


Original Research Article

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A Study on Antimicrobial Susceptibility Pattern of Bacterial Infections in Burn Wound Patients in a Tertiary Care Hospital, Salem

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ABSTRACT

Skin being the largest organ of our human body and Out of numerable immune preclusion mechanisms in our body skin is one of the momentous defence. For those patients who survive initial burn injury infection is an imperative issue. Burn wound injuries are one of the most common, invasive and devastating forms of trauma. Despite the recent advances in burn wound management, bacterial infections persist as an important complication and leading cause of morbidity and mortality among burnt patients. The leading issue in burns wound management is culpability of the burn wound to colonisation of bacteria. Disparity of bacterial flora isolated is appreciated during the period of wound healing. This descriptive study was performed in the department of Microbiology, in conjunction with the burns wards in a Tertiary care hospital. This study was conducted from 2022 October to 2022 December. Organisms are identified based on the morphology of the colonies, Gram stain appearance, testing for presence of catalase enzyme, testing for presence of oxidase enzyme, testing of motility by hanging drop method. Antibiotic susceptibility test was done using modified Kirby Bauer's disc diffusion method on Mueller Hinton Agar as per Clinical and Laboratory Standards Institute Guidelines. Out of 100 patients, majority of them belongs to 21-40 years and female patients were 61%. Out of 100 samples processed, 67 sample showed bacterial growth and 43 sample showed no growth. Among the 67 sample with growth, gram negative isolates (68.6%) were more compared to gram positive isolates(31.3%). *Klebsiella* species (26.8%) were the most predominant organism isolated followed by *Pseudomonas aeruginosa* (16.4%), *Proteus* species (11.9%), *Acinetobacter baumannii* (8.9%), *Escherichia coli* (4.4%) among gram negative organisms. In the gram positive isolates, *Staphylococcus aureus* were (16.4%), followed by Coagulase negative *Staphylococcus* (10.4%), *Enterococcus* species (4.4%). *Klebsiella* is the most common isolate among all followed by *Pseudomonas*, *Staphylococcus aureus*. Piperacillin and tazobactam, carbapenem has increased sensitivity when compared to other drugs for Gram negative pathogens, where as Cotrimoxazole, Amoxicillin Clavulanic acid, Ciprofloxacin all have very high resistance.

Keywords

Burn wounds, trauma, colonization, infection, sepsis-related

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Introduction

Burn wound injuries are one of the most common, invasive and devastating forms of trauma. Despite the recent advances in burn wound management, bacterial infections persist as an important complication and leading cause of morbidity and mortality among burnt patients.

Nearly 73% of post-burn deaths occurring within five days have been reported to be sepsis-related. Burn wounds are highly susceptible to colonization and infection and this is the major problem in the management of burn victims. Initially, the burnt area is considered free of microbial contamination (Liwimbi and Komolafe, 2007; Fatima Kabanangi *et al.*, 2021).

Major burn wounds usually become infected within 3-5 days after admission, so it is obvious that the infection arises from the patient's own bacterial flora and is not an exogenous occurrence.

Predominant risk factors for burn wound infection are the size of burn wound, i.e., the percentage of total body surface area (TBSA) burnt and the duration of hospitalization (Hisham A. Abbas *et al.*, 2013; Vishwajith *et al.*, 2021).

The main aim and objectives of this study to determine the microbial profile of burn wound infection and also to determine the antimicrobial susceptibility patterns of the isolated organisms

Materials and Methods

Surface swab collected from patients with Burn wound infections admitted in Burns ward.

Sample collection

To obtain a culture of burn surface, topical agents were first removed with a gauze soaked in sterile saline. The method of collection was deep swabbing, or aspiration of the bleb. Then the sample was collected by two sterile swab sticks. For dry wounds

the swab was moistened with sterile saline. After the collection, the swab were immediately transported to the laboratory for further processing.

Sample processing

One of the swab sticks was used for direct gram staining. By the other swab stick, the sample was inoculated in Nutrient agar, Blood agar, MacConkey plates and incubated for 18 to 24 hours at 37°C.

At the end of the incubation period, the plates were examined for the growth and the preliminary identification was made based on colony morphology, gram staining, motility and the series of biochemical tests (catalase, coagulase and oxidase tests).

