

Original Research Article

<https://doi.org/10.20546/ijcmas.2017.612.437>

## Antibacterial Activity of *Punica granatum* (Pomegranate) Fruit Peel Extract against Pathogenic and Drug Resistance Bacterial Strains

Alka Chaudhary<sup>1</sup> and Siddarth Nandan Rahul<sup>2\*</sup>

<sup>1</sup>Department of Microbiology C.C.S. University Meerut- 250110, UP, India

<sup>2</sup>Department of Biotechnology, Agriculture Sciences and Agricultural Informatics, Shobhit University, Meerut- 250110, UP, India

\*Corresponding author

### ABSTRACT

The Botanical name of Pomegranate is *Punica granatum* and it is a bearing fruit shrub, belonging to the family Lythraceae. *Punica granatum* originated in the region of northern India. Every part of the *Punica granatum* is useful for our body such as seeds and fibers which provide vitamins-C, vitamins-K, Folate. Seeds of *Punica granatum* contain oil acids such as Punica acid, Palmitic acid, stearic acid and Oleic acid. *Punica granatum* peel also shows Antioxidant and Antibacterial activity. Antibacterial activities of peel extract were studied against some bacterial strains as- *Pseudomonas fluorescens*, *Pseudomonas aeruginosa*, *Shigella flexneri*, *Klebsiella pneumoniae*, *Salmonella typhi*, and *Bacillus subtilis* (all are gram negative and also pathogenic to the human). The *Punica granatum* peel extract were prepared in three organic solvents- Methanol, Ethanol and Benzene. All extract showed the degree of effectiveness against all bacterial strains but the Methanol extract showed the maximum growth inhibition 85.71% against *Klebsiella pneumoniae* bacteria at 100µl/ml extract concentration. *Klebsiella pneumoniae* is pathogenic and cause disease Pneumonia and can progression in to serve bacterial infection to bloodstream infection, wound infection, urinary tract infection and meningitis. It is also resistance for some antibiotics such as Lincomycin and Oleandomycin. The present study depict that the *Punica granatum* fruit peel extract (which is waste in industry) showed good Antibacterial activity against given pathogenic and some drug resistance bacterial strains.

### Keywords

*Punica Granatum* (Pomegranate) fruit peel, Pathogenic bacterial strains, Antimicrobial activity, Disc diffusion method, Minimum Inhibitory Concentration.

### Article Info

#### Accepted:

28 October 2017

#### Available Online:

10 December 2017

### Introduction

Medicinal plants have always been a good source to find new remedies for human health problems. Recently a wide range of these plants have been screened for antimicrobial property (Martin and Ernst 2003, Upadhyay *et al.*, 2010).

*Punica granatum* commonly called as Pomegranate, recently describe as nature power fruit, is a plant used in folkloric medicine for the treatment of various diseases

(Adbel moneim *et al.*, 2011; Ajay kumar *et al.*, 2005) widely cultivated in Mediterranean region. Pomegranate has strong antioxidant and anti-inflammatory properties recent studies have demonstrated its anticancer activity in several human cancers (Adhami and Mukhtar, 2007; Longtin, 2003). In addition pomegranate peel extract with an abundance of flavonoid and tannins has been show to have a high antioxidant activity (Abdel moneim *et al.*, 2011).

Pomegranate (*Punica granatum* L.) is the fruit of a tree belonging to the family Punicaceae. It is native from Iran to the Himalayas in northern India and has been cultivated and naturalized over the entire Mediterranean region since ancient times (Jurenka, 2008 and Meerts *et al.*, 2009). The ripe fruit contains many arils separated by a white, membranous pericarp (Jurenka, 2008). Studies show that pomegranate juice (PJ) has potent antioxidant activity (ability to scavenge free radicals), significantly higher than more commonly consumed fruit juices such as grape, cranberry, grapefruit, and orange (Azadzoi *et al.*, 2005; Basu 2009; Guet *et al.*, 2008).

This activity has been attributed to antioxidant properties of polyphenols, including ellagitannins (hydrolyzable tannins) and anthocyanins (condensed tannins) (Cowan, 1999). Punicalagins are the major ellagitannins in the whole fruit and can be hydrolyzed to ellagic acid and other smaller polyphenols *in vivo* (Jurenka, 2008; Lansky, 2007). Pomegranate has been used in traditional medicine for the treatment of dysentery, diarrhea, helminthiasis, and respiratory pathologies (Lansky 2005–Sánchez-Lamar, 2008). Currently there is considerable interest toward evaluating plant sources for alternative treatments against pathogenic bacteria which are now showing resistant to many drugs. Pomegranate (*Punica granatum*) peel is an inedible part obtained during processing of pomegranate juice which is completely waste in the industry. In the present investigation we have tested various extract of *Punica granatum* against pathogenic and drug resistant bacterial strains.

