

Original Research Article

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Evaluation of Botanical Extracts against *Pyricularia oryzae*, A Causal Agent of Blast of Paddy

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ABSTRACT

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Rice is one of staple food of India and is only second most consumed cereal crop around the world. Blast of paddy is one of the economically important diseases caused by the phytopathogen *Pyricularia oryzae*. The present investigation carried out to evaluate the potentiality of extract of locally available botanicals namely, *Catharanthus roseus*, *Duranta repens*, *Solanum nigrum*, *Crotalaria trichotoma*, *Oxalis latifolia*, *Leucas aspera* and *Parthenium hysterophorus* against *Pyricularia oryzae* *in-vitro*. The experiment was carried out using 5% aqueous extracts of botanicals by poisoned food technique. Out of the seven tested aqueous botanical extracts *D. repens* has shown highly significant results with 100% growth inhibition of pathogen which is on par with fungicide mancozeb, followed by *C. roseus* (31.36%), *S. nigrum* (30.76%), *C. trichotoma* (25.44%), *O. latifolia* (24.14%) and least inhibition was observed in *L. aspera* (15.38%) and *P. hysterophorus* (10.05%).

Introduction

Oryza sativa belongs to the family Gramineae (Poaceae), is one of the major food crops of the world and forms the staple diet of half of the world's population. Asia is the leader in rice production accounting for about 90% of the world's production. Globally, India stands first in the area covered in cultivation and second in production after China (FAOSTAT, 2021). With the current status of the production of rice the demand projected to be increases 30 % by 2050 (Mohanty and Chavez, 2010; FAO-UN, 2015). Rice is prone to several diseases which in turn reduces yield of the crop and affects the economy of farmers specially in developing countries. The crop is susceptible to many viral, bacterial and fungal diseases.

Blast of paddy caused by the phytopathogen *Pyricularia oryzae*, is an economically important and widely distributed phytopathogen throughout the world and it occurs in mild to severe form in different parts of India.

Rice blast is one of the most catastrophic diseases which occur on all above-ground parts of the rice plant. Rice blast caused by *Pyricularia oryzae* was firstly described by Cavara in 1891, which attack the leaf sheath, rachis, the joints of the culm, and even the glumes. Infection to the culms causes greater damage than leaf infection. Severe leaf infection occurring in the post-transplant stage may, however, lead to total destruction of the foliage (Ou, 1985). Natural plant products and their analogous are an important source of new agricultural

chemicals which are gaining more importance in the research area and are used as organic methods in control of plant diseases.

There are many reports on use of botanicals in managing plant diseases both *in vitro* and *in vivo*. Botanicals like *Azadirachta indica*, *Solanum torvum*, *Hyssopus officinalis*, *Crotalaria trichotoma*, *Citrus aurantifolia*, *Asafoetida* sp. and *Pongamia* sp. *Melia azadirachta*, *Allium sativum*, *Allium cepa*, *Curcuma longa*, *Caryophyllus aromaticus* are known to be effective in controlling phytopathogens like *Pyricularia oryzae*, *Alternaria alternata*, *Bipolarize oryzae*, *Curvalaria lunata*, *Fusarium oxysporum*, *Fusarium moniliformae*, *Fusarium solani*, *Xanthomonas oryzae*, *Pyrenophora avenae*, *Alternaria solani*, *Rhizoctonia solani*, *Alternaria trititina* (Amadioha, 2000; Letessier *et al.*, 2001; Shervin, 2012; Pal and Kumar, 2013; Ravikumar and Rajkumar, 2013; Chakrapani *et al.*, 2020 and Puja *et al.*, 2021).

The present investigation focused on evaluating the efficacy of locally available botanicals in managing blast of paddy *in vitro*, which is an effort to demonstrate ecofriendly approach in plant disease management by using locally available resources and also cost effective compared to synthetic fungicide.

Materials and Methods

Collection and Preparation of plant extracts

Botanicals were selected on the basis of three criteria i.e., (i) Presence of antimicrobial toxins. (ii) Presence of pre-infection defense bio-chemicals which provide resistance to plants against pathogens. (iii) Availability of plants materials throughout the year with very little or no commercial value as compared to their chemical alternatives.

