

Original Research Article

<https://doi.org/10.20546/ijcmas.2017.611.017>

Standardization of Isolation Technique of *Rhizobium* from Root Nodules of Lentil

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ABSTRACT

Native plant growth promoting rhizobacteria are unable to compete with other bacterial strains and hence cannot express their full potential in realising the beneficial effects. Hence, inoculation of plants with target microorganism is necessary to exploit full potential for yield enhancement. This study was aimed at standardizing the technique for isolation of *Rhizobium* species from root nodules of lentil so as to explore and maximize their contributions to soil nitrogen fertilization in place of synthetic fertilizers. A total of five nodulated healthy plants collected from North-eastern part of Uttar Pradesh, India were sterilized and macerated under aseptic condition followed by streaking one loop full suspension in a zig-zag manner using the inoculation loop on YEMA medium in laminar air flow and incubated in a B.O.D. incubator at 28±2°C for 24 hours. Growth on YEMA plate was observed after the said incubation period and then maintained at 4°C in 20% (w/v) glycerol at -80°C for long-term storage. Vigorous growth of bacteria was seen in next 24 to 48 hours. Bacterial colonies were found to have distinct colour, morphology, appearance and texture on agar plates.

Keywords

Rhizobacteria,
YEMA, PGPR,
Agar, Lentil.

Article Info

Accepted:

04 September 2017

Available Online:

10 November 2017

Introduction

Lentil (*Lens culinaris* L.), a member of leguminosae family, having origin in Middle East and Egypt, later spreaded to Latin America (Cubero, 1981 and Duke, 1981) is one of the oldest cultivated legume plants (Bahl *et al.*, 1993 and Rehman *et al.*, 1994), grown commercially for its edible seeds. It is a self-pollinating, annual cool season crop with disomic chromosome number of 14 (Muehlbauer, 1991) with a haploid genome size of an estimated 4063 Mbp (Arumuganathan and Earle, 1991). It can be

grown to an altitude of 3000 meters above mean sea level. However, seed yield decreases with increasing altitude (Whyte *et al.*, 1953).

The total cultivated area in the world accounts around 4.6 million hectares producing 4.2 million tons of seeds with an average productivity of 1095 kg/ha (FAO, 2010). India accounts the largest global area with 1.48 million ha and 1.01 million ha production (AICRP, 2010-2011).

Yield levels can be enhanced by efficient use of synthetic fertilizers. However on account of increasing environmental degradation, synthetic fertilizers need to be replaced by a more effective, sustainable and cost-effective method through the use of biofertilizer. Plant Growth Promoting Rhizobacteria (PGPR) residing in rhizosphere in association with plant root surface has positive effects on growth and yield of different crops in a sustainable manner. These beneficial microorganisms are important components of plant productivity and soil fertility because of their participation in several key processes by release of different plant growth regulators (like IAA, GA *etc.*), by solubilising inorganic phosphates, by biological nitrogen fixation, control of plant pathogens *via* synthesis of antibiotics, by nutrient cycling and seedling establishment (Jeffries *et al.*, 2003 and Rosas *et al.*, 2009). They have been also reported to confer upon the plants ability to tolerate different abiotic stresses (Yang *et al.*, 2009 and Lim and Kim, 2013). Native plant growth promoting rhizobacteria are unable to compete with other bacterial strains and hence cannot express their full potential in realising the beneficial effects. Hence, inoculation of plants with target microorganism is necessary to exploit full potential for yield enhancement. This study was aimed at standardizing the isolation of *Rhizobium* species from root nodules of lentil so as to explore and maximize their contributions to soil nitrogen fertilization in place of synthetic fertilizers.

Materials and Methods

A total of five nodulated plants were collected from North-eastern part of Uttar Pradesh, India. Healthy plants were uprooted carefully and those plants possessing healthy nodules with pink colour were selected and transported to the P.G.P.R. laboratory, Department of Genetics and Plant Breeding, Institute of Agricultural Sciences, Banaras Hindu University Varanasi, India in polythene bags for immediate processing. Yeast extract mannitol agar (YEMA) with pH range of 6.8-7.0 was prepared before *Rhizobium* isolation as described by Rajendran *et al.*, In this, healthy, unbroken, firm and pink nodules were selected and washed under tap water to remove adhering mud and soil particles, followed by treating it carefully with 5% hydrogen peroxide for surface sterilization. Repeated washing of nodules in sterile water for 3- 4mins was made to get rid of the sterilant and then treated with 70% ethyl alcohol for about one minute and 0.1% HgCl₂ for two minutes.

