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EMS Induced Variability for Physico-Quality Traits in Groundnut (*Arachis hypogaea* L.)

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ABSTRACT

Keywords

Induced variability, Physico-Quality, Groundnut

Article Info

Accepted: 12 October 2018 Available Online: 10 November 2018 An investigation was carried out to study the effects of various doses (0.2 to 0.6%) of EMS on physico-quality traits of groundnut ($Arachis\ hypogea\ L$) in M_4 families of the groundnut variety TPG 41. Among the different concentrations of EMS treated TPG 41 population, 0.2 % EMS was found to be effective in inducing variability for pod yield per plant, hundred kernel weight, shelling out turn and kernel length in M_4 families. Phenotypic correlation coefficients between different traits in M_4 showed significant and positive correlation between hundred pod and kernel weight, kernel length and width with pod yield per plant in all treatments. Shelling out turn was significantly and negatively correlated with kernel length. Of the total of 129 families, more than 20 families showed superior physical and yield attributes than TPG 41. The promising mutants identified in M_4 generation with respect to physical and quality traits need further confirmation through large scale evaluation.

Introduction

Groundnut is an important edible oilseed legume crop grown in mainly in arid and semi-arid tropics of the world. In India, it covers an area of 45 lakh hectares with a production of 67.7lakh tons and a productivity level of 1484 kg/ha (FAOSTAT, 2015).

In India, oil is the ultimate economic product of groundnut crop. Increased availability of cheaper oils for both food and confectionery purposes has resulted in the change in consumer preference and hence groundnut oil is relegated to the lower ranks (Govindraj *et al.*, 2016), hence the future of groundnut crop

lies in its use as food and confectionery rather than exclusively as oil type.

Edible quality and export worthiness of groundnut is mainly determined by physical factors viz., larger seed size, consistency of seed mass and shape, integrity of seed testa, absence of immature seeds, integrity of the seed at the time of processing, and blanching efficiency (Dwivedi and Nigam, 2005). Seed size coupled with the nutritional quality can also determine the worth of groundnut for direct consumption. The nutritional quality is decided by kernel chemical composition (protein, oil content, fatty acids) of kernels.

Quality of oil mainly depends on fatty acid composition and two unsaturated fatty acids, oleic and linoleic acids which together constitute 80% of the groundnut oil. The higher the O/L ratio, higher the oil stability (Holley and Hammons, 1968). However, variation for O/L ratio is very narrow (1 to 2.5) in cultivated types (Bishi et al., 2015; Nawade et al., 2016). The first high-oleate (80%) mutant line, F435 identified by Nordenas early as 1987. In released groundnut varieties of India, oleic acid content ranged from 38 to 60% oleic acid (Nawade et al., 2016). Recently, Gangadhara and Nadaf (2016) developed high oleate (>70%) groundnut lines coupled with foliar disease (rust and LLS) resistance using high oleate mutant GM 4-3 through backcross breeding. Janila et al., (2016) and Bera et al., (2018) introgressed fatty acid desaturase mutant alleles from SunOleic 95R through marker assisted backcross breeding. However, to create additional variability for both physical and oil quality traits of groundnut mutation breeding was resorted using the popular groundnut variety TPG 41.

Materials and Methods

TPG 41 is a large seeded Spanish groundnut variety released for summer cultivation in India (Kale et al., 2004). One thousand five hundred pure, healthy and dry seeds (moisture, 12%) of the groundnut variety TPG 41 was treated with five concentrations of Ethyl Methane Sulphonate (EMS) namely, 0.2%, 0.3%, 0.4%, 0.5% and 0.6% each with 300 seeds per treatment at ICAR-Directorate of Groundnut Research, Junagadh, Gujarat, India during Rabi-Summer-2015. Seeds of the M₁ generation were sown in Kharif 2015 in the field. Plants harvested in bulk gave rise to the M₂ population. About 138 individual plants were harvested from different doses of EMS treatments in M₂ generations (78 in 0.2%; 28 in 0.3%; 16 in 0.4 %; 10 in 0.5% and 6 in

0.6%) based on distinct morphological and pod features during *Rabi*-Summer, 2016. These plants were planted as progeny rows during *kharif*-2016. Of these 10 mutants could not germinate, hence 129 single plants could be harvested.