Such seven plants with family and plant parts used are listed in Table 1. Fifteen grams of each selected seven plants was macerated with 30 mL of distilled water in mortar and pestle.

The macerated biomass was kept overnight in culture tubes for exudation of biochemicals. The biomass was filtered through muslin cloth, followed by through Whatman No. 1 filter paper. The filtered extracts were stored at 4°C as 50% stock solutions for further experiment.

Determination of efficacy of plant extracts against *Pyricularia oryzae* by poison food technique

Efficacy of plant extracts on the growth of *Pyricularia oryzae in vitro* was determined by poison food technique. Plant extract of 2mL from the each stock solution (50% concentration) was added to 18 mL of sterilized potato dextrose agar (PDA) medium in a conical flask to make final concentration of 5%, mixed thoroughly, and autoclaved. A 5 mm diameter disc of actively growing 6–7 days old culture of *P. oryzae* was placed in the centre of the Petri plates. The Petri plates containing medium with 0.2% mancozeb fungicide (Indofil® mancozeb 75% WP) served as positive control and plates with medium amended with 2mL distilled water served as negative control. Three replicates were maintained for each treatment. Radial growth of mycelium was measured after seven days of incubation. The results were compared with negative control. Percentage of inhibition was calculated with the following formula (Singh and Tripathi, 1999).

$$\text{Percentage of inhibition (\%)} = \frac{dc - dt}{dc} \times 100$$

Where, dc = Mycelia growth in control

dt = Mycelia growth in treatment

Results and Discussion

In the present investigation seven aqueous extracts of the selected botanicals (Table 1) used to assess their efficacy against phytopathogen *P. oryzae*. Among the seven botanicals used, *D. repens* showed highly significant activity with complete inhibition (100%) of the mycelial growth of the pathogen at 5% of concentration which is on par with synthetic chemical fungicide, followed by *C. roseus* (31.36%), *S. nigrum* (30.76%), *C. trichotoma* (25.44%), *O. latifolia* (24.14%), *L. aspera* (15.38%) and *P. hysterophorus* (10.05%) (Table 2 and Fig. 1 and 2) which corroborates with earlier reports that plant extracts of different families have shown inhibitory activity against many fungal pathogens (Choi *et al.*, 2004; Sohbat *et al.*, 2014; Puja *et al.*, 2021; Julián *et al.*, 2015; Madhu *et al.*, 2014; Manasi and Ajit, 2014; Pandukur and Amienyo, 2016; Malarkodi and Manoharan, 2013; Meera and Balabaskar, 2012; Mehajabeen, 2011; Bolatito *et al.*, 2023).

Table.1 Name of Botanicals and parts used for aqueous extraction

SL. No.	Scientific name	Family	Plant part used for extraction
1	<i>Crotalaria trichotoma</i> Linn.	Leguminosae	Leaves
2	<i>Leucas aspera</i>	Lamiaceae	Leaves
3	<i>Duranta repens</i> Linn.	Verbenaceae	Leaves
4	<i>Parthenium hysterophorus</i> Linn.	Asteraceae	Leaves
5	<i>Catharanthus roseus</i> Linn.	Apocynaceae.	Leaves
6	<i>Solanum nigrum</i> Linn.	Solanaceae	Leaves
7	<i>Oxalis latifolia</i>	Oxalidaceae	Leaves

Figure.1 Inhibitory activity of Botanical aqueous extracts at 5% concentration compared to Chemical (0.2%) and Distilled water