Anti-microbial susceptibility testing

Antimicrobial susceptibility test was done for all bacterial isolates as per CLSI guidelines by Kirby Disk Diffusion method on Muller Hinton agar.

Implication

The use of prophylactic antibiotics is common practice with burnt patients. Drug resistant bacteria with intrinsic resistance towards antibiotics, the ability to survive longer in the hospital environment and hand-to-hand transmission of bacteria reflect their easy spread and the possible causes of outbreak. So continuous surveillance and update of antibiotic resistance pattern of microorganisms is imperative for infection control programs and accurate antibiotic treatment in the burnt patients.

Thus this will help to determine the predominant bacterial agents causing burn wound infection in our healthcare setup and their antimicrobial profile will help the policy makers in formulation of strategies for rational and effective use of antimicrobial agents which in turn lead to control of spread of antibiotic resistance genes in the community and in the reduction of morbidity and mortality associated with better management of burn wound patients.

Results and Discussion

Age and sex wise distribution

Out of 100 patients, majority of them belongs to 21-40 years and female patients were 61%.

Distribution of isolates

Out of 100 samples processed, 67 sample showed bacterial growth and 43 sample showed no growth. Among the 67 sample with growth, gram negative isolates (68.6%) were more compared to gram positive isolates (31.3%). *Klebsiella* species (26.8%) were the most predominant organism isolated followed by *Pseudomonas aeruginosa* (16.4%), *Proteus* species (11.9%), *Acinetobacter baumannii* (8.9%), *Escherichia coli* (4.4%) among gram negative organisms. In the gram positive isolates, *Staphylococcus aureus* were (16.4%), followed by Coagulase negative *Staphylococcus* (10.4%), *Enterococcus* species (4.4%) (Table 3).

The gram negative isolates were highly sensitive to Imipenem (100%) followed by Piperacillin-Tazobactam (84%), Cefoperazone-Sulbactam (74%), Amikacin, Gentamicin. They showed least sensitivity to Amoxicillin-Clavulanic acid (36%).

The gram positive isolates showed high sensitivity to Linezolid & Vancomycin (100%) followed by Doxycycline, Erythromycin, Clindamycin. They showed high resistance to Ampicillin.

Burn injuries are a global public health problem and still remain the leading cause of disability and unintentional death (WHO, 2018; Roman *et al.*, 2012).

In developing nations 17% of permanent and 18% of temporary disabilities in children exposed to burns. Even in those who survive results in a convincing

amount of morbidity, sustained hospitalization, impairment (Burns, 2018).

Overall following changes in immune system happens in patients with major burns 1. fall in number of helper T cells, 2. rise in number of suppressor T cells, 3 reduced production of monocyte and macrophage, and 4. Fall of immunoglobulin levels (Lawrence C. Madoff and Florencia Pereyra, 2012) Increased cytokine levels contribute to the dysregulation of immune system.

Initial colonization taking place at the site of burns which then proceed to invade if unchecked leading to bacteremia and sepsis, ending up with increased mortality (Pruitt and McManus, 1984).

The present study is carried out to determine the bacteriological profile and antimicrobial susceptibility patterns among the patients admitted in the burns ward at a tertiary care hospital in Tamil Nadu, swab was collected from these patients and inoculated in appropriate culture media, the isolates are identified based on standard procedures employing methods like Gram staining, standard biochemical reactions etc. the identified isolates sensitivity to antimicrobial agents are obtained by Kirby bauer disc diffusion method according to CLSI guidelines 2021.

Out of 100 patients, majority of them belongs to 21-34 years (34%). Similar to study conducted by Kavitha *et al.*, (2018). Out of the 100 patients 39 percentage are male and 61 percentage are female.

Contrast to our study by study showing increased incidence in male are Agnihotri *et al.*, (2004). Caroline Mohr O'Hara *et al.*, 2000; Proteus *et al.*, 2000) and Kaur *et al.*, (2006). In Kavitha *et al.*, (2018) study conducted in Telangana showed percentage of female is 58% which higher than male (Church *et al.*, 2006).

