

Original Research Article

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## Antibacterial Activities of Endophytic Fungi Liquid Cultures from *Michelia champaca* L. Plant

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### ABSTRACT

Endophytic fungi can produce secondary metabolites containing antibacterial compound. Some species of endophytic fungi have been isolated from *Michelia champaca* L. plant, those are: *Colletotrichum alienum*, *Geotrichum candidum*, *Rhizoctonia sp*, *Aspergillus ochraceus*, *Mycellia sterilia 1*, *Papulosporasp*, *Mycellia sterilia 2*, *Mycellia sterilia 3*, *Curvularia lunata*, *Colletotrichum kahawae*, dan *Aspergillus parasiticus*. The aims of this research are: 1) To examine the antibacterial activity of each endophytic fungi species against *Escherichia coli*, *Bacillus subtilis*, and *Propioni bacterium acnes*, 2) to determine the endophytic fungi species that has the highest antibacterial activity. Each endophytic fungi isolate is inoculated on Potato Dextrose Agar plate medium, and incubated in 27°C during 7x24 hours, then cut with a sterile scalpel and inoculated into Potato Dextrose Broth (PDB) medium and shaken at 120 rpm for 7x24 hours. Then the liquid culture was centrifuged at 3000 rpm for 10 minutes. Furthermore, the supernatant from each endophytic fungi was treated to *E.coli*, *B. subtilis* and *P. acnes* to examine the antibacterial activity by agar diffusion method. The research results showed that: 1) each species of endophytic fungi have antibacterial activity against *E.coli*, *B. subtilis* and *P. acnes*, 2) the *Rhizoctonia sp*. has the highest antibacterial activity against *E. coli*, *Aspergillus parasiticus* has the highest antibacterial activity towards *B. subtilis*, and *Curvularia lunata* has the highest antibacterial activity towards *P. acnes*.

#### Keywords

Endophytic fungi,  
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### Introduction

Some bacteria species have been resistant towards some antibiotics. Some studies have been done to find eco-friendly natural antibiotics. Some medicinal plants can produce secondary metabolites, i.e:

flavonoid, alkaloid, tannin which are potential to be used as a source of antibiotics (Ciocan and Bara, 2007). Medicinal plant parts i.e: rhizome, leaves, twig, bark, and flower petal could be use in this research. Most of the medicinal plants live in a symbiotic mutualism with endophytic fungi that live

in their host plant tissues (Nair and Padmavathy, 2014). The endophytic fungi are able to produce antimicrobial secondary metabolites, that's why they can protect the host plants by inhibit the plant from microbial attack that cause diseases (Mwanga *et al.*, 2019; Mousa and Raizada, 2013). That's why some medicinal plants could stay healthy and are rarely attacked by microbes that cause diseases. The results of the preliminary study showed that there were 11 species of endophytic fungi isolated from *Michelia champaca* L. Each endophytic fungi species are able to produce: alkaloid, flavonoid, tannin, saponin, and terpenoid with different content. The five secondary metabolites could be extracted from the leaves, twigs, and flower petal. It is important to examine the antibacterial activity of 11 the endophytic fungi species liquid cultures towards *E.coli*, *B.subtilis*, and *P.acnes*. If the research result could prove that antibacterial potential of each endophytic fungi liquid cultures from *Michelia champaca* L. plant parts, so it can be used as an alternative for using the endophytic fungi liquid culture for eco-friendly natural antibiotic.

This research was aimed to: 1) To examine the antibacterial activity of each endophytic fungi liquid cultures species against *Escherichia coli*, *Bacillus subtilis*, *Propioni bacterium acnes*, 2) To determine the endophytic fungi species that has the highest antibacterial activity.

## Materials and Methods

Materials : endophytic fungi isolates (*Colletotrichum alienum*, *Geotrichum candidum*, *Rhizoctonia sp.*, *Aspergillus ochraceus*, *Mycellia sterilia* 1, *Papulospora sp.*, *Mycellia sterilia* 2, *Mycellia sterilia* 3, *Curvularia lunata*, *Colletotrichum kahawae*, and *Aspergillus parasiticus*), cork borer, Potato Dextrose Agar (PDA) medium from MERCK brand, Potato Dextrose Broth (PDB) medium (consist of potato tuber essence, dextrose from MERCK and aquadest), Nutrient Broth (NB) medium (consist of beef extract for microbiology from MERCK, pepton from meat from MERCK, and distilled water)), Agar

Nutrient medium from MERCK, chloramphenicol, and 70% alcohol. The antibacterial examination method was Agar Diffusion Method. The procedures are described as follows.

