

Review Article

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Role of Auxin on Growth, Yield and Quality of Tomato - A Review

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ABSTRACT

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A growth regulator, plant growth regulator, or PGR, is a natural or synthetic chemical that is sprayed or otherwise applied to a seed or plant in order to alter its characteristics. They are sometimes referred to as plant hormones. Plant growth regulators function as chemical messengers for intercellular communication. In tomato, different growth regulators play a pivotal role in germination, root development, branching, flower initiation, fruiting, lycopene development, synchronization and early maturation, parthenocarpic fruit development, ripening, TSS, acidity, seed production etcetera. To boost the tomato production in India these versatile resources greatly help the professionals and researchers. By keeping the importance of growth regulator in tomato production this review paper is scripted.

Introduction

Tomato (*Lycopersicon esculentum* Mill.) is one of the most widely cultivated crops in the world. It is an important source of vitamins and an important cash crop for small-holders and medium-scale commercial farmers. It is one of the most popular salad vegetables and is taken with great relish. Food value of tomato is very rich because of higher contents of vitamins A, B and C including calcium and carotene (Bose and Som, 1990). Uddain *et al.*, (2009), Rashid (1983), Davies and Hobes (1981) reported that tomato adds flavor to the foods and it is also rich in medicinal value. Tomato has a significant role in human nutrition because of its rich source of lycopene, minerals and β -carotene which are anti-oxidants and promote good health.

Tomato contains organic acids like citric, malic and acetic acids which is found in fresh tomato fruit, promotes gastric secretion, acts as a blood purifier and works as intestinal antiseptic (Pruthi, 1993).

Among vegetables, tomato occupies 4th position in area and 2nd position in production in India. The present area and production of tomato in the country 8.82 lakh ha and 187.35 lakh tones respectively in 2010 (NHB, 2014). While in Andhra Pradesh, it is cultivated about an area of 1.67 lakh ha with a production of 33 lakh tones (NHB, 2014).

Although tomato plants can grow under a wide range of climatic conditions, they are

extremely sensitive to hot and wet growing conditions (Ahmad, 2002). Increasing temperature, viral diseases and salinity are the major limiting factors in sustaining and increasing tomato productivity (Fekadu and Dandena, 2006). There are generally various constraints resulting in low production of vegetables including tomato which includes poor soil fertility, water scarcity, poor cultivation skills, attack of pest and disease, poor availability of inputs and harsh climate (Baliyan and Kgathi, 2009). Lack of adoptive cultivars and poor fruit setting of existing varieties especially during the hot/dry season where the demand for tomato is very high is one challenge farmers are facing in tomato production even though there is potential land for cultivation. Breeding for heat tolerance in tomato crop has been difficult due to many factors like moderate heritability inheritance being complex or the cultivars becoming lower in yield (George *et al.*, 1984). Tomato fruit set is very sensitive to environmental conditions, in particular, to too low or high temperatures that affect pollen development and anther dehiscence. Fruit set depends on the successful completion of pollination and fertilization (Gillaspy *et al.*, 1993). Tomato requires day temperature of 21–28°C and moderately cool night temperature of 15–20°C for proper fruit setting. High temperature (both day and night), humidity, rainfall and light intensity are the limiting factors of tomato production (Abdulla and Verkerk, 1968). High day and night temperature above 32°C and 21°C, respectively, was reported as limiting factor to fruit-set due to an impaired complex of physiological process in the pistil, which results in floral or fruit abscission (Picken, 1984). High temperatures reduces fruit set, fruit production and yield in tomato (Peet *et al.*, 1997). For good fruit set and better yield, pollination, germination of pollen grains, pollen tubes growth, fertilization and fruit initiation must take place successfully (Kinet

and Peet, 1997). Gelmesa *et al.*, (2010) explained that high relative humidity of the air, low light intensity and extreme low and high temperature, and improper mineral nutrition seems to be involved in the control of those phenomena and result in low fruit set and quality. High day and night temperature above 32°C and 21°C, respectively, was reported as limiting factor to fruit-set due to an impaired complex of physiological process in the pistil, which results in floral or fruit abscission (Picken, 1984).

