

Original Research Article

<https://doi.org/10.20546/ijcmas.2020.910.443>

Studies on Anti-fungal Activity of Leaf Extracts of *Andrographis echiooides* (L.) Nees

P. Hemalatha*, V. Sivakumar, R. Parimaladevi and M. Tilak

Tamil Nadu Agricultural University, Coimbatore, India

*Corresponding author

ABSTRACT

Keywords

Andrographis echiooides, Anti-fungal activity, Leaf extract, Inhibition zone

Article Info

Accepted:
25 September 2020
Available Online:
10 October 2020

Among all the species of *Andrographis*, *Andrographis echiooides* is given importance recently for its excellent medicinal properties. The study of *Andrographis echiooides* as anti-microbial agent was found necessary for gaining insight into this medicinal flora and its real value. The ethyl acetate extract of leaves recorded highest anti-fungal activity against all the selected pathogens viz., *Pythium aphanidermatum*, *Phytophthora capsici*, *Macrophomina phaseolina*, *Fusarium udum* and *Aspergillus niger* and produced inhibition zone of diameter equal to the positive control. *Macrophomina phaseolina* and *Aspergillus niger* were found to be more sensitive to ethyl acetate extract with a MIC value of 100 ppm.

Introduction

Andrographis echiooides (L.) Nees (Gopuram thanki) is one of the important medicinal plant species belonging to the family Acanthaceae. *Justicia echiooides* L. and *Indoneesiella echiooides* (L.) Sreemadh are the synonyms of this plant. The plant is an erect, annual herb, simple or slightly branched, growing up to a height of 20 to 60 cm. In the Indian Systems of Medicine predominantly, it is used against blood cancer. The leaf extract is recommended for oral consumption. Traditionally, the plant has been used as febrifuge, bitter tonic, astringent, anodyne and also for dysentery, cholera and diabetes. The

chemical constituents of this plant are echioidin and echioidin (Guhabakshi *et al.*, 1999).

Antibiotics are defined as substances (secondary metabolites) of microbial origin that have anti-microbial activity in very small amounts. It is reported that, on average, two or three antibiotics derived from microbes are launched each year. But the lifespan of such antibiotics is limited. Further, the awareness of problems due to over prescription and misuse of traditional antibiotics is increasing. This has resulted in increasing interest in the anti-microbial plant extracts (Eisenberg *et al.*, 1993).

Plants have an almost limitless ability to synthesize substances, most of which are phenols or their oxygen-substituted derivatives (Geissman, 1963). Most are secondary metabolites, of which at least 12,000 have been isolated, a number estimated to be less than 10 per cent of the total (Schultes, 1978). In many cases, these substances serve as plant defense mechanisms against predation by microorganisms, insects and herbivores. Since the past research work on antimicrobial aspect is very limited, this study was carried out to determine specifically the antifungal activity of *Andrographis echinoides* (L.) Nees.

Materials and Methods

The present investigation on photochemical screening of *Andrographis echinoides* (L.) Nees was carried out in Department of Microbiology, Tamil Nadu Agricultural University, Coimbatore.

Planting material

Leaves of *Andrographis echinoides* (L.) Nees. were collected from Medicinal Plants Unit of Horticultural College and Research Institute, Tamil Nadu Agricultural University, Coimbatore. The leaves were shade dried at room temperature for four to five days.

Extract preparation

Air dried and powdered plant materials (0.25 to 0.5 kg) were extracted by maceration and percolation with 70 per cent ethanol, 80 per cent methanol, hexane, dichloromethane, petroleum ether, ethyl acetate, chloroform and water at room temperature. The extracts were then filtered and concentrated under vacuum in rotary evaporator to give (as a percentage of powdered plant materials) 6-11 per cent gummy residue. All the extracts were kept in tightly stoppered bottle in a refrigerator until

used for the anti-microbial testing.

Fungi and media

The plant pathogenic fungal cultures used in this study viz., *Pythium aphanidermatum*, *Phytophthora capsici*, *Macrophomina phaseolina*, *Fusarium udum* and *Aspergillus niger* were obtained from Department of Plant Pathology, Tamil Nadu Agricultural University, Coimbatore. Fungi were cultured and maintained on potato dextrose agar at room temperature.