## Endophytic Fungi Liquid Culture Preparation

The endophytic fungi liquid culture was prepare to obtain the secondary metabolites produced by each species of endophytic fungi (Guo *et al.*, 2008). Each endophytic fungi isolate was inoculated on PDA plate medium containing chloramphenicol (100mg/L) and then incubated at  $\pm 27^{\circ}\text{C}$  for 7x24 hours. Then each endophytic fungi culture were cut into five pieces 1x1 cm in size. Each endophytic fungi pieces were inoculated into 100 mL PDB medium and shaken at 120 rpm incubated in  $27^{\circ}\text{C}$  for 7x24 hours. Furthermore, the endophytic fungi liquid culture was filtered and centrifugated at the rate of 3000 rpm for 10 minutes. Each endophytic fungi species supernatant as much in 20  $\mu\text{L}$  volume was used for antibacterial examination in vitro (Siqueira *et al.*, 2011).

## The Antibacterial Effect Examination of Endophytic Fungi Liquid Culture

The antibacterial activity examination of each endophytic fungi species liquid cultures supernatant was done against *Escherichia coli*, *Bacillus subtilis*, and *Propioni bacterium acnes* used agar diffusion method. Each bacterial species was inoculated into NB medium, standardized by McFarland 0.5 ( $1.5 \times 10^8$  cfu/mL) and incubated in  $37^{\circ}\text{C}$  for 18 hours. The bacterial cultures were inoculated on NA plate medium and cut with a sterile cork borer to make wells in the medium then incubated in  $37^{\circ}\text{C}$  for 1x24 hours.

Furthermore, the wells were filled with 20 $\mu\text{L}$  of supernatant for each endophytic fungi species, then incubated at  $37^{\circ}\text{C}$  for 1x24 hours. The positive control used 5mg/mL chloramphenicol and negative control used PDB medium. The antimicrobial effect of each endophytic fungi species against *E. coli*, *B. subtilis*, and *P. acnes* was determined by measuring

the growth inhibition zone around the well on NA medium. The data were analyzed with Anova and continued with Duncan's test to determine which species of endophytic fungi have the highest antibacterial activity compared to another species of endophytic fungi based on the growth inhibition zone diameter.

### **Secondary Metabolite Compound Detection of Each Endophytic Fungi Species**

The flavonoid, alkaloid, tannin, saponin, and terpenoid compound in each endophytic fungi species liquid culture was analyzed by spectrophotometric methods.

### **Results and Discussion**

The antibacterial activity of endophytic fungi liquid cultures can be determined based on the results of measuring diameter of the bacteria growth inhibition zone around the wells that have been filled with endophytic fungi species liquid cultures on NA plate medium. The NA plate medium was inoculated with the bacteria cultures: *E. coli*, *B. subtilis*, and *P. acnes*. The results showed that each endophytic fungi liquid cultures isolated from the *Michelia champaca* plant have antibacterial activity against that bacteria species. The antibacterial activity of endophytic fungi *Rhizoctonia* sp. towards three bacteria species is shown in Fig 3.1.

The *Rhizoctonia* sp. liquid culture was the most effective against *E. coli*, the *Aspergillus parasiticus* liquid culture was the most effective against *B. subtilis*, while the *Curvularia lunata* liquid culture was the most effective against *P. acnes* based on the growth inhibition zone diameter. The clear zone around the well contained with endophytic fungi liquid culture which is containing secondary metabolites proved that there was inhibitory activity on the growth of the bacteria (Fig 3.1). This proves that endophytic fungi secondary metabolites in the wells diffuse into the medium and inhibit bacterial

growth. The results showed that each endophytic fungi species could produce the five antibacterial compounds, although the endophytic fungi species has been isolated from *Michelia champaca* plant as its host plant which could also produce similar secondary metabolites as produced by each endophytic fungi researched species.

