

Original Research Article

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## Assessment of Genetic Diversity in *Labeo gonius* (Hamilton, 1822) through Microsatellite Marker in Nanak Sagar and Dhaura Reservoirs of Uttarakhand, India

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### ABSTRACT

The present study deals with the assessment of genetic diversity using microsatellite marker in the fish *Labeo gonius* from Nanak Sagar and Dhaura reservoirs of Uttarakhand having different morpho-edaphic features and self-recruiting populations of this fish. These reservoirs are distantly located and distinctly separated without any connection having negligible possibility of gene exchange with each other. Total 12 cross amplified microsatellite primers after using software Primer-BLAST and Primer-3 were screened in all 100 DNA samples of fish collected from both the reservoirs. 12 cross amplified microsatellite primers were screened and successfully amplified. After PCR amplification of microsatellite loci and performing native PAGE using amplified DNA samples as above, POP GENE Version 1.32 was used to calculate Nei's observed heterozygosity, expected heterozygosity, Nei's genetic diversity, Fixation index (Fis) and Shannon's information index (SI) and genetic variability indices viz. Gene flow(Nm), the coefficients of genetic differentiation (Fst & Gst) and Nei's genetic distance. Overall Gst value (0.1601) recorded for *L. gonius* suggested the possibility of less gene exchange among the two stocks and indicated that 16.01% variation was attributable to interstock divergence, while 83.99% to individual differences within the stocks. Nei's genetic diversity, calculated from the banding pattern of every primer, ranged from 0.4368 to 0.6922 with mean value of 0.5732 for specimens from Dhaura reservoir whereas it ranged from 0.5136 to 0.8243 with mean value of 0.6770 in specimens from Nanak Sagar reservoir. The observed and expected heterozygosities ranged from 0.4769 to 0.4981(mean value- 0.4901) and 0.5014 to 0.5267(mean value- 0.5169) respectively in *L. gonius* from Dhaura reservoir. The observed and expected heterozygosities ranged from 0.4641 to 0.5314 (mean value- 0.5046) and 0.4768 to 0.5682 (mean value- 0.5225) respectively in Nanak Sagar reservoir stock. Slightly better level of observed heterozygosity observed in fish from Nanak Sagar reservoir than Dhaura reservoir might be due to more differentiated stock of Nanak Sagar. Lesser value of observed heterozygosity compared to expected heterozygosity in fish from Dhaura reservoir might be possibly due to increased inbreeding in successive generations owing to small population size restricting desired germplasm exchange of appropriate genetic diversity. These observations indicated that the *L. gonius* stock of Nanak Sagar reservoir is genetically more diverse and differentiated as compared to its stock from Dhaura reservoir.

### Keywords

Genetic characterization, Heterozygosity microsatellites, Primers, *Labeo gonius*.

### Article Info

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## Introduction

*Labeo gonius* (Hamilton, 1822), a common Cyprinid species, is widely distributed in water bodies of North India, Asom, Odisha and along the east coast up to the Krishna river in India. It is a dominant fish species in different reservoirs of Uttarakhand and has a larger scope as potential candidate species for aquaculture. It spawns during the south-west monsoon during July to August. In Uttarakhand state a major part of aquatic resources is covered by reservoir (18,931 ha) followed by rivers (2700 km) and ponds (600 ha). Two major reservoirs of Uttarakhand viz. Dhaura (1200 ha) and Nanak Sagar (4262 ha) are greatly subjected to environmental aberrations like reduction in water volume, increased sedimentation and water abstraction, catchment area degradation due to siltation, drying of water during summer season, conversion of water area into marshy land due to soil erosion caused by deforestation which makes status of different fishes highly vulnerable and subject to unpredictable genetic changes in these reservoirs. The fish fauna of these reservoirs mainly comprises of various species of catfishes, major carps and minor carps. *L. gonius*, a minor carp contributes substantially in catches of these reservoirs and enjoys a good market value as a food fish in the state. At present, its fishery is flourishing well in these reservoirs but for insuring good future prospects for *L. gonius* fishery early attention is required before its decline as practically no attention has been paid for stock assessment, sustainable utilization and genetic management of this species till now. Status of genetic structure of fish is considered essential for their controlled propagation, stock improvement and developing conservation plans. To conserve intra-specific genetic diversity of the fish the status of genetic diversity of the concerned species is a pre-requisite for developing management