Table.1 Age wise distribution

Age group (years)	No. of patients
13-20	5
21-30	34
31-40	25
41-50	14
51-60	11
>60	10
Total	100

Table.2 Sex wise distribution

Sex	No. of patients
Female	61
Male	39
Total	100

Table.3 Distribution of isolates

Organism (n=67)	No. of isolates (%)
Gram negative isolates (n=46)	
<i>Klebsiella</i> species	18(26.8%)
<i>Pseudomonas aeruginosa</i>	11(16.4%)
<i>Proteus</i> species	8(11.9%)
<i>Acinetobacter baumannii</i>	6(8.9%)
<i>Escherichia coli</i>	3(4.4%)
Gram Positive isolates (n=21)	
<i>Staphylococcus aureus</i>	11(16.4%)
Coagulase negative <i>Staphylococcus</i>	7(10.4%)
<i>Enterococcus</i> species	3(4.4%)
Total	67 (100%)

Table.4 Antimicrobial sensitivity pattern of gram negative bacterial isolates

Organism / Drugs	<i>Klebsiella</i> spp	<i>Pseudomonas</i> spp	<i>Proteus</i> spp	<i>Acinetobacter</i> spp	<i>Escherichia coli</i>
AMC	38%	-	37%	-	33%
CIP	69%	36%	50%	67%	33%
COT	39%	-	37%	50%	67%
GEN	80%	63%	75%	50%	33%
AK	76%	72%	75%	67%	67%
CTX	57%	-	62%	-	67%
CFS	80%	81%	75%	67%	67%
PTZ	92%	90%	87%	83%	67%
IPM	100%	100%	100%	100%	100%
CAZ	-	54%	-	83%	-
CPM	-	72%	-	-	-

Table.5 Antimicrobial sensitivity pattern of gram positive bacterial isolates

Organism / Drugs	<i>Staphylococcus aureus</i>	CoNS	<i>Enterococcus spp</i>
AMP	37%	42%	67%
COT	63%	57%	-
DOXY	71%	57%	100%
ERYTHRO	54%	85%	67%
CD	54%	85%	67%
LZ	100%	100%	100%
VA	100%	100%	100%
CEFOXITIN	81%	57%	-

