Dis. Sci.*, 10(1) : 19-23.
- Ambadkar, C. V., M. A. Atarand G. A. Bhalariao, 2015. Occurrence and distribution of bacterial blight of pomegranate caused by *Xanthomonas axonopodispv. punicae* in Latur and Osmanabad districts of Marathwada region. *Adv. Res. J. crop Improvement* 6(1): 50-55.

### How to cite this article:

Sumant H. Kabade, R. W. Ingle, Punam N. Usendi and Rahul S. Shete. 2020. *In vitro* Evaluation and Sensitivity of Antibiotics and Fungicides against *Xanthomonas axonopodis pv. punicae*. *Int.J.Curr.Microbiol.App.Sci.* 9(08): 2101-2109.  
doi: <https://doi.org/10.20546/ijcmas.2020.908.239>