## **Materials and Methods**

### **Preparation of plant material**

The plant material (fruit part) was collected from the Cotton Research Station D.M. road

Bulandshahr region UP (India) during the starting of winter season. Collected plant material was clean up and crushed into powder form using pestle mortal.

### **Extract preparation**

Fruit peel extract were prepared by immersing 1gm of powder in 5ml with four different organic solvents as Methanol, Ethanol and Benzene for 24 hours after filtration the extract were evaporated by the help of rotator evaporator. For stock solution each extract was re-dissolved with 5ml DMSO (dimethyl sulphoxide).

### **Test microbial strains**

All tested bacterial strains viz., *Pseudomonas aeruginosa* (MTCC 162), *Pseudomonas fluorescens* (MTCC 254), *Proteus vulgaris* (MTCC 123), *Bacillus subtilis* (MTCC 251), *Klebsiella pneumonia* (MTCC 140), and *Shigella flexneri* (MTCC 182), *Salmonella typhi* (MTCC 326) were collected from MTCC (microbial type culture collection) IMTECH Chandigarh. These microorganisms were maintained on nutrient agar media (NAM) at 30°C for further investigation.

### **Antibacterial screening**

The antimicrobial screening of the bacterial strains were carried out disc diffusion method (Grover and Moore 1962). The plant extracts of 0.1ml were mixed in 0.9 ml of pre sterilized nutrient broth and then added 0.1ml bacterial culture suspension. In control sets, DMSO (in place of the plant extract) was used in the medium in appropriate amount. Culture tubes were incubated for 24 hours at 30°C.

After incubation, sterile disc of 6mm (Himedia) were dip in to the broth (treated as well as control separately), disc were

aseptically inoculated on the agar surface of the nutrient agar medium in plates. Inoculated Petriplates were incubated at 30°C and the observations were recorded after 24 hours. Percentage of bacterial growth inhibition (BGI %) was calculated per formula.