Fig.1 Age wise distribution

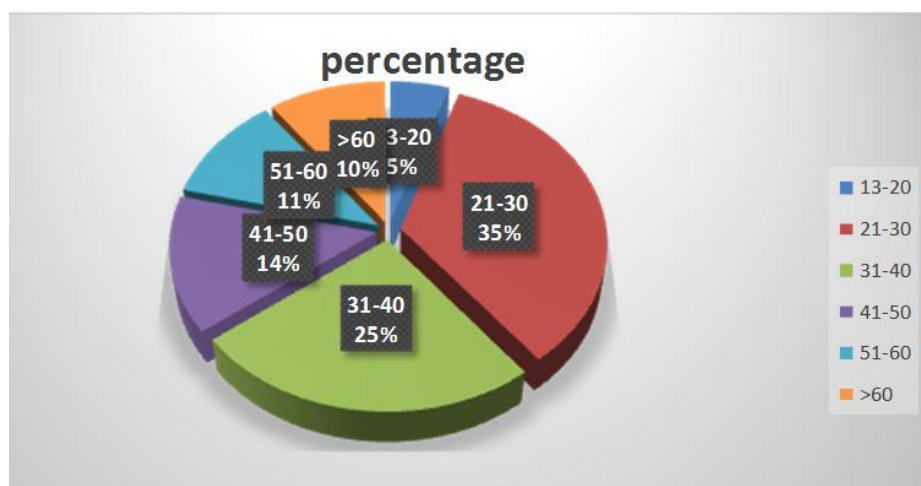


Fig.2 Sex wise distribution

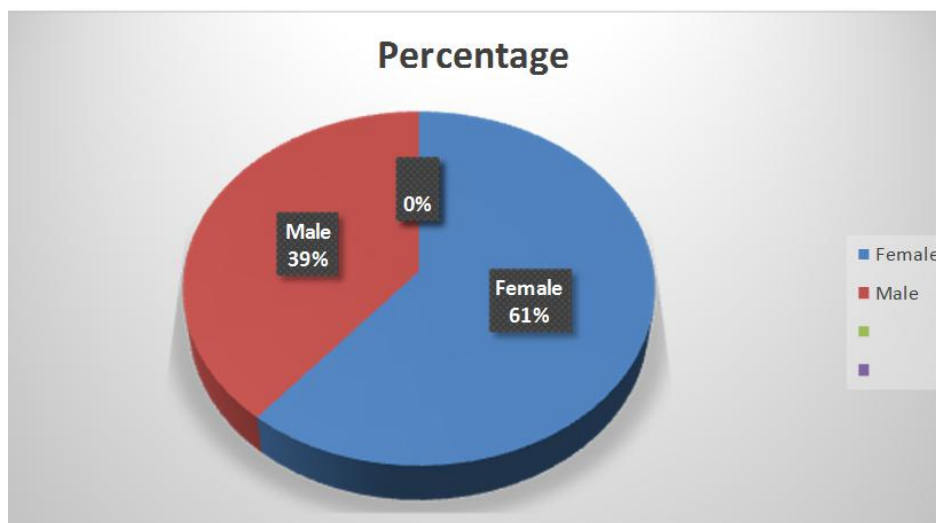
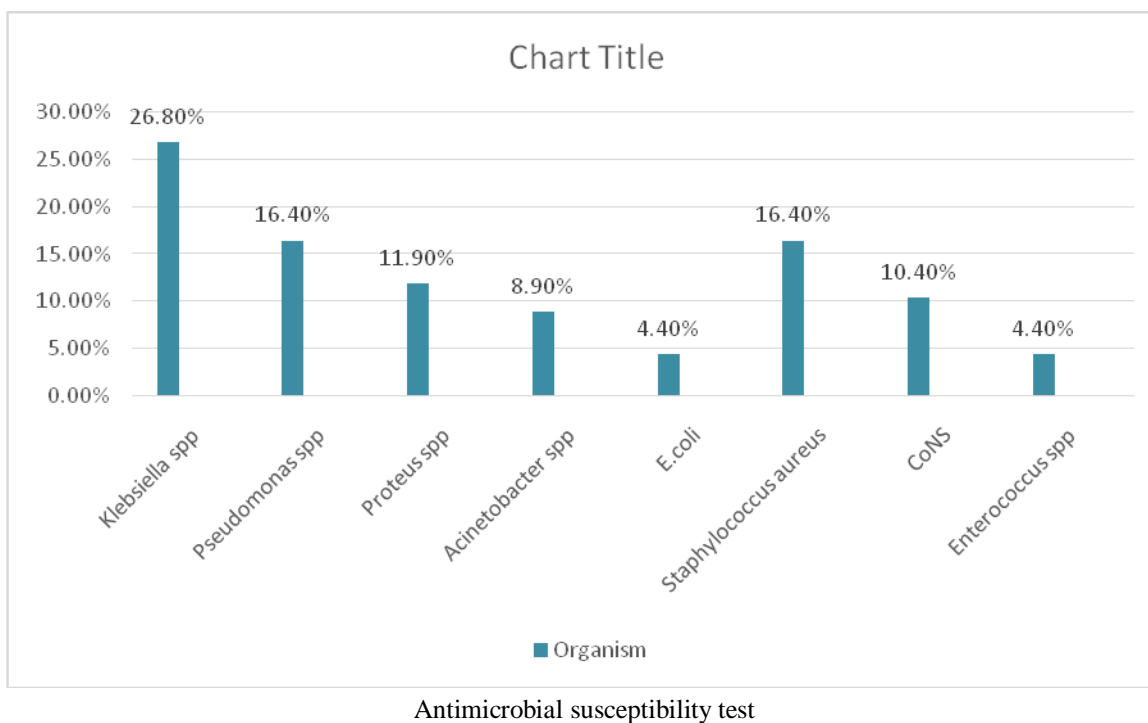


Fig.3 Distribution of isolates



Gram negative bacteria were the predominating bacteria with 46 isolates and 68.6% and Gram positive isolates were only 21 (31.3%). This pattern is similar to the observation in Saaq *et al.*, (2015).

Whereas study conducted by, Lakshmi *et al.*, (2015); Kavitha *et al.*, (2018); Alghalibi *et al.*, (2011) and Naveen Saxena *et al.*, (2013) had Grampositive bacteria predominantly.

Klebsiella spp in our study showed 92% of the isolates are sensitive to piperacillin – Tazobactam and 8% are resistant to it. 100% of the isolates are sensitive to Imipenem and 10% are resistant to it. similar to study by contrast to results obtained by Jyoti sonawane (2010) where 96.3% sensitivity to imipenem obtained.