The data were analyzed by Anova and followed by Duncan's test 5% to determine which endophytic fungi liquid cultures that have the highest antibacterial activity against *E.coli*, *B.subtilis*, and *P.acnes*. There was a difference in growth inhibition zone diameter between each endophytic fungi species towards the three bacteria species. The *Rhizoctonia* sp. liquid culture produced the largest growth inhibition zone diameter against *E.coli*, the *Aspergillus parasiticus* liquid culture produced the largest growth inhibition zone diameter against *B. subtilis*, and the *Curvularia lunata* liquid culture produced the largest growth inhibition zone diameter against *P. acnes*. The data are shown in Table 3.1.

Each endophytic fungi species liquid cultures isolated from *Michelia champaca* plant contained antibacterial compounds secondary metabolites. The antibacterial compounds that have been put into the well diffuse around it, so the bacteria that grow around the well are exposed by active compounds i.e: flavonoids, alkaloids, tannins and saponins, so it caused the death of the bacteria cells and formed clear zone called bacteria growth inhibition zone (Guo *et al.*, 2008; Yu *et al.*, 2010).

Alkaloids are one of the secondary metabolites active compounds which have antibacteria activity (Khameneh *et al.*, 2019). The antibacterial activity of alkaloid has been proven through various research results. Alkaloids are able to inhibit the formation of peptidoglycan in the bacteria cell wall so that the bacteria cell wall layer is not fully formed. The bacteria cell wall is a protector to the entire cell contents.

**Table.1** Duncan's Test of Antibacterial Activity of Each Liquid Culture of Endophytic Fungi against Test Bacteria of *E. coli*, *P. acnes*, and *B. subtilis*

Treatment	Test Bacteria	Average (mm)	Notation
<i>Colletotrichum alienum</i>	<i>E. coli</i>	0	a
<i>Colletotrichum alienum</i>	<i>P. acnes</i>	0	a
<i>Colletotrichum alienum</i>	<i>B. subtilis</i>	0	a
<i>Geotrichum candidum</i>	<i>E. coli</i>	0	a
<i>Aspergillus ochraceus</i>	<i>E. coli</i>	0	a
<i>Aspergillus ochraceus</i>	<i>P. acnes</i>	0	a
<i>Aspergillus ochraceus</i>	<i>B. subtilis</i>	0	a
<b>Mycelia sterilia 1</b>	<i>E. coli</i>	0	a
<b>Mycelia sterilia 1</b>	<i>P. acnes</i>	0	a
<b>Mycelia sterilia 1</b>	<i>B. subtilis</i>	0	a
<i>Papulospora</i> sp.	<i>E. coli</i>	0	a
<i>Papulospora</i> sp.	<i>P. acnes</i>	0	a
<i>Papulospora</i> sp.	<i>B. subtilis</i>	0	a
<b>Mycelia sterilia 2</b>	<i>E.coli</i>	0	a
<b>Mycelia sterilia 2</b>	<i>B. subtilis</i>	0	a
<b>Mycelia sterilia 3</b>	<i>P. acnes</i>	0	a
<b>Mycelia sterilia 3</b>	<i>B. subtilis</i>	0	a
<i>Curvularia lunata</i>	<i>E. coli</i>	0	a
<i>Colletotrichum kahawae</i>	<i>E. coli</i>	0	a
<i>Colletotrichum kahawae</i>	<i>P. acnes</i>	0	a
<i>Colletotrichum kahawae</i>	<i>B. subtilis</i>	0	a
<i>Aspergillus parasiticus</i>	<i>E. coli</i>	0	a
<i>Aspergillus parasiticus</i>	<i>P. acnes</i>	0	a
<b>K-</b>	<i>E. coli</i>	0	a
<b>K-</b>	<i>P. acnes</i>	0	a
<b>K-</b>	<i>B. subtilis</i>	0	a
<b>Mycelia sterilia 3</b>	<i>E. coli</i>	1,075	a
<b>Mycelia sterilia 2</b>	<i>P. acnes</i>	2,2	a
<i>Geotrichum candidum</i>	<i>P. acnes</i>	2,34	a
<i>Rhizoctonia</i> sp.	<i>B. subtilis</i>	2,525	a
<i>Rhizoctonia</i> sp.	<i>P. acnes</i>	2,875	a
<i>Curvularia lunata</i>	<i>P. acnes</i>	2,98	a
<i>Geotrichum candidum</i>	<i>B. subtilis</i>	3,205	a
<i>Curvularia lunata</i>	<i>B. subtilis</i>	4,125	a
<i>Aspergillus parasiticus</i>	<i>B. subtilis</i>	4,34	a
<i>Rhizoctonia</i> sp.	<i>E. coli</i>	14,44	b
<b>K+</b>	<i>B. subtilis</i>	18,45	b
<b>K+</b>	<i>E. coli</i>	18,76	b
<b>K+</b>	<i>P. acnes</i>	20,02	b