During kharif-2017, 129 mutant families were planted as M₄ generation along with parent control (TPG 41) and evaluated for physical and oil quality traits. In M₄ generation, physical traits viz., hundred pod and kernel weight, shelling out turn (%), sound mature kernel (%), kernel length and width and pod yield per plant(g) were measured. The fatty acids profiling of 22 selected mutant lines along with TPG 41 were analysed using gas chromatography system (Thermo fisher, Trace GC 1100) equipped with flame ionization detector (FID). The fatty acid methyl esters were passed through capillary column (TRwax) and esters of fatty acids were estimated (Misra and Mathur, 1998). The inlet, FID detector were set to 2400 C and oven at 1900 C whereas carrier gas (nitrogen) and fuel gas (hydrogen) were maintained at 30 mL per min. Total run time for each sample was 12 min and the peaks (Fig. 2) were identified by comparison to a FAME standard mix RM-3 (sigma-Aldrich, St. Louis, Mo). The data were averaged on M₄ lines belonging to each treatment and subjected to the statistical analyses. The distribution of different physical and yield related traits were plotted using Past Software (Fig. 3).

Results and Discussion

Variability for physical and productive traits induced by different doses of EMS

There were significant differences among M_4 families (Table 1) with respect to physical (shelling out turn (%), sound mature kernel (%) and kernel length) traits. Lower concentration of EMS (0.2%) could able to

produce a range of pod yield per plant varying from 4.5 g to 19.1 g with an average value of 11.3g. At 0.5 % EMS, pod yield plant ranged from 6.4 g to 18.5g with an average value of 12.5g. Hundred pod weight was minimum (55g) in 0.2% EMS derived families and maximum (129g) in 0.3% EMS derived families (Fig. 1). Hundred kernel weight ranged from 33g to 45g with an average value of 39g in 0.5% EMS derived families, whereas in 0.2% EMS derived families, it ranged from 24 g to 46g with an average value of 38g. Shelling out turn was minimum (66%) in 0.5% EMS derived families and maximum (76%) in both 0.2% and 0.3% EMS derived families.

Sound mature kernel (%) ranged from 54% to 87% in 0.2% EMS derived families with mean value of 71%. Kernel width was minimum (7.2mm) in 0.2% derived EMS families and maximum (9.6mm) in 0.5% EMS derived families. Kernel length was minimum (11.7 mm) in 0.2% EMS derived families and maximum (17mm) in 0.3% and 0.5% EMS derived families. Thus it has been observed that, among the different doses of EMS derived TPG 41 families, 0.2% EMS was found to be effective in inducing variability for five important traits *viz.*, pod yield per plant, hundred kernel weight, shelling out turn and kernel length in M₄ generation.

Isolation of superior families for physical traits

As the market price and consumer preference are determined by shape and size of the pod and seed of groundnut, identification of high yielding genotypes coupled with attractive pod and kernel characteristics are very important. Various physical traits *viz.*, sound mature kernel(%), hundred kernel weight, kernels with elongated shape, tapering ends are useful for table purpose (Nigam *et al.*, 1989). Superior mutants isolated for physical and yield related traits are presented in Table 2.

Mutants *viz.*, #94(129g), #121(121g), #118 (120g) and #105(115g) showed higher hundred pod weight (g) compared to TPG 41 (96g). Two mutants namely #34 and #105 exhibited 46 g of hundred kernel weight compared to TPG 41(38 g). Higher shelling out turn indicated more kernel weight and preferable as shelling out turn is one of the important traits of trade. For sound mature kernel (%), four mutants #34(87%), #45(86%), #62(85%) and #130(83%) were found superior over TPG 41(69%).