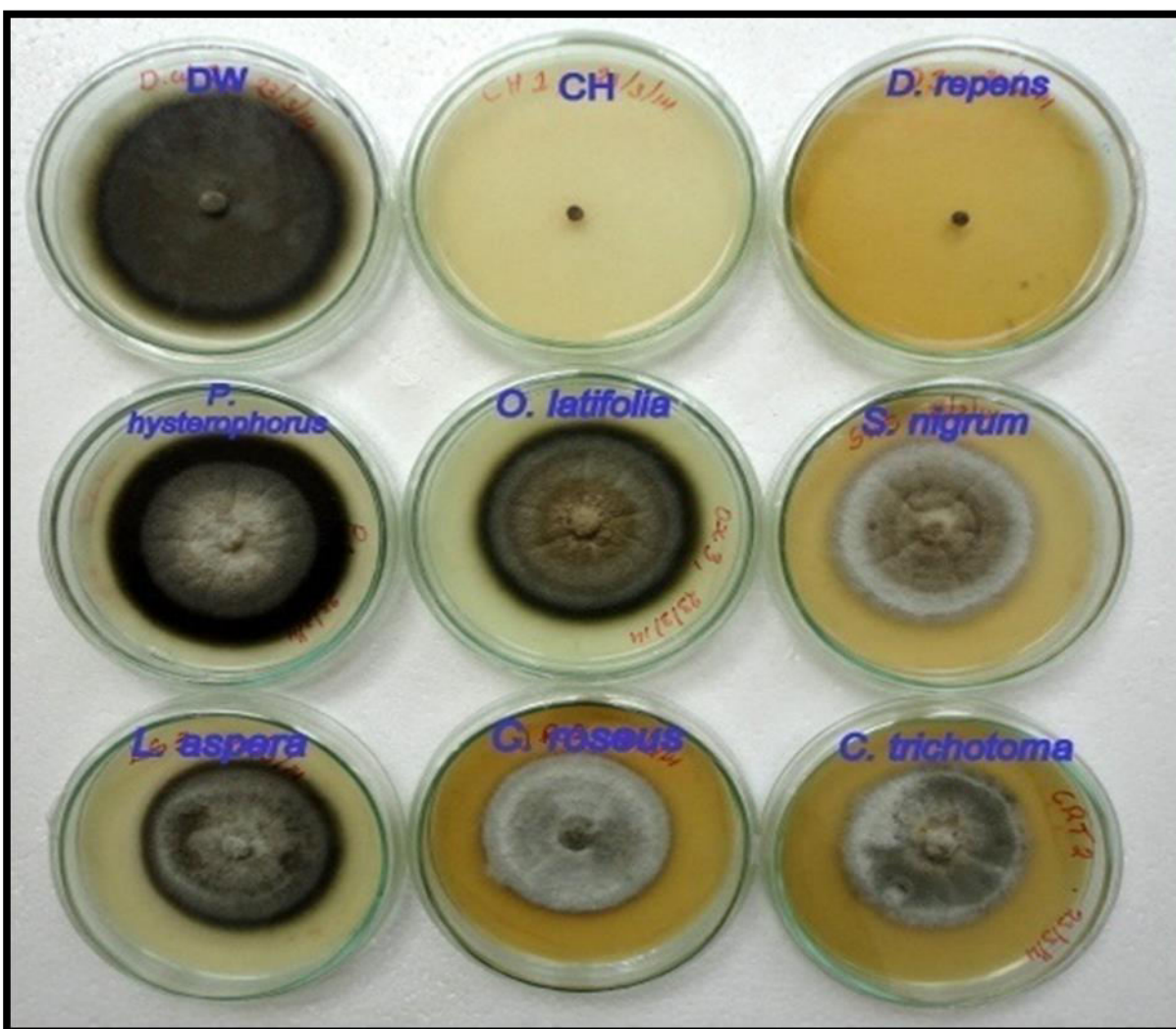
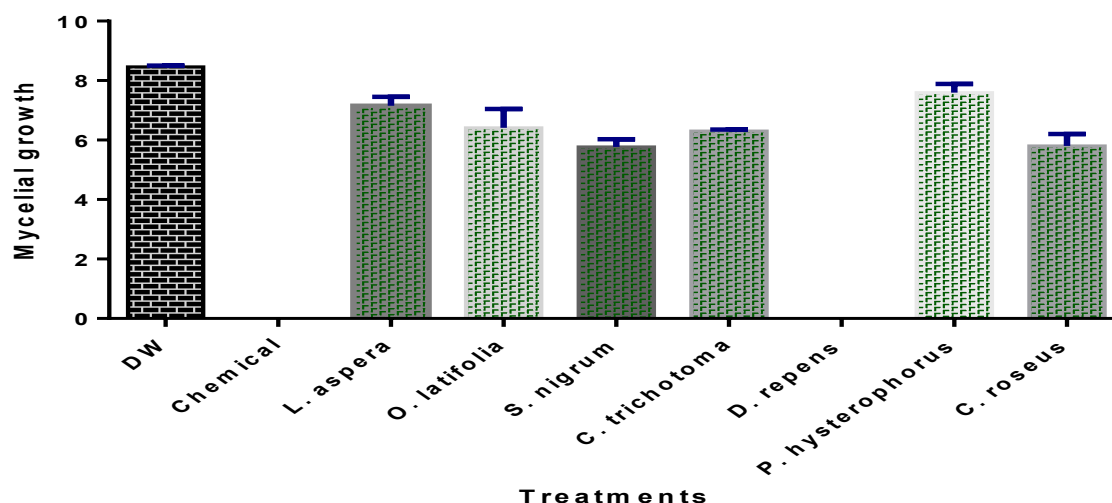