Explanation: K<sup>+</sup> = Chloramphenicol; K<sup>-</sup> = PDB medium

**Fig.1** Antibacterial activity of Endophytic Fungi *Rhizoctonia* sp against *E. coli*, *B. subtilis*, *P.acnes*.  
Description: A. *Rhizoctonia* sp. liquid culture against *E.coli*; B. *Rhizoctonia* sp. liquid culture against *B. subtilis*; C. *Rhizoctonia* sp. liquid culture against *P. acnes*.



The inhibition of cell wall formation could cause cell membranes susceptible to damage, so it can reduce cell wall semipermeability. Furthermore, the cell contents will come out from the cytoplasm and cause cellular metabolism inhibition. Furthermore, it caused inhibition in ATP formation, so the bacteria colony growth inhibited and followed by bacteria cell death (Sudrajat *et al.*, 2012).

Flavonoids are known as antibacterial agent against various pathogenic microorganisms (Xie *et al.*, 2015; Lamb and Cushnie, 2005). Flavonoid which have antibacteria activity can bind extracellular protein on the bacterial cell membrane. This can cause changes in the extracellular proteins structure. Furthermore, there will be a decrease in semipermeability of the cell membrane, thereby reducing cell metabolism. It can also cause inhibition of bacterial colony growth (Konate *et al.*, 2012). The OH<sup>-</sup> ion in the phenol component can bind to H<sup>+</sup> ion in hydrogen bonds on the structural protein cell wall and cause protein denaturation (Zainab, 2013). Furthermore, it causes damage to the bacteria cell wall and cell membrane.

Tannins are a group of polyphenol that is commonly found in nature, besides that tannin is secondary metabolites result that can take a role as antimicrobial substance (Abbas, 2014; Kumar and

Goel, 2019). Tannin can dissolve the fat layer on the bacterial cell wall and also denature structural protein on the bacteria cell membrane (Al-Ani *et al.*, 2008; Kaczmarek, 2020). This causes the cell wall structural damage and reduces the semipermeability of the cell membrane. Furthermore, it causes nutrient and enzymes come out of the cell, so that will cause cellular metabolism and colony growth inhibition.

Saponins are one of the secondary metabolites produced by endophytic fungi and have antimicrobial potential (Sadananda *et al.*, 2011; Jin *et al.*, 2017). Saponins can reduce surface tension, so it can increase in cell wall permeability. This causes intracellular compounds are released from the cytoplasm, so it could inhibit the cellular metabolism and cause bacteria cell death (Warganegara and Restina, 2016).

Terpenoid is an active chemical compound that can inhibit the growth of pathogenic bacteria (Abdel-Rahman *et al.*, 2019). Terpenoid can bind with purine, a transmembrane protein on the outer layer bacteria cell wall, to form a strong polymeric bonds that cause damage to purine. The cell wall damage causes the cell membrane will be susceptible to damage and then cause decrease in the cell membrane semipermeability. This fact causes the



nutrients in the cytoplasm come out, and it cause an obstacle in cellular metabolism. Furthermore, the ATP produced by cellular metabolism will be decreases, so it caused bacteria colony forming inhibition (Salni *et al.*, 2011; Sathya *et al.*, 2012).

The research results showed that the liquid cultures of 11 species of endophytic fungi isolated from *Michelia champaca* are able to produce secondary metabolites containing antibacteria compounds, those are flavonoid, alkaloid, tannin, saponin and terpenoid. The 11 endophytic fungi species liquid culture isolated from *Michelia champaca* plant could inhibit *E. coli*, *B. subtilis*, and *P. acnes* colony growth.

The research result can be used as the basic for using liquid cultures of these endophytic fungi as the base material for eco-friendly natural antibiotics. The use of this liquid cultures useful as an another alternative for picking activity on *Michelia champaca* plant parts in order to take as a natural medicine.