strategies. The data on genetic diversity can be effectively used to provide scientific basis for developing measures for conservation and management of natural population of the targeted species. Heterozygosity is an important measurement of genetic diversity level and has got much attention from ecologists and aqua-culturists (Xu *et al.*, 2001). The best estimate of genetic variation in natural population is the mean observed heterozygosity ( $H_o$ ) per locus which varies non-randomly between loci, populations and species (Allendorf and Utter, 1979). Use of molecular markers, especially microsatellites, has been recognized to have great potential in revolutionizing the genetic management of fishery stocks for controlling the level of inbreeding and loss of genetic diversity through its ability to detect genetic uniqueness of individuals, populations or species (Lakra *et al.*, 2007). Microsatellites or simple sequence repeats (SSRs) are short tandem repeat motifs (1-6 bases) with high level of allelic polymorphism and co-dominant inheritance (DeWoody and Avise, 2000). Microsatellites tend to be evenly distributed in the genome on all chromosomes and all regions of the chromosome (Chistiakov *et al.*, 2006). Microsatellites have been found inside gene coding regions, introns and in the non-gene sequences (Liu *et al.*, 2001). Microsatellites markers are preferable because they are co-dominant and highly polymorphic as compared to other genetic markers (Liu and Cordes, 2004) and have been proved to be very useful in revealing information about allele frequency, heterozygosity, population differentiation, inbreeding co-efficient, gene flow, linkage disequilibrium, stock identification and other parameters that are crucial measures of genetic diversity and population genetics. Recently, microsatellite markers have been developed for some Indian fishes such as *Labeo rohita* (Das *et al.*, 2005; Patel *et al.*, 2009) and *Catla catla* (McConnell *et al.*,

2001), *Chitala chitala* (Puniya *et al.*,2006) and *Cirrhinus mrigala* (Lal *et al.*,2011). Previously genetic diversity studies in *L. gonius* from three reservoirs of Uttarkhand has been done through allozyme and RAPD markers (Tewari *et al.*, 2013a and b) but accuracy of predictions based on these markers has always been a matter of concern. Present study is the first attempt to assess the genetic variability status of *L. gonius* from two reservoirs- Dhaura and Nanak Sagar located in Tarai region of Uttarakhand through microsatellite marker with the aim of devising desirable management practices to conserve the available genetic resources of this fish for its sustainable production in these reservoirs.

### **Materials and Methods**

Hundred live specimens (50 from each reservoir) of the fish, *L. gonius* were collected for present study from commercial catches of Nanak Sagar and Dhaura reservoirs. Kidney tissue were dissected out by using sterilized scissors and forceps and stored at -86°C in deep freezer for further analysis. DNA was isolated from the dissected kidney tissue using DNA isolation kit purchased from Genei (Ltd.), Bangalore, India. Total twelve cross amplified microsatellite primers were used. To amplify the repeat regions, primers were analysed using the web based tool (<http://primer3.sourceforge.net>) Primer3 (Rozen and Skaletsky, 2000) with an optimum annealing temperature of 55°C and a minimum GC content of 40-70%.