Plant growth regulators (PGRs) are used extensively in horticulture to enhance plant growth and improve yield by increasing fruit number, fruit set and size (Batlang, 2008 and Serrani *et al.*, 2007a). Use of growth regulators had improved the production of tomato including other vegetables in respect of better growth and quality (Saha, 2009). Fruit set in tomato can be increased by applying plant growth regulators to compensate the deficiency of natural growth substances required for its development (Singh and Choudhury, 1966). Induction of artificial parthenocarpy through application of PGRs enables fertilization-independent fruit development that can reduce yield fluctuation in crops like tomato, pepper and likes (Heuvelink and Korner, 2001). Plant growth regulators such as auxins and gibberellins are known to affect parthenocarpy (Matlob and Kelly, 1975), fruit setting (Rappaport, 1957) and fruit size (Osborne and Went, 1953); therefore synthesized auxins and gibberellins are often used for promotion of fruit set in some fruit vegetable production including tomatoes (Kuo and Tsai, 1984) and yields can increase dramatically to four times Abdulla (1978).

As more native auxin is transported down the stem to the roots, the overall development of the roots is stimulated. The longer and branched root can uptake more nutrients from

the soil which are accumulated to the plant sink and increase the yield (Wang *et al.*, 2005). If the source of IAA is removed, such as by trimming the tips of stems, the roots are less stimulated accordingly.

Rahul *et al.*, (2005) investigated that plant growth was not affected significantly by any treatment and interaction between different doses of PGRs (control, 25 or 75 ppm IAA, and 25 or 75 ppm NAA) and micronutrient (control, 2500 ppm Multiplex or 2000 ppm Humaur) mixtures.

On the other hand, most report indicated that synthetic auxin like 2, 4-D has herbicidal or ephynastic effect (Pandolfini *et al.*, 2002) which lead to flower bud abscission, poor fruit set, fruit defects and puffiness beyond certain concentrations. This is due to herbicidal effect of 2, 4-D at higher concentrations hastened maturity of the plants so that they complete their life cycle in a short period of time. Serrani *et al.*, (2007) indicated that high doses of 2, 4-D resulted in some malformations in tomato.

In fact the use of growth regulators had improved the production of tomato including other vegetable in respect of better growth and quality which ultimately led to generate interest between the scientists and farmers for commercial application of growth regulators. Keeping in view, this review paper is enlisted to summarize the importance of growth regulators in tomato cultivation.

Role of auxin

The most important Auxin produced by plants is indole-3-acetic acid (IAA). It plays important roles in a number of plant activities, including phototropism, gravitropism, apical dominance, fruit development, abscission and root initiation. A couple of synthetic auxins are 2, 4-D and 2, 4, 5-T.

Role of auxin on flowering and fruiting of tomato

Synthesized auxin are often used for promotion of fruit set in some fruit and vegetable production including tomatoes (Gemici *et al.*, 2006; Khan *et al.*, 2006; Serrani *et al.*, 2007; Batlang, 2008). IAA is required for fruit growth and development and delays fruit senescence and plays also a minor role in the initiation of flowering and development of reproductive organs (Asahira *et al.*, 1967). Patel *et al.*, 2012 revealed that application NAA increases the fruit diameter in tomato. Verma *et al.*, 2014 revealed that fruit set in tomato was successfully improved by application of NAA. Leopold (1964) observed that with the increase in concentration of auxin there was a comparable increase in percentage of flower cluster. Mukharji and Roy (1966) found that application of IAA had protected the flower and premature fruit drop and increased length of size fruit in tomato plant. Singh and Upadhayaya (1967) studied the effect of IAA and NAA on tomato and reported that the regulators induce parthenocarpic fruit.