*Rhizoctonia* sp. is an endophytic fungi species isolated from *Michelia champaca* plant, which is able to produce antibacterial active compounds, with have the highest antibacterial effect against *E.coli* compared to 10 other endophytic fungi species. *Aspergillus parasiticus* was able to produce antibacterial active compounds with the highest antibacterial activity against *B. subtilis*. *Curvularia lunata* was able to produce active compounds, which is antibacterial, with the highest antibacterial effect against *P.acnes*, compared to 10 other species of endophytic fungi.

The conclusion of this research are:

Each endophytic fungi species isolated from *Michelia champaca* L. have antibacterial activity against *E. coli*, *B. subtilis*, *P. acnes*;

Secondary metabolite in *Rhizoctonia* sp. liquid culture has the highest antibacterial activity towards the three bacterial species. The *Curvularia lunata*

liquid culture is the most effective against *P.acnes*, the *Aspergillus parasiticus* liquid culture is the most effective against *B. subtilis*, and *Rhizoctonia* sp. liquid culture is the most effective against *E.coli*.

## References

- Abbas, S. R., Sabir, S. M., Ahmad, S. D. Boligon, A. A., & Athayde, M. L. 2014. Phenolic Profile, Antioxidant Potential and DNA Damage Protecting Activity of Sugarcane (*Saccharum officinarum*). *Food Chemistry*. Vol.147(1): 10–16.  
<https://doi.org/10.1016/j.foodchem.2013.09.113>.
- Abdel-Rahman, T., Hussein, A., Beshir, S., Hamed, A. R., Ali, E., & El-Tanany, S. S. 2019. Antimicrobial Activity of Terpenoids Extracted from *Annona muricata* Seeds and its Endophytic *Aspergillus niger* Strain SH3 Either Singly or in Combination. *Open Access Macedonian Journal of Medical Sciences*. Vol.1: 1-5.  
<https://doi.org/10.3889/oamjms.2019.793>.
- Al-Ani, R. T., Mohammed, N., Atheer, V. M., & Mohammed, S. 2008. Antibacterial Activity of Tannins Extracted from Some Medicinal Plants *in vitro*, *Iraqi Academy Scientific Journal*. Vol. 6(1):1-7.
- Ciocan, I. D & Bara, I. I. 2007. Plant Products As Antimicrobial Agents. *Anale Stiintifice ale Universitatii, Alexandru Ioan Cuza*. *Setiunea Geneticasi Biologie Moluculara*, TOM VIII.
- Guo, B., Y. Wang, X. Sun, & K. Tang, 2008. Bioactive natural products from endophytes: A review. *Applied Biochem.Microbiol*. Vol. 44: 136-142.  
<https://doi.org/10.1134/S0003683808020026>
- Jin, Z., Gao, L., Zhang, L., Liu, T., Yu, F., Zhang, Z., Guo, Q., & Wang, B. 2017. Antimicrobial Activity of Saponins Produced by Two Novel Endophytic Fungi from *Panaxnotoginseng*. *Natural Product Research*. Vol.1: 1-4.  
<https://doi.org/10.1080/14786419.2017.1292265>.