### **Amplification of microsatellite loci and analysis of microsatellite data**

All the twenty microsatellite primers were initially screened in 2-2 DNA samples of *L. gonius* collected from Nanak Sagar and Dhaura reservoirs. A total of 12 cross amplified microsatellite loci were

successfully amplified which produced clear and polymorphic bands. PCR amplification of microsatellite loci were performed in a 25 µl reaction mixture, which included 1X PCR buffer (10 mM Tris-HCl pH 9.0, 50 mM KCl), 0.2 mM of each dNTP, 2.0 mM of MgCl<sub>2</sub>, 5 p mol of each primer, 1.5 U Taq DNA polymerase and 25–50 ng of template DNA using PCR (Eppendorf, USA). Initial denaturation was performed at 94°C for 3 minutes followed by 30 cycles of 94°C for 30 seconds, locus specific annealing temperatures (52-64°C) for 60 seconds and 72°C for 90 seconds and a final elongation of 1 cycle at 72 °C for 8 min followed by storing it at 4°C. Amplified products were mixed with 2 µl of gel loading dye and then separated on 6% denaturing polyacrylamide gel with 1x TBE on PAGE Gel along with standard marker Φ X 174/ Hinf I marker at constant power supply of 25 volts for 2 hrs. Polymorphic information content (PIC) of individual primer was estimated using the

formula:  $PIC = 1 - 1/n \sum_{i=1}^n P_{ij}^2$  Where  $P_{ij}$  is the frequency of  $j$ th allele. After performing native PAGE, POP GENE Version 1.32 (Raymond and Rousset, 1998) was used to calculate Nei's observed heterozygosity ( $H_o$ ), expected heterozygosity ( $H_e$ ), Fixation index ( $F_{is}$ ) and genetic variability indices viz. Gene flow ( $N_m$ ), the coefficients of genetic differentiation ( $F_{st}$  &  $G_{st}$ ) and Nei's genetic distance. Nei's genetic diversity ( $H_i$ ) was calculated from the banding pattern of every screened primer across all the loci. Individual genotypes were scored using the Gene Mapper (version 4.0; Applied Biosystems) with a size standard and an internal control for allele calling. Each allele was coded according to its size in nucleotide base pairs (bp). Possible null alleles and genotyping errors caused by stuttering and/or large-allele dropout were tested using MICRO-CHECKER (Van Oosterhout *et al.*, 2004). Scoring and human error were estimated by

duplicate analyses. The polymorphic information content (PIC) was calculated by using the CERVUS version 3.03 (Kalinowski *et al.*, 2007).

## **Results and Discussion**

### **Screening of microsatellite primers in *L. gonius***

Microsatellite primers selected for further study based on amplifying successfully and exhibiting desired level of polymorphism have been summarized in table 1. Twelve Primer-BLAST cross amplified microsatellite primers for *L. gonius* exhibited polymorphism.

### **Microsatellite primers amplification in *L. gonius* from Nanak Sagar reservoir**

Microsatellite primers amplification values across all loci in *L. gonius* from Nanak Sagar reservoir are recorded in table 2. Number of alleles per locus ranged from 5-9 with mean value of 7.08 per locus. A total of 8 SSR loci were scored by the primer PL-01. The product size ranged from 0.11Kb to 0.30 Kb and the average expected genetic diversity and PIC value of the primer were 0.635 and 0.71 respectively.

A total number of 7 SSR loci were scored by the primer PL-02. The product size ranged from 0.14 Kb to 0.35 Kb and the average expected genetic diversity and PIC value of the primer were 0.783 and 0.68 respectively. The totals of 6 SSR loci were scored for the primer PL-03 with product size ranged from 0.25 to 0.37 Kb and the average expected genetic diversity and PIC value of the primer were 0.635 and 0.63 respectively. A total of 8 SSR polymorphic loci were scored for the primer PL-08. The product size ranged from 0.22 Kb to 0.48 Kb and the average expected genetic diversity and PIC value of the primer

were 0.680 and 0.69 respectively. Total numbers of 6 SSR loci were scored by the primer PL-10. The product size ranged from 0.18 Kb to 0.70 Kb and the average expected genetic diversity and PIC value of the primer were 0.765 and 0.60 respectively. 7 SSR loci were scored by the primer PL-11 and the product size was 0.25 Kb to 0.65 Kb and the average expected genetic diversity and PIC value of the primer were 0.737 and 0.64. A total of 8 SSR loci with product size ranged 0.20 Kb to 0.70 Kb were scored for the primer PL-13. The average expected genetic diversity and PIC value were 0.824 and 0.68 respectively.