$$\text{BGI (\%)} = \frac{\text{dc} - \text{dt}}{\text{dc}} \times 100$$

Where,

dc= diameter of control

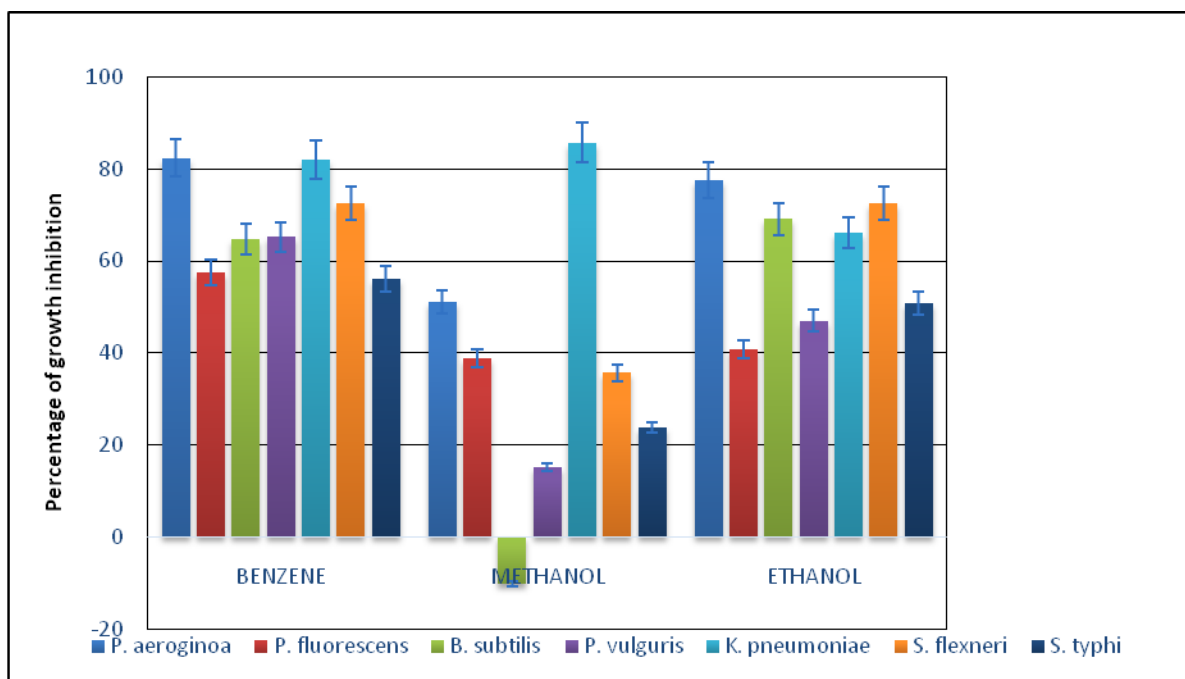
dt= diameter of test

### Results and Discussion

Some bacterial strains were drug resistance for some antibiotic viz., *Klebsiella pneumoniae* was resistance for many drugs as Lincomycin and Oleandomycin. *Pseudomonas aeruginosa* was resistance for many drugs as Tobramycin, Lincomycin and Oleandomycin (Table 1). The result of

antibacterial activity of Pomegranate (*Punica granatum*) fruit peel was determined by disc diffusion method. All the extract of Pomegranate showed the antibacterial activity against all the pathogenic bacterial strains. The Methanol extract of Pomegranate fruit peel showed maximum growth inhibition 85.71% against *Klebsiella pneumoniae* and also showed minimum growth inhibition - 10.29% against *Bacillus subtilis*. The Ethanol extract of Pomegranate fruit peel showed maximum growth inhibition 77.45% against *Pseudomonas aeruginosa* and also showed minimum growth inhibition 40.07% against *Pseudomonas fluorescens*. The Benzene extract of Pomegranate fruit peel showed maximum growth inhibition 82.35% against *Pseudomonas aeruginosa* and also showed minimum growth inhibition 55.93% against *Salmonella typhi* (Table 2). The MIC value of Methanol extract against *Klebsiella pneumoniae*, in Ethanol extract against *Pseudomonas aeruginosa*, and in Benzene extract against *Pseudomonas aeruginosa* showed in Table 3.

Fig.1 Effect of different extracts on bacterial growth



**Table.1** Zone of inhibition by different drugs against bacterial strains (Sahrawat and Shahi, 2013)

Antibiotics	Zone of drug against bacterial strains (mm)						
	<i>Proteus vulgaris</i>	<i>Pseudomonas aeruginosa</i>	<i>Pseudomonas fluorescens</i>	<i>Bacillus subtilis</i>	<i>Klebsiella pneumoniae</i>	<i>Shigella flexneri</i>	<i>Salmonella typhi</i>
Tobramycin	21	-	-	14.2	23	19.5	10.0
Cephaloridine	17.5	12.02	-	-	14.5	-	8.50
Kanamycin	22.5	10.0	11.5	15.6	12.5	16.5	7.50
Lincomycin	-	-	-	-	-	-	-
Norfloxacin	13	-	-	17.5	18.5	18.5	17.5
Oleandomycin	-	-	-	-	-	-	-

**Table.2** Antibacterial screening of different extract of Pomegranate fruit peel against pathogenic and drug resistance bacterial strains by Disc diffusion method

Pathogens	Percentage of growth inhibition at 100µl/ml		
	Methanol	Ethanol	Benzene
<i>Pseudomonas aeruginosa</i>	41.17%	48.52%	-32.03%
<i>Pseudomonas fluorescens</i>	1.85%	25.92%	40.74%
<i>Bacillus subtilis</i>	42.15%	62.74%	67.64%
<i>Proteus vulgaris</i>	25.07%	0%	-28.07%
<i>Klebsiella pneumoniae</i>	61.65%	80.04%	81.20%
<i>Bacillus subtilis</i>	-10.09	69.11	64.70
<i>Shigella flexneri</i>	35.63	72.41	72.41

**Table.3** Minimum inhibitory concentration of Pomegranate fruit peel against bacterial strains

Bacterial strains	MIC against pathogens in µl/ml		
	Methanol	Ethanol	Benzene
<i>Klebsiella pneumoniae</i>	$1.56 \times 10^{-6}$	-	-
<i>Pseudomonas aeruginosa</i>	-	$12.5 \times 10^{-3}$	-
<i>Pseudomonas aeruginosa</i>	-	-	$6.25 \times 10^{-4}$

As the present study on the antimicrobial activity of Pomegranate extract on the several human pathogenic bacteria (Fig. 1). The study shows that the various extract of Pomegranate can successfully control these kinds of bacteria as the other researchers have revealed that pomegranate peel is a rich source of tannins, flavonoids and other phenolic compounds (Li *et al.*, 2006). Antioxidant and antibacterial properties of pomegranate peel in in-vitro model systems have been reported (Negi and Jayaprakasha, 2003; Reddy *et al.*,