Table.2 Inhibitory activity of aqueous plant extracts

Sl.No.	Treatment	Treatment of aqueous extract in %	Colony diameter (in cm)	% of Mycelial inhibition
1	Distilled water	5	8.45 ± 0.033	0
2	Chemical (Mancozeb)	0.2 [#]	0.0 ± 0.0***	100 %
3	<i>Leucas aspera</i>	5	7.15 ± 0.30	15.38 %
4	<i>Parthenium hysterophorus</i>	5	7.6 ± 0.39	10.05 %
5	<i>Oxalis latifolia</i>	5	6.41 ± 0.62*	24.14 %
6	<i>Crotalaria trichotoma</i>	5	6.3 ± 0.05*	25.45 %
7	<i>Catharanthus roseus</i>	5	5.8 ± 0.45**	31.36 %
8	<i>Duranta repens</i>	5	0.0 ± 0.0***	100 %
9	<i>Solanum nigrum</i>	5	5.85 ± 0.31*	30.76 %

#Recommended dose

*** Data was analyzed by Tukey's post-test at $P \leq 0.05$ level of significance using Graph pad prism 5 software**Figure.2** Graphical representation of Mycelial inhibition of *Pyricularia oryzae* by various treatments

Control of crop diseases and managing phytopathogens which are responsible for causing diseases is one of the most economically challenging tasks to both farmers and scientists. As the diseases of the crop plants progressed, the research over such diseases and study of causal agents responsible also increased which resulted in discovery of chemical fungicide to control and manage the disease caused by phytopathogens. However, overuse of chemical fungicides eventually led to harmful effects on humans and other animals (Sanjay *et al.*, 2018; Jochen *et al.*, 2019). This forced the researchers to find alternative approaches in managing plant diseases and inclined towards natural botanicals to find the antimicrobial substances.

The Aqueous extract of *D. repens* with numerous biochemical compounds was significantly effective which is on par with fungicide. The aqueous extracts of *D. repens* with significant activity against tested pathogen *P. oryzae* proved that, even crude extracts of botanicals are capable of controlling the pathogens equivalent to purified chemical fungicides.

Hubert *et al.*, (2015) also found that plant extracts of *Coffee arabica*, *Nicotiana tabacum*, *Aloe vera* and *Azadirachta indica* were effective in mycelial inhibition of *Pyricularia grisea*. The similar results were obtained with aqueous plant extracts of *Azadirachta indica*, *Aloe vera*, *Nicotiana tabacum*, against *Magnaporthe oryzae*

as reported by Falade and Asowata-Ayodele (2022). Based on the results from the present findings it is found that biological control of phytopathogens and diseases is becoming more and more evident and gaining more importance as they are not hazardous to the farmers, soil microbial flora and fauna. Thus from the present findings *D. repens* can be used in control of *P. oryzae*.