Agar well diffusion method

The anti-fungal activity of leaf extracts of *Andrographis echinoides* (L.) Nees. was determined by modified agar well diffusion method of Perez *et al.*, (1990). Once the agar was solidified, it was punched with six millimeters diameter wells and filled with 25 μ l of the plant extracts and blanks (70 % ethanol, 80 % methanol, hexane, chloroform, dichloromethane, ethyl acetate, acetone and water).

The concentration of the extracts employed was 25 μ gml⁻¹. Simultaneously, ketoconazole was used as positive control at a concentration of 1.0 μ gml⁻¹ respectively. The dilution medium for the positive control was sterile distilled water. The test was carried out in triplicates. The plaques were incubated at room temperature for 48 hours. The anti-microbial activity was calculated by applying the expression.

$$\text{RIZD (\%)} = \frac{\text{IZD of sample} - \text{IZD of negative control}}{\text{IZD of antibiotic standard}} \times 100$$

Where,

RIZD is the percentage of relative inhibition zone diameter

IZD is the inhibition zone diameter (mm).

Minimal inhibitory concentration (MIC) evaluation

The MIC was evaluated on plant extracts that showed anti-microbial activity. This was performed at four concentrations of each extract (1 ppm, 10 ppm, 100 ppm and 1000 ppm) employing the same modified agar well diffusion method.

Results and Discussion

Anti-fungal activity of different solvent extracts

The treatment T₁ (70% ethanol), T₂ (Methanol), T₃ (Acetone) and T₆ (Ethyl acetate) recorded anti-fungal activity against the selected fungal plant pathogens (Table 1). The ethanol (T₁) and methanol (T₂) extracts recorded activity in particular against *Fusarium udum* and *Aspergillus niger* whereas acetone extract (T₃) recorded the activity against *Macrophomina phaseolina* and *Fusarium udum*. The ethyl acetate extract (T₆) recorded the highest activity against all

the fungus plant pathogens viz., *Pythium aphanidermatum*, *Phytophthora capsici*, *Macrophomina phaseolina*, *Fusarium udum* and *Aspergillus niger*. The positive control (Ketaconazole) recorded activity against all test organisms, while negative control (respective solvents) did not express any activity.

In accordance with these results, it was clear that andrographolides, which were present in the ethyl acetate fraction, could be considered responsible for the anti-microbial activity. Atta-ur-Rahman and Choudhary (1995) stated that diterpenoid alkaloids are commonly found to have anti-microbial properties. Similarly, Ghosh *et al.*, (2004) reported that the crude protein extract from the leaves of *Andrographis paniculata* was found to inhibit the spore germination of two major pathogens *Aspergillus flavus* and *Macrophomina phaseolina*. The anti-fungal protein component was further purified from the crude extract and the molecular mass of toxic protein was estimated to be 39.5 KDa.

Table.1 Anti-fungal activity of different solvent extracts of *Andrographis echioides* (L.) Nees leaves

Test samples	Test organisms				
	<i>Pythium aphanidermatum</i>	<i>Phytophthora capsici</i>	<i>Macrophomina phaseolina</i>	<i>Fusarium udum</i>	<i>Aspergillus niger</i>
T ₁ – 70% ethanol	-	-	-	+	+
T ₂ – Methanol	-	-	-	+	+
T ₃ – Acetone	-	-	+	+	
T ₄ – Hexane	-	-	-	-	-
T ₅ – Petroleum ether	-	-	-	-	-
T ₆ – Ethyl acetate	+	+	+	+	+
T ₇ – Dichloromethane	-	-	-	-	-
T ₈ – Aqueous extract	-	-	-	-	-
T ₉ – Positive control	+	+	+	+	+
T ₁₀ – Negative control	-	-	-	-	-
-: No activity	+: Activity recorded		Positive control: Ketaconazole		Negative control: Respective solvents