- Kaczmarek, B. 2020. Tannic Acid with Antiviral and Antibacterial Activities as a Promising Component of Biomaterials—A Minireview. *MDPI Journal-Materials*. Vol.13: 1-13.  
<https://doi.org/10.3390/ma13143224>.
- Khameneh, B., Iranshahy, M., Soheili, V., & Bassas, B. S. F. 2019. Review on Plant Antimicrobials: A Mechanistic Viewpoint. *Antimicrobial Resistance and Infection Control*. Vol.8(118): 1-28.  
<https://doi.org/10.1186/s13756-019-0559-6>.
- Konate, K., Hilow, A., Mavoungou, J. F., Lepengue, A. N., Souza, A., Barro, N., Datte, J. Y., Batchi, B. M., & Nacoulma, O. G. 2012. Antimicrobial Activity of Polyphenol Rich Fractions from *Sida alba* L. (Malvaceae) Against Cotrixazol-Resistant Bacteria Strains. *Annals Of Clinical Microbiology and Antimicrobials*, Vol.11(2): 1-6.
- Kumar, N. & Goel, N. 2019. Phenolic Acids: Natural Versatile Molecules with Promising Therapeutic Applications. *Biotechnology Reports*. Vol.24: 1-10.  
<https://doi.org/10.1016/j.btre.2019.e00370>.
- Lamb, A. J. & Cushnie, T. P. T. 2005. Antimicrobial Activity of Flavonoids. *International Journal of Antimicrobial Agents*. Vol.26(1): 343–356.  
<https://doi.org/10.1016/j.ijantimicag.2005.09.002>.
- Mousa, W. K. & Raizada, N. 2013. The Diversity of Anti-microbial Secondary Metabolites Produced by Fungal Endophytes: An Interdisciplinary Perspective. *Frontiers in Microbiology*. Vol.4(65): 1-18.  
<https://doi.org/10.3389/fmicb.2013.00065>.
- Mwanga, Z. N., Mvungi, E. F., & Tibuhwa, D. D. 2019. Antimicrobial Activities of Endophytic Fungi Secondary Metabolites from *Moringa oleifera* (Lam). *Tanzania Journal of Science*. Vol.45(3): 463-476.
- Nair, D. N. & Padmavathy, S. 2014. Impact of Endophytic Microorganisms on Plants, Environment and Humans. *Scientific World Journal*. Vol 2014:1-11.  
<https://doi.org/10.1155/2014/250693>
- Sadananda, T. S., Nirupama, R., Chaithra, K., Govindappa, M., Chandrappa C. P., & Vinay, R. B. 2011. Antimicrobial and Antioxidant Activities of Endophytes from *Tabebuia argentea* and Identification of Anticancer Agent (Lapachol). *Journal of Medicinal Plants Research*. Vol. 5(16): 3643-3652.  
<https://doi.org/10.5897/JMPR.9000170>
- Salni., Marisa, H & Mukti, R. W. 2011. Isolasi Senyawa Antibakteridari Daun Jengkol (*Pithecolobium lobatum* Benth) dan Penentuan Nilai KHM-nya. *Jurnal Penelitian Sains*. Vol.14(1).  
<https://doi.org/10.36706/jps.v14i1.125>
- Sathya, B. S., Jayasurya, K. S. Sankaranarayanan, S., & Bama, P. 2012. Antibacterial Activity of Different Phytochemical Extracts from The Leaves of *T. Procumbens* Linn.: Identification and Mode of Action of The Terpenoid Compound As Antibacterial. *International Journal of Pharmacy and Pharmaceutical Sciences*. Vol.4(1): 557-564.
- Siqueira, V. M., Conti, R., Araujo, J. M., & Motta, C. M. S. 2011. Endophytic Fungi from the Medicinal Plant *Lippiasidoides* Cham. and their Antimicrobial Activity. *Symbiosis* 53: 89-95. <https://doi.org/10.1007/s13199-011-0113-7>
- Sudrajat, S., Sadani, S., & Sudiastuti, S. 2012. Analisis Fitokimia Senyawa Metabolit Sekunder Ekstrak Kasar Etanol Daun Meranti Merah (*Shorea leprosula* Miq.) dan Sofat Antibakterinyaterhadap *Staphylococcus aureus* dan *Eschericia coli*. *Journal of Tropical Pharmacy and Chemistry*, Vol. 1 (4):303-311.  
<https://doi.org/10.25026/jtpc.v1ii4.41>
- Warganegara, E., & Restina, D. 2016. Getah Jarak (*Jatropha curcas* L.) sebagai Penghambat Pertumbuhan Bakteri *Streptococcus mutans* pada Karies Gigi. *Majority*. Vol.5(3): 62.
- Xie, Y., Yang, W., Tang, F., Chen, X., & Ren, L. 2015. Antibacterial Activities of Flavonoids: Structure-Activity Relationship and

- Mechanism. *Current Medicinal Chemistry*. Vol.22(1): 132-149.  
<https://doi.org/10.2174/0929867321666140916113443>.
- Yu, H., L. Zhang, L. Li, C. Zheng, & Guo, L.. 2010. Recent Developments and Future Prospects Of Antimicrobial Metabolites Produced By Endophytes. *Microbiol. Res.*, Vol.165: 437-449.  
<https://doi.org/10.1016/j.micres.2009.11.009>
- Zainab. 2013. The Ethanol Concentration As Extraction Solvent Effect towards *Lawsonia inermis* L. leaf extract. *J. Farmasiana*. Vol.3(2): 63-68.

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