Again they were washed with sterile water (3 successive times) under aseptic conditions and crushed with sterile crucible. A suspension made of the crushed nodules, streaked one loop full suspension in a zig-zag manner using the inoculation loop on YEMA medium under aseptic condition in laminar air flow and incubated in a B.O.D. incubator (Narang Scientific, India) at 28±2 °C (Vincent, 1970) for 24 hours.

Table.1 Selected rhizobial strains and their sources

Strains	Source
LR1	North-eastern part of Uttar Pradesh, India
LR 2	
LR 3	
LR 4	
LR 5	

Growth on YEMA plate was observed after the said incubation period and then maintained at 4°C in 20% (w/v) glycerol at -80°C for long-term storage. Glass vessels containing nutrient media were sterilized and autoclaved at 15 lbs square⁻¹ inch pressure at 121°C for 15 minutes. Double distilled water was used throughout the experiment for preparation of media, reagents *etc.*

Results and Discussion

Bacterial colonies of *Rhizobium* strains (Table 1) isolated from the samples collected from regions of north-eastern part of Uttar Pradesh, India were visible on agar plates after 12 hours of inoculation. Vigorous growth of bacteria was seen in next 24 to 48 hours. Bacterial colonies were found to have distinct colour, morphology, appearance and texture on agar plates. Further bacterial colonies were transferred in the bakelite tube having YEMA media through streaking for medium term storage. Interestingly these isolates could be used in future for further testing the biochemical, physiological, morphological characteristics of bacteria as well as their potential for providing the crop better growth in biotic and abiotic stress conditions.

References

- All India Coordinated Research Project 2010–11 Project coordinator's report on Mullarp, rabi (2009–2010). Indian Institute of Pulses Research Kanpur, India.
- Arumuganathan, K., and Earle ED 1991. Nuclear DNA content of some important plant species. *Pl Mol Biol Rep* 9: 208-218.
- Bahl, P.N., Lal S and Sharma BM 1993. An overview of the production and problems in Southeast Asia. In W Erskine and M C Saxena (Eds.), *Lentil in South Asia*. Proceedings of the seminar on lentils in south Asia 1-10 ICARDA, Aleppo, Syria.
- Cubero, J.I., 1981. Origin, taxonomy and domestication. *Lentils Slough*, CAB 15-38.
- Duke, J.A., 1981. *Handbook of legumes of world economic importance*. Plenum Press, New York. Pp. 52-57.
- FAO, 2010. Faostat, Fao Statistical database. <http://www.fao.org>
- Jeffries, P., Gianinazzi S, Perotto S, Turanu K and Barea M 2003. The contribution of arbuscular mycorrhizal fungi in sustainable maintenance of plant health and soil fertility. *Bio and Fert of Soils* 37: 1-16.
- Lim Jong-Hiu, and Kim Sang-Dal 2013. Induction of Drought Stress Resistance by Multi-Functional PGPR *Bacillus licheniformis* K11 in Pepper. *Pl Pathol J* 29(2): 201-208.
- Rajendran, G., Singh F, Desai AJ, and Archana G 2008. Enhanced growth and nodulation of pigeon pea by co-inoculation of *Bacillus* strains along with *Rhizobium* sp. *Biores Techno* 99: 4544–4550.
- Rehman, S., and Altaf CHM 1994. Karyotypic studies in *Lens culinaris* Medic, ssp. *Macrosperma* cv. Laird X Precoz. *Pak J Bot* 26(2): 347-352.
- Rejili, M., Mahdhi M, and Fterich A 2012. Symbiotic nitrogen fixation of wild legumes in Tunisia: Soil fertility dynamics, field nodulation and nodules effectiveness. *Agr Ecosyst Environ* 157:60–69.
- Whyte, R.O., Leissner, GN and Trumble HC 1953. *Legume in agriculture*. FAO Agricultural Studies 21: 323-325.
- Yang, R.C., Crosa J, Cornelius PL, Burgueno J Biplot 2009. Analysis of genotype environment interaction: proceed with caution. *Crop science* 49:1563- 1576.

How to cite this article:

Ajay Varun, Arun Patel, Umakant Banjare, Santosh Kumar and Ramesh Kumar Singh. 2017. Standardization of Isolation Technique of *Rhizobium* from Root Nodules of Lentil. *Int.J.Curr.Microbiol.App.Sci*. 6(11): 132-134. doi: <https://doi.org/10.20546/ijcmas.2017.611.017>