Kernel length was longer in mutants viz., #121, #118 and #94 (17mm each) than TPG 41 (13.8mm). Three mutants viz., #27(19 g), #124(18.5 g) and #20(18 g) exhibited higher pod yield per plant than TPG 41 (8 g). Recovery of productive mutants in groundnut for pod yield and related traits using different doses (0.3% to 0.5%) of EMS has been well documented in groundnut (Prasad, 1984; Gowda et al., 1996; Mathur et al., 2000). In groundnut using mutagenesis followed by hybridization and selection high yielding and large seeded varieties (TG1, TG17, TG 22, TG 39, Somnath, TPG 41) have been developed by Bhaba Atomic Research Center (BARC), Mumbai and SAUs (Patil, 1975; Patil 1977; Mouli et al., 1989; Mouli et al., 1990, Kale et al., 2000, 2004).

Fatty acid profiling of selected EMS mutagenized mutants derived from TPG 41

Seed size coupled with the better fatty acid composition (High oleic acid and low linoleic acid content) is preferred for direct consumption as confectionery or table purpose due to enhanced shelf life and health benefits. High oleic acid content in groundnut is governed by two recessive alleles (Moore and Knauft, 1989; Gangadhara and Nadaf, 2016) but role of modifiers and additional epistatic interactions was also reported (Isleib *et al.*, 1996) in Virginia/Spanish types.

 $\textbf{Table.1} \ \, \textbf{Trait variation for physical and yield related traits in } \, M_4 \ generation$

Trait	Treatmen	Minimu	Maximu	Mean	SD	SE
Pod yield per plant (g)	TPG 41	8.00	8.57	8.25	0.23	0.10
	0.2 %	4.50	19.15	11.33	2.50	0.29
	0.3 %	3.98	16.90	10.50	2.82	0.56
	0.4 %	5.62	15.12	10.20	2.66	0.71
	0.5 %	6.41	18.52	12.54	3.95	1.32
	0.6 %	5.75	15.21	10.39	3.53	1.44
Hundred pod weight (g)	TPG 41	94.60	97.44	96.50	1.21	0.54
	0.2 %	55.29	108.64	91.32	7.61	0.88
	0.3 %	89.06	129.21	97.59	7.57	1.51
	0.4 %	80.62	114.93	90.79	8.43	2.25
	0.5 %	84.76	120.52	102.5	12.0	4.00
	0.6 %	74.01	103.51	89.48	9.98	4.07
Hundred kernel weight	TPG 41	38.14	40.12	38.82	0.93	0.42
(g)	0.2 %	24.73	46.82	38.07	3.02	0.35
	0.3 %	35.81	45.45	39.35	2.02	0.40
	0.4 %	33.59	46.36	37.62	3.01	0.80
	0.5 %	33.60	45.93	39.72	3.67	1.22
	0.6 %	31.75	42.37	36.13	3.68	1.50
	TPG 41	71.00	72.84	72.14	0.76	0.34
Shelling Out turn (%)	0.2 %	68.08	76.24	73.69	1.58	0.18
	0.3 %	67.19	76.64	74.02	1.84	0.37
	0.4 %	71.78	75.83	73.78	0.90	0.24
	0.5 %	66.90	75.22	72.77	2.48	0.83
	0.6 %	71.07	75.37	73.43	1.47	0.60
Sound Mature kernel	TPG 41	68.00	69.70	69.08	0.65	0.29
(%)	0.2 %	54.55	87.92	71.66	6.48	0.75
	0.3 %	64.80	79.78	73.03	4.04	0.81
	0.4 %	64.60	80.32	71.11	4.96	1.33
	0.5 %	62.63	74.37	68.46	4.24	1.41
	0.6 %	65.55	83.89	71.48	6.85	2.80
Kernel width (mm)	TPG 41	8.40	8.60	8.52	0.08	0.04
	0.2 %	7.20	9.10	8.35	0.34	0.04
	0.3 %	7.90	9.20	8.42	0.34	0.07
	0.4 %	7.80	8.80	8.26	0.33	0.09
	0.5 %	8.30	9.60	8.79	0.48	0.16
	0.6 %	8.40	8.60	8.45	0.08	0.03
Kernel length (mm)	TPG 41	13.70	14.30	13.88	0.25	0.11
	0.2 %	11.70	16.50	15.02	0.82	0.09
	0.3 %	14.30	17.10	15.17	0.69	0.14
	0.4 %	14.60	16.10	15.19	0.48	0.13
	0.5 %	14.60	17.10	15.78	0.90	0.30
	0.6 %	14.50	15.90	15.13	0.57	0.23