Further, *D. repens* could be developed as an alternative bio-formulation to fungicide which can be used directly by the farmers with proper guidelines and hence can reduce the dependency on fungicides reducing the contamination of soil.

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Author Contributions

M. C. Ravikumar: Investigation, formal analysis, writing—original draft. G. S. Nandeesha: Validation, methodology, writing—reviewing. Rajkumar H. Garampalli:—Formal analysis, writing—review and editing.

Data Availability

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethical Approval Not applicable.

Consent to Participate Not applicable.

Consent to Publish Not applicable.

Conflict of Interest The authors declare no competing interests.

References

Amadioha, A C (2000). Controlling rice blast in vitro and in vivo with extracts of *Azadirachta indic* Crop protection. 19: 287-290.

- [http://dx.doi.org/10.1016/S0261-2194\(99\)00080-0](http://dx.doi.org/10.1016/S0261-2194(99)00080-0)
- Bolatito, A S O., Fatoki, O A., Oluwalana, O A., Adeniji, A A (2023). Antifungal Activities of *Moringa oleifera* and *Ageratum conyzoides* against Fungi Causing Deterioration of Plantain (*Musa paradisiaca* L.). Asian Journal of Research in Botany. 6(2): 269-278
- Cavara, F (1891). Fungi *Longobardiae exsiccati* sive mycetum specimina in *Longobardia collecta*, exsiccata et speciebus novis vel criticis, iconibus illustrate. *Pugillus I no 49* (Cited in Padwick GW. 1950 p. 18)
- Chakrapani, K., Bireswar, S W., Tampakleima, C., Tusi, C., Tokmem, S (2020). Assessing *in vitro* antifungal activity of plant extracts against *Rhizoctonia solani* causing sheath blight of rice (*Oryza sativa*. L). *Journal of Pharmacognosy and Phytochemistry* 9(1): 1497-1501.
- Choi, G J., Jang K S., Kim, J S., Lee, S W., Cho, J Y., Cho, K Y., Kim, J C (2004). In vivo antifungal activities of 57 plant Extracts against six plant pathogenic fungi. *The Plant Pathology Journal* 3:184-191. <https://doi.org/10.5423/PPJ.2004.20.3.184>
- Falade, M J. and Asowata-Ayodele, A M (2022). Multi-seasonal evaluation of selected aqueous plant extracts for antimicrobial activities against rice blast disease. *International Journal of Plant Pathology and Microbiology*. 2(2): 38-45
- FAO. 2015. FAOSTAT Database. Available online at <http://faostat.fao.org/>
- FAO. 2021. FAO statistical data, <https://www.fao.org/faostat/en/#data/QCL>
- Hubert, J., Mabagala, R B., Mamiro, D P (2015). Efficacy of Selected Plant Extracts against *Pyricularia grisea*, Causal Agent of Rice Blast Disease. *American Journal of Plant Sciences*. 6: 602-611. <https://doi.org/10.4236/ajps.2015.65065>
- Jochen, P Z., Mirco, B., Gertie, A., Carsten, A B., Gwenael, I., Anja, K., Sylvain, P., Jes, J., Rasmussen, J R., Andreas, S., Kelly, S., Sebastian, S., Ralf, A., Ralf, B S (2019). Fungicides: An Overlooked Pesticide Class? *Environmental Science and Technology* 53: 3347–3365 <https://doi.org/10.1021/acs.est.8b04392>
- Julian, R A., Jens, C S., Hilmer, S., Nguyen, D K., David, B C., Hans, J L J (2015). Activity-guided separation of *Chromolaena odorata* leaf extract reveals fractions with rice disease-reducing properties. *European Journal of Plant Pathology*. 143:331–341. <https://doi.org/10.1007/s10658-015->