Total 5 SSR loci were scored by the primer PL-14 and the average expected genetic diversity and PIC value of the primer were 0.796 and 0.63 respectively and product size ranged from 0.10 to 0.25 kb. 6 SSR loci were scored by the primer PL-15 and the average expected genetic diversity and PIC value of the primer were 0.513 and 0.61 respectively and product size ranged from 0.13 to 0.40 kb. 7 SSR loci were scored by the primer PL-16 and the average expected genetic diversity and PIC value of the primer were 0.576 and 0.65 respectively. Product size ranged from 0.17 to 0.40 kb. 9 SSR loci were scored by the primer PL-17 and the average expected genetic diversity and PIC value of the primer were 0.650 and 0.74 respectively and product size ranged from 0.15 to 0.30 kb. 8 SSR loci were scored by the primer PL-20 and the average expected genetic diversity and PIC value of the primer were 0.630 and 0.67 respectively and product size ranged from 0.20 to 0.40 kb.

### **Microsatellite primers amplification in *L. gonius* from Dhaura reservoir**

Microsatellite primers amplification values across all loci in *L. gonius* from Dhaura reservoir are recorded in table 2. Number of

alleles per locus ranged from 5-9 with mean value of 6.91 per locus. A total of 7 SSR loci were scored by the primer PL-01. The product size ranged from 0.10 Kb to 0.25 Kb and the average expected genetic diversity and PIC value of the primer were 0.510 and 0.67 respectively. A total number of 6 SSR loci were scored by the primer PL-02. The product size ranged from 0.20 Kb to 0.40 Kb and the average expected genetic diversity and PIC value of the primer were 0.634 and 0.60 respectively.

The total of 7 SSR loci were scored for the primer PL-03 with product size ranged from 0.20 to 0.42 Kb and the average expected genetic diversity and PIC value of the primer were 0.540 and 0.65 respectively. The total of 9 SSR loci was scored for the primer PL-08. The product size ranged from 0.18 Kb to 0.53 Kb and the average expected genetic diversity and PIC value of the primer were 0.436 and 0.73 respectively. Total numbers of 6 SSR loci were scored by the primer PL-10 and the product size ranged from 0.38 Kb to 0.65 Kb and the average expected genetic diversity and PIC value of the primer were 0.595 and 0.61 respectively. 7 SSR loci was scored by the primer PL-11 and the product size was 0.32 Kb to 0.68 Kb and the expected genetic diversity and PIC value of the primer were 0.652 and 0.64 respectively. 9 SSR loci with product size ranged 0.28 Kb to 0.74 Kb was scored for the primer PL-13.

The average expected genetic diversity and PIC value were 0.623 and 0.73 respectively. 6 SSR loci were scored by the primer PL-14 and the average expected genetic diversity and PIC value of the primer were 0.571 and 0.61 respectively and product size ranged from 0.10 to 0.20 kb. 8 SSR loci were scored by the primer PL-15 and the average expected genetic diversity and PIC value of the primer were 0.692 and 0.66 respectively and product size ranged from 0.25 to 0.35 kb. 5 SSR loci were scored by the primer PL-16 and the

average expected genetic diversity and PIC value of the primer were 0.497 and 0.58 respectively.

Product size ranged from 0.25 to 0.45 kb. A total of 7 SSR loci were scored by the primer PL-17 and the average expected genetic diversity and PIC value of the primer were 0.548 and 0.63 respectively and product size ranged from 0.15 to 0.30 kb. 6 SSR loci were scored by the primer PL-20 and the average expected genetic diversity and PIC value of the primer were 0.578 and 0.61 respectively and product size ranged from 0.20 to 0.45 kb.