In *Pseudomonas aeruginosa* 100 % of the isolates are sensitive to imipenem and no resistant to it. Contrast to our study by study showing 76% sensitivity shown by Prabat Ranjan *et al.*, (2010). Ciprofloxacin in our study sensitive to 37% similar to study by Prabat Ranjan *et al.*, (2010).

Proteus spp in our study showed sensitive to both Amoxyclav and cotrimoxazole showed 37% and resistance were 63%. This goes well in line with Desai *et al.*, (2011). 87 % of the isolates are sensitive to Piperacillin- Tazobactam and. 75% of the iolates are sensitive to Cefoperazone – Sulbactam, gentamycin and Amikacin. In contrast to work done by Madhavi and Shaziaparveen (2015) in which 100% resistance to Cotrimoxazole and 100% sensitivity to Amikacin mentioned.

Escherichia coli in our study showed sensitivity about 67% of the isolates are sensitive to Cefoperazone – sulbactam, 33% to Gentamycin and 20% a. 67% of the isolates are sensitive to Ceftriaxone and contrast to our study by Mishra *et al.*, (2000).

Staphylococcus aureus in our study showed 100% of the isolates are sensitive to vancomycin, Linezolid, and 71 % to Doxycyclin. This is similar to study done by Fantahunbiadglegne *et al.*, (2009). *Klebsiella* is the most common isolate among all followed by *Pseudomonas Staphylococcus aureus*.

Piperacillin and tazobactam, carbapenem has increased sensitivity when compared to other drugs for Gram negative pathogens, where as Cotrimoxazole, Amoxicillin Clavulanic acid, Ciprofloxacin all have very high resistance

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So, continuous surveillance and update of antibiotic resistance pattern of microorganisms is imperative for infection control programs and accurate antibiotic treatment in the burnt patients.

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References

- Agnihotri N, Gupta V, Joshi R M. Aerobic bacterial isolates from burn wound infections and their antibiograms – a five year study. *Burns* 2004; 30(3): 241-243
<https://doi.org/10.1016/j.burns.2003.11.010>.
- Alghalibi, S. M. S.; Humaid, A. A.; Alshaibani, E. A. S. and Alhamzy, E. H. L. Microorganisms Associated With Burn Wound Infection; *Egypt. Acad. J. biolog. Sci.*, 2011; 3(1): 19-25
<https://doi.org/10.21608/EAJBSG.2011.16691>
- Burns. (2018). Accessed: May 27, 2019: <http://www.who.int/news-room/factsheets/detail/burns>
- Caroline Mohr O'Hara, Frances W. Brenner and J. Michael Miller. Classification, identification, and clinical significance of *Proteus*, *Providencia*, and *Morganella* clinical microbiology reviews, oct. 2000, p. 534–546 vol. 13, no 4
<https://doi.org/10.1128/cmr.13.4.534-546.2000>
- Church D, Elsayed S, Reid O, Winston B, lindsay R, Burn wound infections. *Clin Microbiology Review*, 2006 Apr; 19(2): 403-34. doi: <https://doi.org/10.1128/CMR.19.2.403-434.2006>.
- Desai, K. J., S. S. Malek, A. Parek Neonatal Septicemia Bacterial isolates and their antibiotic susceptibility pattern *Gujarat Medical Journal* Feb 2011 Vol 66 No.1.
- Fantahun Biadlegne, Bayeh Abera, Atenaf Alam, Belay Anagaw. Bacterial isolates from wound infections, *Ethiopia Journal of Health sciences*, 2009,19,3,173-177.
- Fatima Kabanangi, Agricola Joachim, Emmanuel James Nkuwi, Joel Manyahi, Sabrina Moyo, Mtebe Majigo. 2021. High Level of Multidrug-Resistant Gram-Negative Pathogens Causing Burn Wound Infections in Hospitalized Children in Dar es Salaam, Tanzania, *International Journal of Microbiology*, 8 pages.
<https://doi.org/10.1155/2021/6644185>
- Hisham A. Abbas, Eman M. El-Masry, Ghada H. Shaker, Islam Mohsen. 2013. Bacterial etiology and antimicrobial resistance of burn wound infections in a burn unit in hehia general hospital in Egypt, *International Journal of Biological & Pharmaceutical Research*. 4(12): 1251-1255.
- Jyoti Sonawane, Narayan Kamath, Rita Swaminathan, kausal Dhosani. Bacterial profile of SSI and their antibiograms in a tertiary care hospital in Navi Mumbai., *Bombay hospital Journal* 2010, 52, 3,
- Prabhat Ranjan, K, Neelamaranjan, Sathish K Bansal, D R Arora. 2010 Prevalence of *pseudomonas aeruginosa* in post operative wound infection in a referral hospital