2007; Opara *et al.*, 2009; Alzoreky 2009). As the study says that Benzene extract of Pomegranate fruit peel can inhibit the population of both *Pseudomonas aeruginosa* and *Salmonella typhi* respectively which is already shown by many scientist like Lansky, 2005; Al-Zoreky, 2009; Gould *et al.*, 2009; Machado *et al.*, 2003; Opara *et al.*, 2008 in their study which was on antimicrobial activity of pomegranate has been documented with different extracts and the potential therapeutic applications of the antimicrobial

properties of pomegranate components have been investigated in human and murine models (Jurenka, 2008; Basu, 2009; Aviram *et al.*, 2008; Betanzos *et al.*, 2011; Sumner *et al.*, 2005; Rock *et al.*, 2008; Rosenblat *et al.*, 2006; Shukla *et al.*, 2008). However, FPJ antimicrobial activity has not been explored in spite of its high content of antioxidants.

Overall it can be concluded that Pomegranate (*Punica granatum* L.) peel is a good source of antibacterial compounds against pathogenic and drug resistance bacterial strains and it can be used as a drug against pathogens. The study reveals that different extraction with different solvents has antimicrobial activity and it should be further investigated.

## References

- Abdel MAE, Dkhil MA, Al-Quraishy S (2011) Studies on the effect of pomegranate (*Punica granatum*) juice and peel on liver and kidney in adult male rat. JMPR. In press.
- Adhami VM, Mukhtar H (2007). Antioxidant from green tea and pomegranate for chemoprevention of prostate cancer. Mol biotechnol. 37, 52-57.
- Ajay kumar KB, Asheef M, Babu BH, Padikkala, J(2005). The inhibition of gastric mucosal injury by *Punica granatum* methanolic extract. J. Ethnopharmacol., 96, 171-176.
- Alzoreky NS. Antimicrobial activity of pomegranate (*Punica granatum* L) fruit peels. Int J Food Microbiol. 2009; 13: 24–28.
- Al-Zoreky NS. Antimicrobial activity of pomegranate (*Punica granatum* L.) fruit peels. Int J Food Microbiol. 2009; 134: 244–8.
- Aviram M, Volkova N, Coleman R, Dreher M, Reddy MK, Ferreira D, *et al.*, Pomegranate phenolics from the peels, arils, and flowers are antiatherogenic: studies in vivo in atherosclerotic apolipoprotein e-deficient (E 0) mice and in vitro in cultured macrophages and lipoproteins. J Agric Food Chem. 2008; 56: 1148–57.
- Azadzoï KM, Schulman RN, Aviram M, Siroky MB. Oxidative stress in arteriogenic erectile dysfunction: prophylactic role of antioxidants. J Urol. 2005; 174: 386–93.
- Basu A, Penugonda K. Pomegranate juice: a heart-healthy fruit juice. Nutr Rev. 2009; 67:49–56.
- Betanzos-Cabrera G, Guerrero-Solano JA, Martínez-Pérez MM, Calderon-Ramos ZG, Belefant-Miller H, Cancino-Diaz JC. Pomegranate juice increases levels of paraoxonase1 (PON1) expression and enzymatic activity in streptozotocin-induced diabetic mice fed with a high-fat diet. Food Res Int. 2011; 44: 1381–5.
- Braga LC, Shupp JW, Cummings C, Jett M, Takahashi JA, Carmo LS, *et al.*, Pomegranate extract inhibits *Staphylococcus aureus* growth and subsequent enterotoxin production. J Ethnopharmacol. 2005; 96: 335–9.
- Cowan MM. Plant products as antimicrobial agents. Clin Microbiol Rev. 1999; 12: 564–82.
- Gould SWJ, Fielder MD, Kelly AF, Naughton DP. Anti-microbial activities of pomegranate rind extracts: enhancement by cupric sulphate against clinical isolates of *S. aureus*, MRSA and PVL positive CA-MSSA. BMC Complement Altern Med. 2009; 9: 23.
- Grover, R.K. and Moore, J.D. 1962. Toximetric studies of fungicides against brown rot organism. *Sclerotinia fruticola*. Phytopathology 52: 876-880.
- Guo C, Wei J, Yang J, Xu J, Pang W, Jiang Y. Pomegranate juice is potentially better than apple juice in improving antioxidant function in elderly subjects. Nutr Res. 2008; 28: 72–7.
- Jurenka JS. Therapeutic applications of pomegranate (*Punica granatum* L.): a review. Altern Med Rev. 2008; 13: 128–44.
- Lansky EP, Newman RA. *Punica granatum* (pomegranate) and its potential for prevention and treatment of inflammation and cancer. J Ethnopharmacol. 2007; 109:177–206.
- Li Y, Guo C, Yang J, Wei J, Xu J, Cheng S. Evaluation of antioxidant properties of pomegranate peel extract in comparison with pomegranate pulp extract. Food Chem.