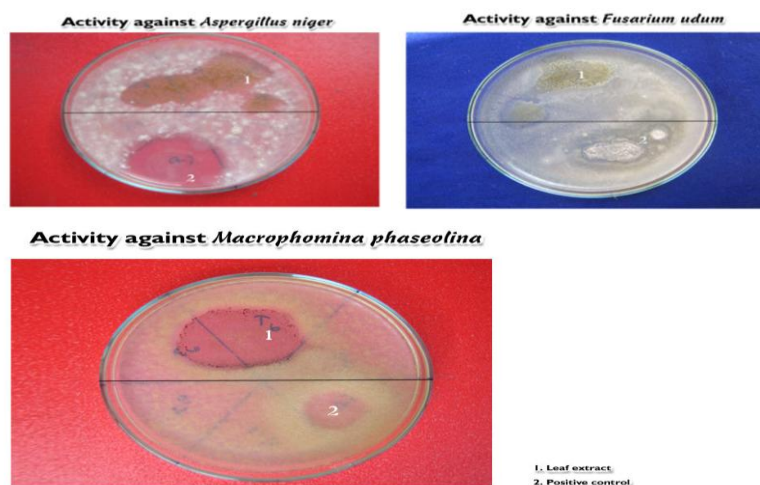
Table.2 Determination of inhibition zone of different solvent extracts of *Andrographis echiioides* (L.) Nees leaves

Test samples	Zone of inhibition (cm)				
	Test organisms				
	<i>Pythium apanidermatum</i>	<i>Phytophthora capsici</i>	<i>Macrophomina phaseolina</i>	<i>Fusarium udum</i>	<i>Aspergillus niger</i>
T ₁ - 70% ethanol	NT	NT	NT	1.9, 2.0	2.2, 2.3
T ₂ - Methanol	NT	NT	NT	2.2, 2.3	2.7, 2.6
T ₃ - Acetone	NT	NT	2.4, 2.1	2.5, 2.6	NT
T ₄ - Ethyl acetate	2.8, 2.8	2.5, 2.7	3.5, 3.6	3.2, 3.3	3.3, 3.4
T ₅ - Positive control	2.9, 3.0	2.6, 2.7	3.4, 3.5	3.3, 3.5	3.2, 3.3
T ₆ - Negative control	-	-	-	-	-
NT: Not tested	-: No activity	Positive control: Ketaconazole		Negative control: Respective solvents	

Table.3 Determination of Minimum Inhibitory Concentration (MIC) for ethyl acetate extract of *Andrographis echiioides* (L.) Nees leaves and Ketaconazole

Organisms	Dilution 1		Dilution 2		Dilution 3		Dilution 4	
	Ethyl acetate extract	Ketaco-nazole	Ethyl acetate extract	Ketaco-nazole	Ethyl acetate extract	Ketaco-nazole	Ethyl acetate extract	Ketaco-nazole
<i>Pythium apanidermatum</i>	NG	NG	G	NG	G	G	G	G
<i>Phytophthora capsici</i>	NG	NG	G	NG	G	G	G	G
<i>Macrophomina phaseolina</i>	NG	NG	NG	G	G	G	G	G
<i>Fusarium udum</i>	NG	NG	G	NG	G	G	G	G
<i>Aspergillus niger</i>	NG	NG	NG	NG	G	G	G	G
Dilution 1: 1000 ppm	Dilution 2: 100 ppm		Dilution 3: 10 ppm		Dilution 4: 1 ppm.		NG: No growth	G: Growth

Plate.1 Antifungal activity of *Andrographis echiioides* (L.) Nees



The hexane, petroleum ether, dichloromethane and aqueous extracts did not produce any activity. This might have resulted from the lack of solubility of the active constituents in these solutions (Romero *et al.*, 2005).

Inhibition zone of different solvent extracts

The inhibition zone of different solvent extracts of *Andrographis echinoides* leaves was determined by agar well diffusion method. The diameter of inhibition zone of ethanol extract (T₁) against *Fusarium udum* and *Aspergillus niger* was found to be 1.9, 2.0 cm and 2.2, 2.3 cm respectively. The methanol extract (T₂) produced the inhibition zone of about 2.2, 2.3 cm and 2.7, 2.6 cm diameter against *Fusarium udum* and *Aspergillus niger*. The ethyl acetate extract (T₄) produced inhibition zone of diameter equal to the positive control against all the selected fungal pathogens (Table 2). The negative control did not produce any inhibition zone against the fungal pathogens.