- Harayana, India Journal of Laborator <https://doi.org/10.4103/0974-2727.72153>.
- Kaur H, Bhat J, Anvikar A R, Rao S, Gadge V. Bacterial profile of blood and burn wound infections in burn patients. In: Proceedings of National Symposium on Tribal Health; 2006 Oct 19-20; Jabalpur, India: p. 89-95
- Kavitha, M. L., S L. Annapoorna and Nagaprasad (2018). Bacteriological Profile of Burns Wound Isolated from a Teaching Hospital in Telangana, India. *Int.J.Curr.Microbiol.App.Sci.* 7(12): 3195-3202. <https://doi.org/10.20546/ijcmas.2018.712.369>
- Lakshmi, N, R Koripella, J Manem, P B Krishna - ... of Dental and Medical Sciences, 2015 Shehzad Ahmad, and Muhammad Salman Zaib
- Lawrence C. Madoff, Florencia Pereyra. (2012). Infectious Complications of Burns. In: Harrison's E-Medicine. The McGraw Hill Companies, Inc;e23:1-4
- Liwimbi O M, Komolafe I O. 2007. Epidemiology and bacterial colonization of burn injuries in Blantyre. *Malawi Med J.* Mar;19(1):25-7.
- Madhavi S.* and S. Shaziaparveen Bacteriological profile and Antibiotic sensitivity of wound infections *International Journal Of Current Microbiology and Applied Sciences* (2015)4(12):248-254
- Misra R N, Yogesh Chander, N K Debata, V C Ohri. (2000) Antibiotic resistance pattern of isolates from wound and soft tissue infections, *MJAFI*, 56(3):205-208. [https://doi.org/10.1016/S0377-1237\(17\)30167-3](https://doi.org/10.1016/S0377-1237(17)30167-3)
- Muhammad Saaiq, Burn Wound Infections and Antibiotic Susceptibility Patterns at Pakistan Institute of Medical Sciences, Islamabad, Pakistan (2015). Bacteriological profile and antibiogram of burn wound infections in a tertiary care hospital
- Naveen Saxena, Divya Dadhich, Deepak Maheshwari. "Aerobic Bacterial isolates from burn wound infection patients and their antimicrobial susceptibility pattern in Kota, Rajasthan." *Journal of Evolution of Medical and Dental Sciences* 2013; 2(23): 4156-60 <https://doi.org/10.14260/jemds/815>
- Proteus, Morganella, and Providentia Bernard W. Senior, Topley and Wilson's Microbiology and Microbial Infection. 10th ed. Edward Arnold Ltd.Italy.2005. Volume-2, part-3 Chapter -31 page. 1-14 O'Hara C M, Brenner F W, and Miller J M. Classification, identification and 125 clinical significance of Proteus, Providencia, and Morganella. *Clin Microbiol Review* 2000;13:534-546
- Pruitt B A, Jr, McManus A T. (1984). Opportunistic infections in severely burned patients. *Am. J. Med*;76:146-154. [https://doi.org/10.1016/0002-9343\(84\)90334-6](https://doi.org/10.1016/0002-9343(84)90334-6).
- Roman, I. M., E. R. Lewis, H. A. Kigwangalla, and M. L. Wilson, "Child burn injury in Dar es Salaam, Tanzania: results from a community survey," *International Journal of Injury Control and Safety Promotion*, vol. 19, no. 2, pp. 135– 139, 2012.
- Vishwajith, K. Anuradha and Venkatesha, D. 2021. Bacteriological Profile and Antimicrobial Resistance Patterns of Burn Wound Infections in a Tertiary Care Hospital. *Int.J.Curr.Microbiol.App.Sci.* 10(02): 2769-2775. <https://doi.org/10.20546/ijcmas.2021.1002.306>
- World Health Organisation, Burns; Fact Sheet, WHO, Geneva, Switzerland, 2018. [2]

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