0684-x

- Letessier, M P., Svoboda, K P., Walters, D R (2001). Antifungal Activity of the Essential Oil of Hyssop (*Hyssopus officinalis*). *Journal of Phytopathology* 149: 673–678. <https://doi.org/10.1046/j.1439-0434.2001.00692.x>
- Madhu, G., Sushil, S., Rekha, B (2014). Fungitoxic activity of fruit extracts of *Syzygium cumini* (L.) Skeels against plant pathogenic fungi *Alternaria alternata* and *Fusarium oxysporum*. *Archives of Phytopathology and Plant Protection* 48(4): 354-364. <https://doi.org/10.1080/03235408.2014.888875>
- Malarkodi, E. and Manoharan, A (2013). Antifungal activity of *parthenium hysterophorus*. *J Chemical and Pharmaceutical Research* 5(1): 137-139.
- Manasi, K B. and Ajit, G D (2014). Antifungal activity of herbal extracts against plant pathogenic fungi. *Archives of Phytopathology and Plant Protection* 47(8): 959-965. <https://doi.org/10.1080/03235408.2013.826857>
- Meera, T. and Balabaskar, P (2012). Antifungal activity of botanicals against *Sarocladium oryzae* causing rice sheath rot disease. *International Journal of Food Agriculture and Veterinary Sciences* 2(1): 121-127.
- Mehajabeen, A M., Jahan, N., Zia-ul-haq, M., Alam, S M., Wazir, A., Hassan, S (2011). Antimicrobial screening of some plants of medicinal importance. *Pakistan journal of Botany* 43(3): 1773-1775.
- Mohanty, S., Wailes, E., Chavez, E (2010). The Global Rice Supply and Demand Outlook: The Need for Greater Productivity Growth to Keep Rice Affordable. In: *Rice in the Global Economy: Strategic Research and Policy Issues for Food Security*. S Pandey D, D Byerlee, D Dawe, A Dobermann, S Mohanty, S Rozelle, and B Hardy(Eds). International Rice Research Institute, Philippines, Manila.
- Ou, S H (1985). *Rice disease*, 2nd Edition Commonwealth Mycological Institute. Kew.
- Pal, G K. and Kumar, B (2013). Antifungal activity of some common weed extracts against wilt causing fungi, *Fusarium oxysporum*. *Current Discovery* 2(1): 62-67.
- Pandukur, S G. and Amienyo, C A (2016). Effect of *Azadirachta indica* extract on the radial growth of some test fungi isolated from two varieties of cocoyam (*Colocasia esculenta* L.) corms and cormels in some markets in Plateau State, Nigeria. *Journal of Phytopathology and Pest Management*. 3(1): 46-59 <https://ppmj.net/index.php/ppmj/article/view/39>
- Puja, K., Sunil, K C., Prabhat, K (2021). Comparative study of mushrooms and botanicals extract against *Alternaria trititica* causing leaf blight disease of wheat. *Journal of Mycology and Plant Pathology*. 51(2): 2021
- Ravikumar, M C. and Rajkumar, H G (2013). Antifungal activity of plants extracts against *Alternaria solani*, the causal agent of early blight of tomato. *Archives of Phytopathology and Plant Protection*. 46 (16): 1897-1903.
- Sanjay, K G., Vineeta, S., Hillol, C., Prassan, C (2018). Harmful Effects of Fungicides-Current Status. *International Journal of Agriculture Environment and Biotechnology*. 779: 1025-1033
- Shervin, H (2012). Antifungal Activity of Some Plant Extracts against Some Plant Pathogenic Fungi in Iran. *Asian Journal of Experimental Biological Sciences*. 3(4):714-718
- Singh, J. and Tripathi, N N (1999). Inhibition of storage fungi of blackgram (*Vigna mungo* L.) by some essential Oils. *Flavour Fragrance Journal*. 14: 1-4 [https://doi.org/10.1002/\(SICI\)1099-1026\(199901/02\)14:1<1::AID-FFJ735>3.0.CO;2-R](https://doi.org/10.1002/(SICI)1099-1026(199901/02)14:1<1::AID-FFJ735>3.0.CO;2-R)
- Sohbat, B., Reza, A., Saeed, A (2014). Anti-fungal properties of 43 plant species against *Alternaria solani* and *Botrytis cinerea*. *Archives of Phytopathology and Plant Protection*. 48(4): 336-344. <https://doi.org/10.1080/03235408.2014.888236>

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