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Table.2 Superior mutants for physical and yield traits in M₄ generation

Mutant	Т	Pod yield per plant (g)	Mutant	Т	Hundred pod weight (g)	Mutant	Т	Hundred kernel weight (g)	Mutant	Т	Shelling out turn (%)	Mutant	Т	Sound mature kernel (%)	Mutant	Т	Kernel length (mm)
27	T1	19.1	94	T2	129	34	T1	46.82	83		76.64	34	T1	87.92	121	T4	17.1
124	T4	18.5	121	T4	121	105	T3	46.36	12	T1	76.24	45	T1	86.41	118	T4	17.1
20	T1	18.0	118	T4	120	118	T4	45.93	11	T1	76.00	62	T1	85.31	94	T2	17.1
98	T2	16.9	105	T3	115	94	T2	45.45	100	T2	75.99	130	T5	83.89	78	T2	16.8
18	T 1	16.7	45	T1	109	121	T4	44.72	71	T1	75.97	41	T1	80.95	17	T1	16.5
75	T1	16.6	117	T4	108	53	T1	44.06	105	T3	75.83	71	T1	80.68	29	T1	16.5
57	T1	16.2	83	T2	106	45	T1	43.24	51	T1	75.82	38	T1	80.56	57	T1	16.4
100	T2	16.2	124	T4	104	32	T1	42.82	23	T1	75.68	105	T3	80.32	50	T1	16.4
121	T4	16.0	130	T5	104	83	T2	42.43	98	T2	75.68	76	T1	79.78	116	T4	16.4
117	T4	15.9	40	T1	102	130	T5	42.37	6	T1	75.57	92	T2	79.78	14	T1	16.2
130	T5	15.2	18	T1	102	50	T 1	41.61	127	T5	75.37	68	T1	79.67	3	T1	16.2
107	T3	15.1	116	T4	102	13	T1	41.29	80	T2	75.34	83	T2	79.17	77	T2	16.2
54	T1	15.0	29	T1	101	12	T1	41.26	18	T1	75.32	57	T1	79.10	27	T1	16.2
21	T1	14.5	49	T1	101	98	T2	41.22	82	T2	75.22	91	T2	78.77	102	T3	16.1
66	T1	14.5	81	T2	100	18	T1	41.12	123	T4	75.22	70	T1	78.21	35	T1	16.1
77	T2	14.3	87	T2	100	55	T1	41.11	13	T1	75.21	64	T1	78.09	76	T1	16.1
64	T 1	14.1	82	T2	100	92	T2	40.98	2	T1	75.20	53	T1	77.98	113	T3	16.1
108	T3	14.0	98	T2	100	75	T1	40.92	88	T2	75.14	52	T1	77.97	130	T5	15.9
25	T 1	13.6	12	T1	100	62	T1	40.89	57	T1	75.14	85	T2	77.90	45	T1	15.9
51	T1	13.6	39	T1	100	84	T2	40.88	34	T1	75.11	75	T1	77.51	79	T2	15.9
10	T 1	13.5	16	T1	99	97	T2	40.87	10	T1	75.09	94	T2	77.49	64	T1	15.8
118	T4	13.5	51	T1	99	27	T1	40.85	31	T1	75.09	96	T2	77.17	8	T1	15.8
TPG 41		8.25			96			38.8			72			69			13.8