Khaled *et al.* (2015) reported that days required for 50% flowering, days required for fruit setting, fruit cluster plant⁻¹ and fruit plant⁻¹ were significant influenced by the combined application of IAA in BARI tomato 7, Manik and Ratan varieties of tomato. Singh *et al.*, (2003) stated that the beta naphthoxyacetic acid (BNOA) have positive effect on seed germination and fruit set, and earlier flowering in tomato. Baliyan *et al.*, 2013 revealed that application of 4-CPA in summer tomato increases the number of tomato fruit set. The application of 4-CPA hormone has significant effect on the tomato fruit set (Ozguven *et al.*, 1997; Sasaki *et al.*, 2005). Mehta and Mathai (1976) observed that spraying of tomato plant with NAA 0.2 ppm and 0.1 ppm gave significantly increased

fruit set, and number of days taken to fruit setting was significantly lesser to control. Kaushik *et al.*, (1978) showed that alpha NAA at 1, 10, or 100 mg/l increased fruit set per plant at lowest concentration, the highest concentration markedly reduced fruit number when sprayed on tomato plants at the 2 leaf stage. Sagar *et al.*, (1978) noted that NAA 20 ppm as a whole plant spray at flowering stage gave significantly increased fruit number in tomato. It was also noted that significantly higher yield in tomato by foliar application of NAA 10 and 20 ppm at time of first flowering. Younis and Tigani (1978) reported that 2 sprays of NAA 10 ppm at time of flowering stage gave significantly increased flower and fruit set in tomato. Gupta *et al.*, (2001) recorded minimum day for fruit setting in plant was 42 DAT, observed significantly with the treatment of 25 ppm NAA alone with Humaur (P3M2).

Rodrigues *et al.*, (2001) studied that tomato spraying with 10 ppm NAA followed by pollination on initial trusses resulted in the highest number of fruits (45.63) and seed yield (0.58 g per plant). Jagdish *et al.*, (2002) confirmed that spraying PCPA at 50 ppm to the flower clusters significantly improved the fruit set per cluster compared with the control, but increasing the concentration to 100 and 150 ppm had no significant effect on fruit set. NAA spray had no effect on fruit set per cluster when compared with the control. Mukherji and Roy (1966) and Howlett (1941) reported that fruit set in tomato was successfully improved by application of NAA and IAA.

Similarly, sprays of NAA or β -NAA at the time of flowering resulted in reduced pre-harvest fruit drop and increased the number of fruits per plant (Alam and Khan, 2002). Synthetic auxin 4-CPA (4-chloro phenoxy acetic acid) reduced pre-harvest fruit drop with increased number of fruits per plant and

yield (Sasaki *et al.*, 2005). Application of 4-CPA is more effective during anthesis period than one week after anthesis (Poliquit *et al.*, 2007). Gelmesa *et al.*, 2012 reported that application of 2, 4-D at 5 and 10 ppm resulted in reduced plant growth and hastened early flowering and fruiting with concentrate pick harvest. Gemici *et al.*, (2006) that suggested high concentrations of 2, 4-D at 10 ppm produced fewer fruits in tomato. Pandolfini *et al.*, 2002 indicated that synthetic auxin like 2, 4-D has herbicidal or ephynastic effect which lead to flower bud abscission, poor fruit set, fruit defects and puffiness beyond certain concentrations. Gelmesa *et al.*, 2010 indicated that, 2, 4-D beyond certain concentration leads to flower bud abscission and fruit drop due to its herbicidal effect. Gimici *et al.*, (2006) who suggested that high concentrations of 2, 4-D at 10 mg l⁻¹ produced fewer fruits than with 4-CPA. Gelmesa *et al.*, 2012 revealed that the interaction effect of 2, 4-D and GA3 indicated that fruit length was maximum for both levels of 5 and 10 mg l⁻¹ 2, 4-D with 10 mg l⁻¹ of GA3 but significantly reduced when the concentration of GA3 increased. Serrani *et al.*, (2007a) reported that, tomato fruits induced by 2, 4-D had thicker pericarp than pollinated fruits throughout its development, and more in response to 2, 4-D than GA3.

Role of auxin on cell division and cell elongation of tomato

IAA stimulates cell elongation by stimulating wall-loosening factors, such as elastins, to loosen cell walls and the effect is stronger if gibberellins are also present (Bunger-Kibler and Bangerth, 1983). IAA also stimulates cell division if cytokinins are present (Zhao, 2008). IAA induces the formation and organization of phloem and xylem. When the plant is wounded, the IAA may induce the cell differentiation and regeneration of the vascular tissues (Ulmasov *et al.*, 1999).