Determination of Minimum Inhibitory Concentration (MIC)

Four different dilutions of ethyl acetate extract and Ketaconazole *viz.*, 1000 ppm, 100 ppm, 10 ppm and 1 ppm were tested to determine the minimum inhibitory concentration. It was evident that both the ethyl acetate extract and Ketaconazole inhibited the growth of all the fungal pathogens in the dilution one (1000 ppm). In the dilution 2 (100 ppm), the Ketaconazole exhibited activity against all the pathogens except *Macrophomina phaseolina* (Table 3) whereas the ethyl acetate extract exhibited activity against *Macrophomina phaseolina* and *Aspergillus niger* (Table 3 and Plate 1). No activity was recorded with the other two dilutions of Ketaconazole and ethyl acetate extract.

In conclusion the anti-microbial activity depends on numerous factors like plant material, techniques employed, growth medium and microorganisms tested. Though, there was nil inhibition at lower concentrations (10 ppm and 1 ppm) of ethyl acetate extract, it was hoped that they might produce comparable effect on further purifications and/or isolation of the active constituents. The study of *Andrographis echinoides* as anti-microbial agent was found necessary for gaining insight into this medicinal flora and its real value, for which the studies on mechanisms of action, interactions with antibiotics or other medicinal plants or compounds and the pharmacokinetic profile of the extracts should be given high priority. The extract could be utilized for preparing economic and effective herbal drugs for pathogenic infection and lead to the identification of active ingredients which is the need of the hour, because successful prediction of lead molecule and its properties as a drug at the onset of drug discovery will pay off later in drug development.

References

- Atta-ur-Rahman and Choudhary MI. 1995. Diterpenoid and steroidal alkaloids. *Nat. Prod. Rep.* 12: 361-379.
- Eisenberg, D.M., R.C. Kessler, C. Foster, F.E. Norlock, D.R. Calkins and T.L. Delbanco. 1993. Unconventional medicine in the United States. *N. Engl. J. Med.*, 328: 246-252.
- Geissman, T.A. 1963. Flavonoid compounds, tannins, lignins and related compounds. In: Pyrrole pigments, isoprenoid compounds and phenolic plant constituents. (Eds.) Florkin, M. and E.H. Stotz, Elsevier, NewYork, 9: 265.
- Ghosh, M., D. Thangamani, M. Thapliyal, R. Yasodha and K. Gurumurthi. 2004. Isolation of *Andrographis paniculata*

- Leaf Protein with Anti-fungal Property. *Acta Phytopathologica et Entomologica Hungarica*, 39(4): 377-381.
- Guhabakshi GN, Sensarma P, Pal DC. 1999. *A Lexicon of Medicinal Plants in India*. NAYA PROKASH, Calcutta.
- Parez, C., M. Pauli and P. Bazevgue. 1990. An antibiotic assay by the agar well diffusion method. *Acta Biologiae et Medicine Experimentalis*, 15: 113-115.
- Romero CD, Chopin SF, Buck G, Martinez E, Garcia M, Bixby L. 2005. Antibacterial properties of common herbal remedies of the south west. *Journal of Ethnopharmacology* 99: 253-257.
- Schultes, R.E. 1978. The Kingdom of Plants. In: *Medicines from the Earth*. (Ed.) Thomson, W.A.R. McGraw-Hill Book Co., New York: 208.

How to cite this article:

Hemalatha, P., V. Sivakumar, R. Parimaladevi and Tilak, M. 2020. Studies on Anti-fungal Activity of Leaf Extracts of *Andrographis echinoides* (L.) Nees. *Int.J.Curr.Microbiol.App.Sci*. 9(10): 3853-3858. doi: <https://doi.org/10.20546/ijcmas.2020.910.443>