EMS 0.2 % (T₁) EMS 0.5 % (T₄) EMS 0.3 % (T₂) EMS 0.6 % (T₅) EMS 0.4 % (T₃) Control (TPG 41)

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 $\textbf{Table.3} \ \text{Fatty acid composition of selected mutants in } M_4 \ \text{generation of groundnut}$

Mutant	EMS	%	0/0	0/0	0/0	%	%	0/0	0/0	0/0	0/0	%
	Treatment	(C16:0)	(C16:1)	(C18:0)	(C18:1)	(C18:2)	(C18:3)	(C20:0)	(C20:1)	(C22:0)	(C22:1)	(C24:0)
17	0.2 %	11.034	0.054	2.311	56.30	23.97	0.066	1.207	1.289	2.54	0.069	1.16
21	0.2 %	11.284	0.042	2.258	56.39	23.72	0.07	1.206	1.289	2.53	0.084	1.13
35	0.2 %	10.255	0.045	2.594	58.54	21.43	0.072	1.259	1.42	2.77	0.097	1.52
36	0.2 %	10.337	0.037	3.123	60.35	19.85	0.056	1.297	1.229	2.42	0.072	1.23
39	0.2 %	11.065	0.041	3.112	61.26	18.62	0.056	1.203	1.18	2.33	0.074	1.06
46	0.2 %	10.81	0.049	2.763	58.27	21.72	0.051	1.205	1.259	2.61	0.074	1.19
53	0.2 %	10.022	0.069	2.72	61.54	19.11	0.056	1.327	1.361	2.55	0.076	1.17
57	0.2 %	10.745	0.069	2.986	58.64	20.87	0.07	1.344	1.196	2.68	0.078	1.32
64	0.2 %	10.356	0.049	2.649	59.75	20.62	0.045	1.256	1.278	2.63	0.084	1.28
67	0.2 %	11.123	0.05	2.66	56.85	22.97	0.073	1.215	1.234	2.58	0.072	1.17
72	0.2 %	10.42	0.029	2.369	58.33	21.60	0.073	1.227	1.512	3.00	0.095	1.35
77	0.3 %	10.67	0.044	2.182	58.99	21.35	0.063	1.241	1.383	2.67	0.092	1.32
78	0.3 %	11.374	0.037	2.228	55.09	24.61	0.064	1.221	1.301	2.70	0.077	1.31
93	0.3 %	11.683	0.041	2.384	55.30	23.64	0.076	1.213	1.364	2.80	0.096	1.40
106	0.4 %	10.447	0.054	2.67	58.53	21.54	0.068	1.305	1.348	2.63	0.083	1.32
108	0.4 %	10.685	0.037	2.855	59.04	21.01	0.059	1.228	1.313	2.61	0.083	1.09
117	0.5 %	10.586	0.044	2.767	54.42	24.59	0.065	1.432	1.286	3.12	0.077	1.62
122	0.5 %	10.297	0.045	2.773	60.96	18.98	0.054	1.271	1.424	2.75	0.094	1.36
124	0.5 %	10.088	0.058	2.429	60.11	20.67	0.054	1.298	1.347	2.60	0.083	1.26
128	0.6 %	10.852	0.055	2.671	57.71	22.27	0.072	1.262	1.296	2.52	0.068	1.23
130	0.6 %	11.037	0.025	2.296	57.96	22.80	0.045	1.208	1.178	2.35	0.059	1.04
1	TPG 41 (C)	11.147	0.06	3.298	54.30	25.19	0.0553	1.433	0.9987	2.4257	0.053	1.04

C16:0-Palmitic Acid	C16:1-Palmitoleic Acid	C18:0-Stearic Acid	C18:1-Oleic Acid	C18:2-Linoleic Acid	C18:3-Linolenic Acid
C20:0-Arachidic Acid	C20:1-Gadoleic Acid	C22:0-Behenic Acid	C22:1-Erucic Acid	C24:0-Lignoceric Acid	