### **Microsatellite variation based Genetic diversity in *L. gonius* from Nanak Sagar and Dhaura reservoirs**

Microsatellite variation based genetic diversity values in *L. gonius* from Dhaura and Nanak Sagar reservoirs are recorded in tables 3 and 4 respectively. Nei's genetic diversity values ranged from 0.4368 to 0.6922 with mean value of 0.5732 for specimens from Dhaura reservoir whereas it ranged from 0.5136 to 0.8243 with mean value of 0.6770 in specimens from Nanak Sagar reservoir.

The observed and expected heterozygosities ranged from 0.4769 to 0.4981 (mean value 0.4901) and 0.5014 to 0.5267 (mean value-0.5169) respectively in *L. gonius* from Dhaura reservoir.

The observed and expected heterozygosities ranged from 0.4641 to 0.5314 (mean value-0.5046) and 0.4768 to 0.5682 (mean value-0.5225) respectively in Nanak Sagar reservoir stock. The mean values of Fis were found to be 0.124 in Nanak Sagar reservoir and 0.145 in Dhaura reservoir. Mean values for Shannon's information index for all microsatellite loci in *L. gonius* were 1.1862 for Dhaura reservoir population and 1.2342 for Nanak Sagar population.

**Table.1** Primer-BLAST designed microsatellite primers for *L. gonius*

Locus	Primer Sequence(5'-3')	Annealing Temp	Annealing Time
PL-01	F-GAAAGCTGCTCGTCCTTGAA R-GAAAGCTGCTCGTCCTTGAA	52 °C	1min 30 sec
PL-02	F-GGGTGTGGGAGAGAAAGAGAG R-GGAGTCTGACAAATGCAGCAAG	64 °C	1 min
PL-03	F-TCTCAGTGGGTGTCATTACCTG R-CCCATCAAACCATCTCTCTAGC	52 °C	1min
PL-08	F-CTGACACTCTTATCTCGCTGCC R-GACCTGAGCAAACAAACCTCAT	53 °C	1min 30 sec
PL-10	F-TCTCTCTTTGTCTTTCCCTTG R-CACAAGCCACTGTTTAGCTTCA	64 °C	1min
PL-11	F-CAAATCTGTGAACATGCAAGC R-CCTAGTCCCCTCTAGTCAGCA	58 °C	1 min 30 sec
PL-13	F-AGATAAGACCCTTCTTCCTCGG R-TTTATTAGGGAGCGTTCGAGTG	64 °C	1min
PL-14	F-CTGTTGGTGACTGTAGGGTGAA R-GAGAACTCGGTTTGAACATGC	58 °C	1min 30 sec
PL-15	F-ACAGTAATCTTGTGTCTGTCTCTC R-GTCTAAACGTGTCTGAGCTGTG	57 °C	1 min 30 sec
PL-16	F-TGAATGTTTCCAGTCACCACAT R-GTAATGCAGCGGAGAATAAACC	57 °C	1min
PL-17	F-ACAATTCCTGTGTCAACTGTGC R-TACCGTCTCAGTCTCTTTTCGG	55 °C	1min
PL-20	F-ATAGTCGAAATTGGTCCTCTGC R- CAATACCATGACTGAAGTGCC	55 °C	1min 30 sec

**Table.2** Microsatellite primers amplification in *L. gonius* from Nanak Sagar and Dhaura reservoir

Locus	Nanak Sagar reservoir			Dhaura reservoir		
	Amplified Product (Kb)	Number of alleles	(PIC)	Amplified Product (Kb)	Number of alleles	(PIC)
PL- 01	0.11-0.30	8	0.71	0.10-0.25	7	0.67
PL-02	0.14-0.35	7	0.68	0.20-0.40	6	0.60
PL-03	0.25-0.37	6	0.63	0.20-0.42	7	0.65
PL-08	0.22-0.48	8	0.69	0.18-0.53	9	0.73
PL-10	0.18-0.70	6	0.60	0.38-0.65	6	0.61
PL-11	0.25-0.65	7	0.64	0.32-0.68	7	0.64
PL-13	0.20-0.70	8	0.68	0.28-0.74	9	0.73
PL-14	0.10-0.25	5	0.59	0.10-0.20	6	0.61
PL-15	0.13-0.4	6	0.61	0.25-0.35	8	0.66
PL-16	0.17-0.40	7	0.65	0.25-0.45	5	0.58
PL-17	0.15-0.30	9	0.74	0.15-0.30	7	0.63
PL-20	0.2-0.40	8	0.67	0.2-0.45	6	0.61