- 2006; 96:254–260. Doi: 10.1016/j.foodchem.2005.02.033.
- Machado TB, Pinto AV, Pinto MCFR, Leal ICR, Silva MG, Amaral ACF, *et al.*, In vitro activity of Brazilian medicinal plants, naturally occurring naphthoquinones and their analogues, against methicillin-resistant *Staphylococcus aureus*. *Int J Antimicrob Agents*. 2003; 21:279–84.
- Martin KW, Earnst E. 2003 Herbal medicine for treatment of bacterial infection' a review of controlled clinical trials. *J antimicrobes chemother* 51, 241-6
- Meerts IATM, Verspeek-Rip CM, Buskens CAF, Keizer HG, Bassaganya-Riera J, Jouni ZE, *et al.*, Toxicological evaluation of pomegranate seed oil. *Food Chem Toxicol*. 2009; 47:1085–92.
- Negi PS, Jayaprakasha GK. Antioxidant and antibacterial activities of *Punica granatum* peel extracts. *J Food Sci*. 2003; 68:1473–1477. doi: 10.1111/j.1365-2621.2003.tb09669.x.
- Opara LU, Al-Ani MR, Al-Shuabi YS. Physico-chemical properties, vitamin C content and antimicrobial properties of pomegranate fruit (*Punica granatum L*) *Food Bioprocess Tech*. 2009; 2:315–321. doi: 10.1007/s11947-008-0095-5.
- Opara LU, Al-Ani MR, Al-Shuaibi YS. Physico-chemical properties, vitamin C content, and antimicrobial properties of pomegranate fruit (*Punica granatum L.*) *Food Bioprocess Technol*. 2008; 2:315–21.
- Reddy M, Gupta S, Jacob M, Khan S, Ferreir D. Antioxidant, antimalarial and antimicrobial activities of tannin-rich fractions, ellagitannins and phenolic acids from *Punica granatum L*. *Planta Med*. 2007; 73: 461–467. doi: 10.1055/s-2007-967167.
- Rock W, Rosenblat M, Miller-Lotan R, Levy AP, Elias M, Aviram M. Consumption of Wonderful variety pomegranate juice and extract by diabetic patient's increases paraoxonase 1 association with high-density lipoprotein and stimulates its catalytic activities. *J Agric Food Chem*. 2008;56:8704–13.
- Rosenblat M, Hayek T, Aviram M. Anti-oxidative effects of pomegranate juice (PJ) consumption by diabetic patients on serum and on macrophages. *Atherosclerosis*. 2006; 187:363–71.
- Sahrawat, A. and Shahi M.P. 2013. Antibacterial screening of *Sapindus mukorossi* geartn fruit and *Daucus carota L.* root extract against pathogenic bacterial strains. *Current discovery* (2) 1. 76-80.
- Sánchez-Lamar A, Fonseca G, Fuentes JL, Cozzi R, Cundari E, Fiore M, *et al.*, Assessment of the genotoxic risk of *Punica granatum L.* (Punicaceae) whole fruit extracts. *J Ethnopharmacol*. 2008; 115:416–22.]
- Shukla M, Gupta K, Rasheed Z, Khan KA, Haqqi TM. Consumption of hydrolyzable tannins-rich pomegranate extract suppresses inflammation and joint damage in rheumatoid arthritis. *Nutrition*. 2008; 24:733–43.
- Sumner MD, Elliott-Eller M, Weidner G, Daubenmier JJ, Chew MH, Marlin R, *et al.*, Effects of pomegranate juice consumption on myocardial perfusion in patients with coronary heart disease. *Am J Cardiol*. 2005; 96:810–14.
- Upadhyay RK, Dwivedi P, Ahmad S. (2010) Screening of antibacterial activity of six plant essential oil against pathogenic bacterial strains. *Asian J Med sci*, 2:152-158

#### How to cite this article:

Alka Chaudhary and Siddarth Nandan Rahul. 2017. Antibacterial Activity of *Punica granatum* (Pomegranate) Fruit Peel Extract against Pathogenic and Drug Resistance Bacterial Strains. *Int.J.Curr.Microbiol.App.Sci*. 6(12): 3802-3807. doi: <https://doi.org/10.20546/ijcmas.2017.612.437>