 $\textbf{Table.4} \ \text{Estimated oil quality parameters of selected mutants in} \ M_4 \ \text{generation of groundnut}$

Mutant	EMS Treatment	LCSFA	SFA	MUFA	PUFA	USFA	PUFA/SFA ratio	USFA/SFA ratio	(C18:1)/ (C18:2) ratio
17	0.2 %	4.91	18.25	57.71	24.04	81.75	1.317	4.479	2.349
21	0.2 %	4.86	18.40	57.81	23.79	81.60	1.293	4.434	2.378
35	0.2 %	5.55	18.40	60.10	21.50	81.60	1.169	4.436	2.731
36	0.2 %	4.95	18.41	61.69	19.90	81.59	1.081	4.433	3.041
39	0.2 %	4.59	18.77	62.55	18.68	81.23	0.995	4.328	3.290
46	0.2 %	5.00	18.58	59.66	21.77	81.42	1.172	4.383	2.683
53	0.2 %	5.05	17.80	63.05	19.16	82.21	1.077	4.620	3.221
57	0.2 %	5.35	19.08	59.98	20.94	80.92	1.098	4.242	2.809
64	0.2 %	5.17	18.17	61.16	20.67	81.83	1.137	4.503	2.898
67	0.2 %	4.97	18.75	58.21	23.04	81.25	1.229	4.334	2.475
72	0.2 %	5.58	18.37	59.97	21.67	81.64	1.180	4.445	2.701
77	0.3 %	5.22	18.08	60.51	21.41	81.92	1.184	4.532	2.763
78	0.3 %	5.23	18.83	56.50	24.67	81.17	1.310	4.311	2.239
93	0.3 %	5.41	19.48	56.80	23.72	80.52	1.218	4.134	2.339
106	0.4 %	5.26	18.37	60.02	21.61	81.63	1.176	4.443	2.717
108	0.4 %	4.92	18.46	60.47	21.07	81.54	1.141	4.416	2.811
117	0.5 %	6.17	19.52	55.83	24.65	80.48	1.263	4.122	2.213
122	0.5 %	5.38	18.45	62.52	19.04	81.56	1.032	4.422	3.211
124	0.5 %	5.16	17.68	61.60	20.72	82.32	1.172	4.657	2.909
128	0.6 %	5.01	18.53	59.13	22.34	81.47	1.205	4.396	2.592
130	0.6 %	4.60	17.93	59.22	22.85	82.07	1.274	4.577	2.542
1	TPG 41 (C)	4.90	19.35	55.41	25.24	80.65	1.305	4.168	2.156

LCSFA-Long chain saturated fatty acids	SFA- Saturated fatty acids	MUFA-Monounsaturated fatty acids
PUFA- Polyunsaturated fatty acids	USFA- Unsaturated fatty acids	

Table.5 Phenotypic correlation coefficients for physical and yield traits

	Pod yield per plant (g)	Hundred pod weight (g)	Hundred Kernel weight (g)	Shelling Out turn (%)	Sound Mature kernel (%)	Kernel width (mm)	Kernel length (mm)
Pod yield per plant (g)	1						
Hundred pod weight (g)	.391** (T1)	1					
How does d	.672* (T4)	012** (T1)					
Hundred Kernel weight	0.40tr (G)	.813** (T1)					
(g)	.949* (C)	.859** (T2)	1				
	.329** (T1)	.804** (T4)					
		.944** (T5)					
Shelling Out		.311** (T1)	.365** (T1)	1			
turn (%)	-	561** (T2)					
Sound Mature		.921* (C)	.537** (T1)				
kernel (%)		.431** (T1)	.575** (T2)				
	-	.400* (T2)	.748** (T3)	.262* (T1)	1		
		.590* (T3)	.974** (T5)				
		.752*(T4)					
		.852* (T5)					
Kernel width		.933* (C)	.279* (T1)				
(mm)	.822* (T5)	.310** (T1)	.442* (T2)		.820* (T5)	1	
		.501* (T2)	.792* (T4)	-			
		.737* (T5)	.813* (T5)				
Kernel length	.884* (C)	.449** (T1)	.916* (C)	952* (C)	.700* (T4)	.351** (T1)	
(mm)	.389** (T1)	.467* (T2)	.458** (T1)	628** (T2)	.928** (T5)		1
	.765** (T3)	.845* (T5)	.812** (T4)	691** (T3)			
			.922** (T5)				
EMS 0.2 %	(T_{\star}) EN	MS 0.3 % (T ₂)	EMS 0.4 9	6 (T _a)			