**Table.3** Genetic Diversity of *L. gonius* from Dhaura Reservoir based on Microsatellite markers

Locus	Observed Heterozygosity (Ho)	Expected Heterozygosity (He)	Nei's genetic Diversity (Hi)	Shannon's Information Index	Fixation Index Fis (-ve)
PL- 01	0.4769	0.5014	0.510	1.2596	0.123
PL-02	0.4961	0.5042	0.634	1.0764	0.133
PL-03	0.4883	0.5286	0.540	1.1889	0.148
PL-08	0.4961	0.5143	0.436	1.1743	0.185
PL-10	0.4865	0.5221	0.595	1.2573	0.132
PL-11	0.4947	0.5129	0.652	1.2589	0.110
PL-13	0.4838	0.5224	0.623	1.1320	0.105
PL-14	0.4919	0.5231	0.571	1.1250	0.103
PL-15	0.4861	0.5127	0.692	1.1124	0.111
PL-16	0.4959	0.5228	0.497	1.4021	0.109
PL-17	0.4872	0.5125	0.548	1.2269	0.099
PL-20	0.4981	0.5267	0.578	1.0214	0.092
<b>Mean</b>	<b>0.4901</b>	<b>0.5169</b>	<b>0.5732</b>	<b>1.1862</b>	<b>0.145</b>

**Table.4** Genetic Diversity of *L. gonius* from Nanak Sagar based on Microsatellite markers

Locus	Observed Heterozygosity (Ho)	Expected Heterozygosity (He)	Nei's genetic Diversity (Hi)	Shannon's Information Index	Fixation Index Fis (-ve)
PL- 01	0.5171	0.5289	0.635	1.3675	0.103
PL-02	0.5282	0.5475	0.783	1.3354	0.120
PL-03	0.4737	0.4958	0.635	1.2519	0.118
PL-08	0.5314	0.5682	0.680	1.4005	0.155
PL-10	0.5287	0.5494	0.765	1.4299	0.105
PL-11	0.4721	0.4986	0.737	1.0563	0.090
PL-13	0.5135	0.5327	0.824	1.3321	0.085
PL-14	0.4641	0.4768	0.796	1.1042	0.098
PL-15	0.4859	0.4981	0.513	1.0368	0.091
PL-16	0.5278	0.5379	0.576	1.1525	0.081
PL-17	0.5282	0.5380	0.650	1.1103	0.115
PL-20	0.4853	0.4988	0.630	1.2335	0.081
<b>Mean</b>	<b>0.5046</b>	<b>0.5225</b>	<b>0.6770</b>	<b>1.2342</b>	<b>0.124</b>

**Table.5** Genetic variability indices in *L. gonius* stocks from Nanak Sagar and Dhaura reservoirs

Parameters	Values	
Coefficient of genetic differentiation (Fst)	0.093	
Estimation of Gene flow (Nm)	1.276	
Total genetic diversity in population (Ht)	0.5274	
Within sample genetic diversity (Hs)	0.4430	
Coefficient of genetic differentiation (Gst)	0.1601	
Nei's genetic distance	0.2134	
P-Value	0.0184*	
	Nanak Sagar Reservoir	Dhaura Reservoir
Observed number of alleles (na)	4.9805	4.8126
Effective number of alleles (ne)	4.7762	4.5531