Role of auxin on root development and apical dominance of tomato

IAA promotes root initiation and induces both growth of pre-existing roots and adventitious root formation, i.e., branching of the roots (Varga and Bruinsma, 1976). As more native auxin is transported down the stem to the roots, the overall development of the roots is stimulated. The longer and branched root can uptake more nutrients from the soil which are accumulated to the plant sink and increase the yield (Wang *et al.*, 2005). If the source of IAA is removed, such as by trimming the tips of stems, the roots are less stimulated accordingly. IAA induces shoot apical dominance and the axillary buds are inhibited by IAA (Woodward and Bartel, 2005). IAA promotes root initiation and induces both growth of pre-existing roots and adventitious root formation, i.e., branching of the roots (Varga and Bruinsma, 1976). Alfonso and Alonso (1981) found that the root proliferation occurred in tomato (cv. Manalucie) shoot segments cultured on Murashige and Skoog media with combination of 0.01, 0.1, 0.5 and 1.0mg/ NAA and 0.01, 0.1, and 0.5 mg/l BA. Higher concentrations of NAA (2 and 4 mg/l) and BA (1, 2 and 4mg/l) inhibited rooting. Singh (1999) observed that the IBA and NAA (250-500 ppm) promoted rooting in tomatoes. Higher net returns were observed in the treatment with IBA at 500 ppm alone or in combination with (250-500 ppm). Taylor and Scheuring (2004) reported that the frequency of lateral root initiation in tomato seedling roots is increased over eightfold in response to 1.6 M-naphthalenacetic acid (NAA).

Role of auxin on growth of tomato

Patel *et al.*, 2012 revealed that application NAA increases the plant height and number of branches in tomato. Abdel Rahman (2008) assessed changes in growth, endogenous

levels of hormones, and ethylene evolution and cellulite and pectolytic enzyme activities of cherry tomato fruits from anthesis through ripening. After anthesis, growth of cherry tomato fruit follows a three – dimensional and sigmoid growth pattern which consists of cell division, cell enlargement and mature green. Pink and red stages. Cytokinins and auxins were abundant and reached their peak during early development (cell division). Ali *et al.*, (2012) reported that IAA produces highest number of branches per plant in tomato. Chhonkar and Singh (1959) reported that high concentration of IAA reduced plant height. Khaled *et al.*, (2015) reported that Plant height, number of leaves and number of branches were significant influenced by the combined application of IAA in BARI tomato 7, Manik and Ratan varieties of tomato. Singh *et al.*, (2005) saw that plant growth and number of branches of tomato positively affected by IAA and NAA. Singh and Upadhyay (1967) observed that NAA 10ppm increased the height and higher doses significantly reduced the height. Mehrotra *et al.*, (1971) reported that NAA 25 ppm had little effects on plant height but there was no effect on number of branches when tomato seedling where treated for 30 minutes before transplanting. It was also reported that quality of fruit was improved with the application of NAA 25 ppm. Patil and Mahajan (1971) noted that higher concentration of NAA 0.4 ppm induced more height in tomato seedling when seedling roots were dipped for 24 hours prior to transplanting, while the average diameter of branches number and leaves were not affected.

Singh *et al.*, (1981) reported that height of the main stem of the plant varied due to the plant growth regulators treatment. IAA and NAA resulted in production of taller plants, maximum number of branches and yield q/ha in tomato. Gupta *et al.*, (2001) recorded significantly maximum plant height at 75

DAT and maximum number of branches at 60 with 75 ppm NAA alone with 2000 ppm NAA as compared to control. Kishan – Swaroop *et al.*, (2001) recorded maximum number of primary branches in the treatment NAA 25 ppm. While the lowest number of primary branches was recorded in the treatment boron 50 ppm. Number of primary branches was not influenced by the growth regulatory substances. Singh *et al.*, (2011) revealed application NAA have positive effect on vegetative growth of three tomato cultivars viz., NUN-1560 (V1), NUN-964 (V2) and NUN-963 (V3). Application of IAA as foliar sprays or to the by moderately high rainfall during Kharif (April growing media of tomato plants had a stimulatory effect September) season and low temperature (15°-20°C) in on plant growth and development (Hathout *et al.*, 1993). Chhonkar and Ghufra (1968) reported that plant height decreased with the increased concentration of NAA concentration.