EMS 0.2 % (T₁) EMS 0.5 % (T₄) EMS 0.3 % (T₂) EMS 0.6 % (T₅) EMS 0.4 % (T₃) Control (TPG 41)

Fig.1 Box plots showing distribution of physical and yield related traits among different EMS treatments in M_4 generations of TPG 41

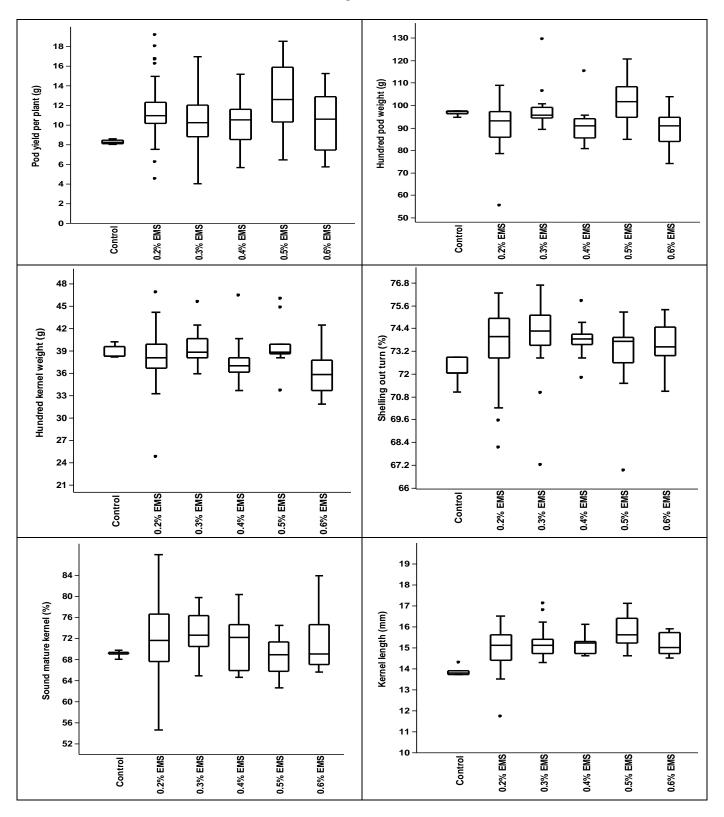


Fig.2 Chromatogram of fatty acid methyl esters separated in gas chromatography of fatty acid profiles of TPG 41 (control)

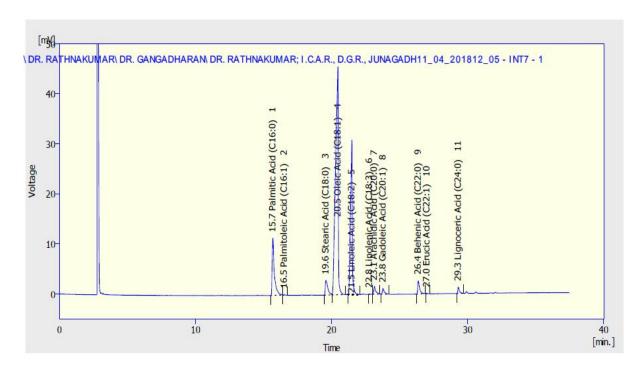
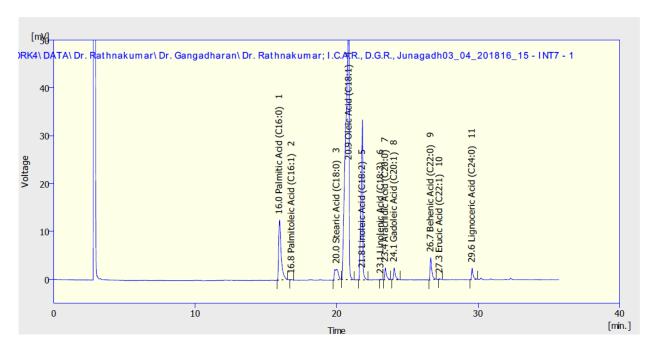