\*Significant at  $P < 0.05$

### **Genetic divergence in *L. gonius* stocks from Nanak Sagar and Dhaura reservoirs**

Observations related with genetic divergence in *L. gonius* stocks from both reservoirs are presented in table 5. Genetic differentiation (P-value) for *L. gonius* across all loci among different population pairs in Nanak Sagar and Dhaura reservoirs was found to be 0.0184. Values of Gene flow (Nm) and Nei's genetic distance among reservoir populations of *L. gonius* were found to be 1.276 and 0.1768 respectively. Values of coefficients of genetic differentiation (Fst & Gst) observed were 0.093 and 0.1632 respectively for overall population of *L. gonius*. Total genetic diversity in overall population (Ht) and within sample genetic diversity (Hs) was 0.5274 and 0.4430, respectively. The observed (na) and effective number of alleles (ne) in *L. gonius* from Nanak Sagar reservoir were found to be 4.9805 and 4.7762 respectively and for Dhaura reservoir these values were 4.8126 and 4.5531 respectively.

The genetic diversity value (0.6770) based on observed and expected heterozygosities (0.5046 and 0.5225) in *L. gonius* from Nanak Sagar reservoir compared to genetic diversity value (0.5732) and observed and expected heterozygosities (0.4901 and 0.5169) from Dhaura reservoir indicated that its stock in Nanak Sagar reservoir exhibited better genetic diversity. These reservoirs are distantly located and distinctly separated without any connection hence having negligible possibility of gene exchange with each other and this also might be responsible for varied sub- structuring of the stocks of *L. gonius*. The values of Nei's genetic diversity in Nanak Sagar reservoir (0.6770) and in Dhaura reservoir (0.5732) in *L. gonius* through microsatellite marker are found to be higher than Nei's genetic diversity values in these reservoirs (0.3980 and 0.2243) in *L. gonius* through RAPD marker (Tewari *et al.*, 2013

b). Genetic diversity values based on codominant microsatellite markers are more accurate and preferable as compared to allozyme and dominant RAPD markers as these markers are highly polymorphic, capable of detecting small genetic differences (even single nucleotide base change variation) which is essential in studies on the genetic variability of the populations (Balloux and Lugonae, 2002). Nei's genetic diversity range (0.436 to 0.824) in *L. gonius* (over all loci) was found to be in similar range (0.679 to 0.874) reported in *T. tambroides* through microsatellite marker by Nguyen *et al.*, (2007). Similar level of observed and expected heterozygosity values in Nanak Sagar population of *L. gonius* and lesser observed heterozygosity value compared to expected heterozygosity in fish from Dhaura might be possibly to some extent correlated with the level of inbreeding incidences in successive generations due to less effective population size limiting germplasm exchange required for maintaining appropriate genetic diversity. Large area of Dhaura gets dried up in summer and most fishes including *L. gonius* are extensively exploited even from deeper isolated pockets thus adversely affecting their effective population size (Ne) available for breeding in the following breeding season. The mean values of observed heterozygosity in *L. gonius* from both the reservoirs were found to be comparable with the mean value of observed heterozygosity (0.46) reported for some other freshwater fishes (DeWoody and Avise, 2000). The observed heterozygosity range (over all loci) in *L. gonius* are found to be comparable with the observed heterozygosity range (0.0000 to 0.9000) reported in *Tor putitora* using seven microsatellites loci developed from *Catla catla* and *Barbus barbus* by Mohindra *et al.*, (2004). Same observations (heterozygosity range- 0.10–1.00 and 0.500 to 0.870) were reported for other cyprinid fishes like silver carp and bighead carp by Tong *et al.*, (2002);