Hathout *et al.*, (1993) found that application of 10 ppm IAA as foliar sprays or to the growing media of tomato plants had a stimulatory effect on plant growth, development and fruit which was accompanied by increases in endogenous auxin, gibberellins and cytokinin contents. However, IAA at 80 ppm had an inhibitory effect on plant growth and development, which was accompanied by increase in the level and activity of indigenous inhibitors and by low levels of auxins, cytokines and gibberellins. Karim *et al.*, (2015) observed that 4-chlorophenoxy acetic acid (4-CPA) had a significant influence on growth of tomato var. BARI Hybrid Tomato-8. The synthetic auxin 2, 4-D mimics the function of natural auxins which control “a multitude of plant growth and development processes” (Hess, 1993). Patel *et al.*, (2012) revealed that application 2, 4-D increases the plant height and number of branches in tomato. Anwar (2010) indicated that application of 2, 4-D at

5 mg l⁻¹ significantly improved growth attributes of tomato plant but those attributes decreased beyond this concentration.

Role of auxin on yield of tomato

Patel *et al.*, 2012 revealed that application NAA increases the fruit diameter and yield in tomato. Ali *et al.*, (2012) reported that IAA produces highest number of flower per plant and yield in tomato. Application of NAA increases the yield in tomato due to enhanced plant growth and faster rate of plant development by the action of NAA in cell elongation and there by increased cell enlargement, cell division and differentiation which in turn result into increase in number of flowers, fruit set, size and weight of fruit as reported by Rodrigues *et al.*, (2001), Kishan *et al.*, (2001), Rai *et al.*, (2002), Nibhavanti *et al.*, (2004), Singh and Sant (2005) and Bokade (2006). Chhonkar and Singh (1959) recorded increasing yield of tomato by seedling treatment with growth substances. They reported that high concentration of IAA increases yield through increased flower induction and fruit set. Singh and Upadhyaya (1967) studied the effect of IAA and NAA on tomato and reported that the regulators increased the fruit set, size and yield of fruit. The chemicals could be applied on seeds, roots whole plants or flowers, but foliar application was very effective for increasing the size of fruit and the yield. Kaushik *et al.*, (1974) reported that 10 ppm of IAA increased the number and weight of fruits per plant significantly. Khaled *et al.*, (2015) reported that weight tomato-1, yield plant-1, yield plot-1 and yield hectare-1 were significant influenced by the combined application of IAA in BARI tomato 7, Manik and Ratan varieties of tomato. Singh *et al.*, (2005) saw that yield of tomato positively affected by IAA and NAA. Gupta *et al.*, (2003) found highest yield with application of IAA + Multiplex micronutrient mixture at the

maturity stage during. Baliyan *et al.*, (2013) revealed that application of 4-CPA in summer tomato increases the number of tomato fruit set which helps in increasing in fruit yield. Alam and Khan (2002) revealed that reduced pre-harvest fruit drop with increased number of fruits per plant and yield was observed due to Naphthalene Acetic Acid (NAA) or b-NAA spray. Habbasha *et al.*, (1999) found that application of IAA increased fruit set percentage and total fruit yield compared to control. Singh and Upadhyay (1967) observed that NAA 10 ppm as soil and foliar application give increased yield in tomato but NAA 20 ppm significantly reduced yield. It is also observed that the largest fruit size was found with 75 ppm NAA along with multiplex at maturity stage of tomato and gave the maximum yield (Gupta *et al.*, 2003). Patil and Mahajan (1971) noted that NAA 0.1 ppm resulted more weight of fruits per plant and NAA 0.05 ppm as seedling root dipping from 24hrs gave higher percent of yield in fruit and second picking. Perenz Zapata *et al.*, (1980) found that tomato Cvs. Floral and Marglobe, treated with NAA at 25 and 35 ppm had large fruit size. Singh *et al.*, (1981) reported that IAA and NAA resulted highest yield q/ha in tomato. Alam and Khan (2002) reported that spray of NAA at variable concentration significantly increased the fruits yield of tomato as compared to control. Gupta *et al.*, (2003) reported that the application of 75ppm NAA along with multiplex resulted in largest fruit size at maturity stage of tomato and gave maximum yield. Singh *et al.*, (2011) revealed application NAA have positive effect on yield of three tomato cultivars viz., NUN-1560 (V1), NUN-964 (V2) and NUN-963 (V3). Arvind (2012) reported that combination of variety “NBH NO-1” and 15 ppm NAA was found best in respect increasing productivity of tomato crop. Karim *et al.*, (2015) observed that 4-chlorophenoxy acetic acid (4-CPA) had a significant influence on yield of tomato var. BARI