Fig.3 Chromatogram of fatty acid methyl esters separated in gas chromatography of fatty acid profiles of mutant Number 22 derived from EMS treatment



The fatty acid profiling of 22 mutants (Table 3), showed narrow range of oleic (54-61%) and linoleic acid (18-24%) content compared

to TPG 41 (54% oleic acid, 25% linoleic acid) suggesting less role of EMS in inducing mutation for *Ol* locus. However, two mutants

#39 (61.26%) and #53 (61.54%) had slightly more oleic acid content compared TPG 41(54.3%). Linoleic (C18:2), a polyunsaturated fatty acid linked with oxidative rancidity of oils when heated at high temperatures (Isleib *et al.*, 2006) leading to formation of trans-fatty acids and causes cardiovascular diseases (Wang *et al.*, 2015). Three mutants #39 (18.62%), #122(18.98%) and #53(19.11%) exhibited low linoleic acid content than TPG 41(25.19%).

The trait high oleic to linoleic acid ratio (high O/L) in groundnut is favoured over low O/L as it confers health benefits (Garcia *et al.*, 2006) and oil stability. Groundnut with high oleate contents could maintain relatively more flavour after roasting as well as after longer periods of storage compared to low oleate types (Nepote *et al.*, 2006).

Regarding oil stability, high O/L ratios in groundnut extend shelf life by delaying the development of rancidity (O'Keefe *et al.*,1993). Four mutants #39(3.2), #53(3.2), #122 (3.2) and #36 (3.04) exhibited slightly improved higher O/L ratio compared to TPG 41(2.5) (Table 4).

Higher dietary intakes of major saturated fats are associated with an increased risk of coronary heart disease (Zong et al., 2016) by increasing LDL cholesterol as well as total cholesterol. Two mutants #53(17.8%) and #124(17.68%) showed slightly lower saturated fatty acids than TPG 41 (19.35). Total monounsaturated fats was higher in #53(63.05%) and #39(62.55%) mutants compared to TPG 41 (55.41). Three mutants #39(18.68%), #122(19.04), #53(19.16) had lower polyunsaturated fats than 41(25.24). In addition to these parameters few mutants also exhibited slightly higher polyunsaturated/unsaturated ratio and unsaturated/saturated fatty acid ratios (Table 5).

Relationship between physical and yield traits

Phenotypic correlation coefficients between the traits in M₄ were calculated and presented in Table 5. The results showed that pod yield per plant was associated positively with hundred pod and kernel weight (Venkataravana et al., 2000), kernel length and width in all treatments. Sound mature kernel (%) showed significant positive correlation with hundred pod and kernel weight as well as kernel length and width in all treatments. Shelling out turn (%) was found to be correlated negatively correlated with kernel length.

Mutagenesis by EMS treatment generated considerable variation for physical and productive traits. Of the total of 129 families, more than 20 families showed superior physical and yield attributes than TPG 41. It has been observed that the lower mutagenic dose (0.2%-0.3% EMS) had beneficial effect on physical and productive characters. The superior mutants identified in M₄ generation with respect to pod yield per plant, hundred kernel weight, kernel length, shelling out turn and O/L ratio could be used as donors in breeding programmes for development of confectionery type groundnuts. These mutants need to be evaluated for further confirmation in large scale evaluation.

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