and in three wild and one farm population of *L. rohita* by Sahoo *et al.*, (2014), respectively. However, the observed heterozygosity range in *L. gonius* was found to be higher than in *Cirrhinus mrigala* (0.247 to 0.333) reported from different rivers by Lal *et al.*, (2004). The mean values of observed and expected heterozygosity in *L. gonius* from Nanak Sagar population using microsatellite marker was found to be comparable with the mean values (0.501 and 0.539) of observed and expected heterozygosity using allozymes marker in *L. gonius* from Nanak Sagar population (Tewari *et al.*, 2013a). Small differences between observed values of population genetic diversity ( $H_s=0.4430$ ) and total genetic diversity ( $H_t=0.5274$ ) indicated moderate genetic differentiation among *L. gonius* stocks. Nei's genetic diversity results in *L. gonius* in present findings are well correlated with the observations made by Singh *et al.*, (2015) where five microsatellite loci was used to study the genetic diversity and characterization of different strains of common carp *C. carpio* L. and reported mean observed heterozygosity ranged (0.45 and 0.62) while expected heterozygosity ranged (0.32–0.68).

On the basis of calculated ( $n_a=4.9805$  and  $4.8126$ ) observed number of alleles and effective ( $n_e=4.7762$  and  $4.5531$ ) number of alleles in Nanak Sagar and Dhaura reservoirs it is indicated that significant genetic variation is there within stocks of *L. gonius* in both reservoirs. However, relatively lower genetic variability in Dhaura stock as compared to Nanak Sagar stocks might be correlated with effective population size owing to fishing pattern in them. The problems of bottleneck, genetic drift and inbreeding depression, are correlated with small populations, effective population size to population genetic structure of fishes (Ayappan, 2011). Overall  $G_{st}$  value (0.1601) calculated for *L. gonius* suggested the possibility of less gene exchange among

the two stocks and indicated that 16.01% variation was attributable to interstock divergence, while 83.99% to individual differences within the stocks. The moderate level genetic differentiation on the basis of calculated value of coefficient of genetic differentiation across all loci ( $F_{st}=0.093$ ) was present due to different population size as Dhaura reservoir have smaller population size than Nanak Sagar reservoir and this pattern of variation is relatively higher corresponds to that obtained (low level of differentiation) in other Indian fresh water fishes through microsatellites reported by Chaturvedi *et al.*, (2011) in *L. dero* ( $F_{st}=0.019$ ) and by Gopalakrishnan *et al.*, (2009) in *L. dussumieri* ( $F_{st}=0.041$ ) whereas Appleyard and Mather, (2002) reported high  $F_{ST}$  values (0.501 to 0.598) in two species of *Oreochromis* indicating there was little evidence of introgression between these species and Singh *et al.*, (2012) reported low  $F_{st}$  value (0.035) in *L. calbasu*. Effective number of migrants per generation, an indicator of gene flow ( $N_m$ ), calculated (1.276) among *L. gonius* populations is an indicative of negligible migration resulting in low gene flow level which indicated that very little gene exchange among the populations because of less number of migrants as these reservoirs are distinctly located from each other. Moreover, they are not interconnected at any point reject the chance of migration and gene exchange. Nei's genetic distance calculated between pairs of stocks of *L. gonius* from two reservoirs reveal larger genetic distance value (0.1768) between Nanak Sagar and Dhaura reservoir stocks. Value of genetic distance among *L. gonius* population is correlated with the value of genetic distance ( $D=0.171$  to  $0.199$ ) reported by Khoo *et al.*, (2002) in guppy fish, population ( $D= 0.085-0.249$ ). Comparison among the separate population pairs from these two reservoirs, distantly located from each other and having no connection with each other,  $P$ -value (0.0184)

show significant genetic differentiation at  $P < 0.05$  among Nanak Sagar and Dhaura reservoirs populations. As genetic distance value increases with the increase in geographic distance, the observations on genetic distance in stocks of *L. gonius* from both the reservoirs might be correlated with the geographical distances (70 km) between the stocks of two distantly separated reservoirs.

On the basis of observations on heterozygosity ( $H_o$  and  $H_e$ ), Nei's genetic diversity ( $H_i$ ), Fixation Index ( $F_{is}$ ), Shannon's Information Index (SI) and Genetic variability indices it can be inferred that *L. gonius* stock of Nanak Sagar reservoir is genetically more diverse and moderately differentiated compared Dhaura reservoir stock.

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