Hybrid Tomato-8. Gemici *et al.*, (2006) indicated increased fruit size and setting in tomato due to application of 2, 4-dichlorophenoxy acetic acid (2, 4-D), 4-chlorophenoxy acetic acid (4-CPA) and β -naphthoxy acetic acid (β -NAA). The synthetic auxin 2, 4-D mimics the function of natural auxins which control “a multitude of plant growth and development processes” (Hess, 1993). Patel *et al.*, (2012) revealed that application 2, 4-D increases the fruit diameter and yield in tomato. Gelmesa *et al.*, (2010) reported that application of 2, 4-D at or lower concentration of 5 mg l⁻¹ improves the fruit yield and quality of tomato variety Fetan. Anwar (2010) indicated that application of 2, 4-D at 5 mg l⁻¹ significantly improved fruit yield of tomato plant but those attributes decreased beyond this concentration. Pudir and Yadav (2001) that indicated improvement in tomato fruit yield at low concentration (5 mg l⁻¹) of 2, 4-D. Khan *et al.*, (2006) stated that 2, 4-D increases longer fruits with bigger size in tomato which could be due to stimulation of parthenocarpic fruit growth that resulted in increased fruit weight. Gemici *et al.*, (2006) also reported that 2, 4-D resulted in increased tomato fruit size, fresh and dry weight when used at recommended concentration.

Role of auxin on quality of tomato

Patel *et al.*, (2012) revealed that application NAA increases the acidity and TSS in tomato. Perez and Ramirez (1980) carried out an experiment with the application of IAA at 25 and 35 ppm on tomato. They found increased fruit size quality with minimum seeds. Gupta *et al.*, (2003) found the largest fruit size, most attractive ripe fruit color highest dry matter and ash content with application of IAA + Multiplex micronutrient mixture at the maturity stage during. NAA application in tomato increased total soluble solid percentage significantly (Pudir and Yadav,

2001). Habbasha *et al.*, (1999) found that application of IAA reduces the percentage of puffy and parthanocarpic fruit and compared to control. Mehrotra *et al.*, (1971) reported that quality of fruit was improved with the application of NAA 25 ppm. Petronk and Loban (1975) obtained quality of tomato seed by treating the seed with IAA at 100 mg/kg or NAA at 50mg/kg before sowing. Pandita *et al.*, (1978) found higher fruit acidity in tomato plant treated with NAA 100 ppm as foliar spray at appearances of first flowering. The highest vitamin „C“ content was in fruit from the plant treated with NAA 50 ppm.

The beneficial effect of NAA at 100 ppm on fruit T.S.S. was also observed. Alam and Khan (2002) reported that spray of NAA at variable concentration significantly increased the nutrient contents of fruits. Rai *et al.*, (2002) reported that maximum chlorophyll content and acidity were obtained with NAA at 75 ppm along with Humaur at 2000 ppm. NAA at 75 ppm along with Multiplex at 2500 ppm gave the highest sugar content. Singh *et al.*, (2011) revealed application NAA have positive effect on quality of three tomato cultivars viz., NUN-1560 (V1), NUN-964 (V2) and NUN-963 (V3). Thakur *et al.*, (1996) indicated that the ascorbic acid content increased with higher concentrations of 2, 4-D and Para-chlorophenoxy acetic acid.

From this review it can be clearly inferred that application of 10 ppm IAA, 50 ppm PCPA, 20 ppm NAA or 5ppm 2, 4-D as foliar sprays had a stimulatory effect on plant growth, flowering and, fruit setting, yield and quality of fruit which was accompanied by increases in endogenous auxin, gibberellins and cytokinin contents in tomato plant. So applying of both natural and synthetic auxin helps farmer in cultivating tomato in adverse climatic condition which can give good fruit yield by increasing vegetative and reproductive growth and reducing the flower

and fruit drop.

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